From Farm to Fork: The Zoonotic Transmission of Salmonella spp. and the Rise of Antimicrobial Resistance

Shameeran Salman Ismael^{1,*}, Bland Husamuldeen Abdullah¹ and Barhav Issa Abdullah¹

¹Department of Medical Laboratory Sciences, College of Health Sciences, University of Duhok, Duhok, Iraq *Corresponding author: <u>shameeran.Ismael@uod.ac</u>

Abstract

Salmonella is a zoonotic foodborne bacterium that causes gastroenteritis in both humans and animals. With more than 2000 serovars known to exist, salmonella is a very dangerous bacterium. The serovars that produce typhoid fever are known as non-typhoidal *Salmonella*, and they are responsible for both gastroenteritis and the illness. Eating foods from animals is often linked to Salmonella infection, which humans can get through the farm-to-fork chain. The next most significant sources, after poultry products, are fish, cattle, pork products, vegetables, and fruit. The rise of antimicrobial resistance and the introduction of resistant strains of Salmonella to multiple drugs have made identifying antibiotic alternatives more urgent, even though antibiotics are still the main treatment for salmonellosis. Many virulence factors are required to adhere to, invade, and evade the host's defense system. Salmonella's pathogenicity, including its adhesion proteins, flagella, plasmids, type 3 secretion systems, and capsule. According to the recent rise in the presence of NTS variants worldwide, control methods meant to reduce food animal contaminants throughout the food chain might not have been successful. The emergence of antibiotic-resistant Salmonella bacteria also raises the possibility of a food safety emergency. Thus, the spread of Salmonellosis and the emergence of outbreaks of antibiotic resistance were the main topics of this chapter.

Keywords: Salmonellosis, Zoonosis, Antimicrobial resistance, Farm, Food

Cite this Article as: Ismael SS, Abdullah BH and Abdullah BI, 2025. From farm to fork: the zoonotic transmission of salmonella spp. and the rise of antimicrobial resistance. In: Zaman MA, Farooqi SH and Khan AMA (eds), Holistic Health and Antimicrobial Resistance: A Zoonotic Perspective. Unique Scientific Publishers, Faisalabad, Pakistan, pp: 147-154. <u>https://doi.org/10.47278/book.HH/2025.284</u>



A Publication of Unique Scientific Publishers Chapter No: 25-308

Received: 18-Feb-2025 Revised: 14-Apr-2025 Accepted: 01-May-2025

Introduction

Salmonellosis is the most prevalent zoonotic foodborne bacteria and a global public health concern. With more than 2000 serov ars, *Salmonella enterica* is the most virulent species. It causes gastroenteritis and typhoid fever, and the serovars that cause the latter are called non-typhoidal Salmonella (Somorin et al., 2021). *Salmonella* is under the family Enterobacteriaceae. It can live in mammals, amphibians, and reptiles and causes gastroenteritis in humans (Sánchez-Vargas et al., 2011; CDC, 2023). Through the farm-to-fork chain, Salmonella gets spread to people and is frequently associated with eating foods derived from animals. Chicken and chicken products are the main contributors among these sources, followed by red meat, pig meat, fish meat, vegetables, and fruits (Lamichhane et al., 2024). *Salmonella* is listed by the World Health Organization as one of the top four global causes of diarrhea (Hoffmann et al., 2012, Chlebicz & Śliżewska, 2018).

The bacterial serotype and the host's immune system determine the severity of the two types of human infections: typhoidal and nontyphoidal (Wei et al., 2023). Fever, cramping in the abdomen, and diarrhea with an abrupt onset are common symptoms of non-typhoidal Salmonella infections (Keestra-Gounder et al., 2015). It typically resolves on its own in one to seven days without therapy, depending on the host (Hoelzer et al., 2011). However, 5% of individuals, particularly those with weakened immune systems, newborns, and the elderly, may get bacterial infections or invasive infections such as septic arthritis, meningitis and osteomyelitis, and endovascular infections (Olnood et al., 2015, Wibisono et al., 2020).

Clinical problems in highly vulnerable people are typically treated with broad-spectrum antibiotics (Galán, 2021). The earliest antibiotics used to treat salmonellosis were trimethoprim/sulfamethoxazole and chloramphenicol (Stoycheva & Murdjeva, 2006). The third generation of quinolones, such as fluoroquinolones, ofloxacin, and ciprofloxacin, is currently the medication for salmonellosis in individuals with weakened immune systems (Jibril et al., 2021). Ceftriaxone and azithromycin are used empirically to treat salmonellosis because of the growing bacterial resistance to fluoroquinolones (Hailu et al., 2021).

Vaccines, like medicine, were used for the prevention of salmonellosis in humans and animals (van Panhuis et al., 2013). The Food and Drug Administration has approved two vaccines to prevent Salmonella: the oral live attenuated Ty21a vaccine and the intra muscular Vi polysaccharide capsular vaccine. New attenuated vaccines, glycoconjugate, O-antigen glycoconjugate, and GMMA-based vaccines are still being developed (Sears et al., 2021). The effectiveness of immunizations against Salmonella is limited by a number of reasons, including complex immune evasion mechanisms, a range of serotypes, and the presence of asymptomatic carriers, making vaccine design difficult (Giannelli et al., 2017). This chapter aimed to determine the transmission of Salmonella spp. and the rise of antimicrobial resistance outbreaks Transmission of Salmonellosis

Since chicken is believed to be the primary cause of *Salmonella* infection in humans, poultry as well as poultry goods are the initial point of infection (Gonzalez-Escobedo & Gunn, 2013). Incorrect handling of contaminated organs, such as the stomach and liver, during carcass processing is the most frequent cause of contaminated meat (*Dos Santos et al., 2020*). The second source is ground meat, 82.2% of Americans eat beef every week, with 67% specifically stating that they prefer ground beef meat (CDC, 2023). Worldwide, domesticated animals and people are mostly exposed to *Salmonella* through wild animals, such as feral pigs and wild boar (Cilia et al., 2021). Humans are usually infected by eating contaminated meat from poultry as well as wild animals, or by coming into close contact with contaminated animal excrement (Gil Molino et al., 2019).

Almost all of the Salmonella-related infections that occur globally each year are foodborne; the infection can be transferred by either indirect or direct contact within the household, at the hospital, or on a farm (Majowicz et al., 2010). *Salmonella* can spread by direct contact, such as when feces-contaminated food or drink is consumed (Ford et al., 2023). Vertical transmission typically occurs in birds and reptiles when pollen via the female reproductive system can reach the eggs (*Griffith et al., 2019*). By employing intermediary objects such as infected tools and live or dead vectors, the germs can spread indirectly (Dróżdż et al., 2021). Figure 1 highlights the transmission of salmonellosis.

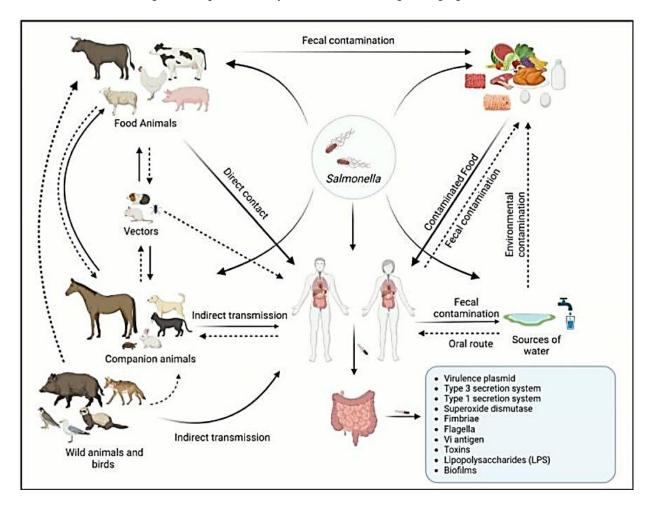


Fig. 1: Routes of transmission of salmonellosis in humans and animals (Lamichhane et al., 2024) https://pmc.ncbi.nlm.nih.gov/articles/PMC10812683/

Clinical Signs of Salmonellosis

There are two common types of salmonellosis typhoidal and non-typhoidal

Typhoidal or Enteric Fever

Commonly referred to as typhoid or paratyphoid, it is caused by typhoidal *Salmonella* serovars, such as *Salmonella typhi* or *Salmonella paratyphi* (Reddy et al., 2010) A week or more is the incubation period for enteric fever, during which time people may found with several symptoms, including a high fever, headache, vomiting, and diarrhea A noticeable temperature pattern appears during enteric fever (Crump, 2019). In the first week, it starts as a low-grade fever ranging between 37.5°C to 38°C, and then progressively increases to a high-grade fever (41.5°C) (Thakur et al., 2022). If proper treatment is not received, the fever may last up to one month (Patel et al., 2010). Infected people may have bradycardia, myalgia, splenomegaly, hepatomegaly, and the appearance of red on the abdomen and chest in addition to fever (Kuvandik et al., 2009). Hepatitis, pancreatitis, and cholecystitis are among the common gastrointestinal problems that affect about 15% of infected

individuals with salmonellosis in endemic areas (Galán, 2016). Bloody diarrhea, or eruption of Peyer's patches, is lymphatic nodules in the last part of the ileum that can result in hemorrhage, one of the most dangerous gastrointestinal adverse effects (Crump, 2019).

Non Typhoidal Salmonellosis

The most common causative agents for this type of infection are *Salmonella typhimurium*, *Salmonella* enteritidis, and *Salmonella newport* (Bakhshandeh et al., 2022). The infection typically lasts 4–7 days, with an incubation period ranging from 6 hours to a week following the initial inoculation (Pulford et al., 2021). Gastroenteritis is the most prevalent symptom in humans, with clinical manifestations such as nausea, vomiting, abdominal pain, diarrhea, and muscle soreness (Hohmann, 2001). The severity of the infections increases, among immunocompromised patients, elderly immunocompromised adults, and newborns and toddlers under five years of age (Scallan et al., 2011). Cholecystitis, pancreatitis, and appendicitis are among the disorders that might appear and worsen, eventually developing into meningitis and sepsis, which are life-threatening. Dehydration, which can be fatal in babies and older individuals, can result from inadequate fluid balance brought on by a protracted loss of body fluids (Schempp et al., 2019, Ehuwa et al., 2021).

Pathogenesis of Salmonellosis

Salmonellosis is more likely to affect the elderly, young children (less than five years old), and people with immunocompromised patients. *Salmonella* exhibited a peculiar trait during its colonization of non-phagocytic cells (Crump, 2019), whereby it initiates phagocytosis in order to gain entry into the host cell. The *Salmonella* pathogenicity islands (SPIs), a group of genes located in a vast section of the chromosome DNA and representing the structural domains that take part in the invasion process, contain these virulence factors (Grassl & Finlay, 2008). After entering the digestive system by tainted food or drink, the bacteria tend to pierce the intestinal wall epithelial cells. The specialized microfold cells found in the lymphoid tissue, commonly known as Peyer's patches (Dillon & Lo, 2019), or the active penetration of nonphagocytic cells through the so-called "trigger" process (Roche et al., 2018) are the primary mechanisms by which *Salmonella* is transported across the intestinal barrier (Dos Santos et al., 2020).

Salmonella's pathogenicity depends on its intracellular persistence, which can differ across strains with high and low virulence (Choi et al., 2019, Pradhan & Negi, 2019). A host-derived membrane sheath called a vacuole also referred to as a *Salmonella*-containing vacuole, or SCV envelops the bacterium upon engulfment. After the phagosome and lysosome fuse, the host cell releases reactive oxygen species and enzymes to eliminate the bacteria it has acquired (Villanueva et al., 2022). The bacterium uses the T₃SS to inject effector proteins straight into the vacuole, changing the structure of the compartment. In order to safeguard the organism in the intracellular niche and facilitate safe reproduction, the altered vacuole shape prevents phagolysosomal fusion (Reddy et al., 2010). A noticeable temperature pattern appears during enteric fever (Thakur et al., 2022). If proper treatment is not received, the fever may last up to one month (Patel et al., 2010). Infected people may have bradycardia, myalgia, splenomegaly, hepatomegaly, and the appearance of red on the abdomen and chest in addition to fever (Kuvandik et al., 2009). Hepatitis, pancreatitis, and cholecystitis (Galán, 2016).

Virulent Factors

There are several virulent factors associated with the pathogenesis of salmonellosis such as:

1. Virulence plasmids are essential for bacteria because they contain genes linked to virulence factors like spvB (ADP-ribosylating toxin) spvC (inhibits inflammation and pyroptosis) and antibiotic resistance (Zuo et al., 2020, Mellor et al., 2022). These plasmids can transmit horizontal gene transfer by transformation and conjugation (Rodríguez-Beltrán et al., 2021). To avoid being retained throughout cell division, they are big and present in small quantities to reduce the burden on the host's cell metabolism (Sengupta & Austin, 2011).

2. *Salmonella's* extracellular area is supplied with a variety of substances, including toxins, surface proteins, lipases, and adenylate cyclase, by the Type 3 secretion system (Li et al., 2019). Many plant and animal Gram-negative pathogens have complex syringe-like nanomachines called type-3 secretion systems (Kosarewicz et al., 2012).

3. There are two separate pathogenicity islands, SPI1 and SPI2, encode the T₃SS in *Salmonella* (Srikanth et al., 201). SPI-1, which encodes the T₃SS1, is necessary for invasive non-phagocytic epithelia (McGhie et al., 2009, LaRock et al., 2015). The T₃SS2 effector proteins, which are encoded by SPI-2, control the kinetics of *Salmonella*-containing vacuole membranes, locate SCVs in certain areas inside host cells, affect immunological responses, alter the cytoskeleton, and affect infected cell motility (Sellin et al., 2014).

4. The enzyme superoxide dismutase (SOD) is essential for preserving the cells' redox potential. One very important job of this substance is to protect healthy cells from the reactive oxygen species that are made when many intracellular pathogens infect cells (Felmy et al., 2013, Tang et al., 2022). Through their ability to intercept reactive oxygen species generated by the host's natural immune system, both enzymes are believed to contribute to Salmonella pathogenicity (Krishnakumar et al., 2007, Wang et al., 2020).

5. Among the most crucial elements of Salmonella attachment is fimbriae. Three different assembly pathways chaperon–usher, nucleation– precipitation, and type IV fimbriae are used to assemble Salmonella fimbriae. Several kinds of fimbriae result from these assembly processes. To start adhesion, *Salmonella* fimbriae attach to receptors on host cells (Grzymajło et al., 2013, Kolenda et al., 2019, Rehman et al., 2019, Uchiya et al., 2019).

6. More than 20 distinct proteins come together to form the intricate mechanism that is the bacterial flagellum (Wang et al., 2020). Although the flagellum has historically solely been thought of as a mobility organelle, it has lately become clear that flagella serve a variety of additional biological purposes (Haiko & Westerlund-Wikström, 2013; Horstmann et al., 2017; Sokaribo et al., 2020).

7. "Vi antigen," a capsular polysaccharide produced by *Salmonella enterica serovar Typhi*, is made up of residues of nonstoichiometrically Oacetylated α-1,4-linked N-acetylgalactosaminuronic acid. Current vaccinations contain this glycoprotein (Hiyoshi et al., 2018).

8. A crucial component of *Salmonella Typhi's* pathogenicity, which causes typhoid fever in humans, is typhoid toxin (Liston et al., 2016). The peculiar biology of this toxin is that Salmonella Typhi only produces it when it is within host cells. After being produced, the toxin is released into the lumen of the vacuole which contains *Salmonella*, where vesicle carrier intermediates carry it to the extracellular area (Chong et al., 2017).

9. Asymmetric lipid bilayers made up of outer leaflet lipopolysaccharides and inner leaflet phospholipids make up Gram-negative bacteria's outer membrane (OM) (Needham & Trent, 2013). Lipid A, O-antigen polysaccharide (O-PS), and core oligosaccharide (C-OS) comprise LPS (Kong et al., 2011). Additionally, LPS is in charge of host epithelial adhesion or invasion (Richards et al., 2010, Baptista et al., 2023).

10. The ability to produce biofilms and the capacity to induce acute, latent, or chronic disease are traits shared by many Salmonella species (Römling, 2005, Ismael et al., 2024). The adaptive response known as biofilms has the potential to modify bacterial gene expression, hence enhancing resistance to environmental stresses and drugs (Kader et al., 2006, Chen et al., 2012). The secretion of a polymeric matrix that is defined by the expression of several elements, including the two main components, cellulose and curli fimbriae, forms a *Salmonella* biofilm (Sanad et al., 2016).

Antibiotic Resistance to Salmonellosis

Recently, *Salmonella typhimurium* has developed multidrug resistance, rendering it untreatable with conventional antibiotics and potentially leading to deadly infections. In contrast to *Salmonella enterica* serovar *Typhimurium* LT2, *Salmonella enterica* serovar *Typhimurium* DT104 showed multidrug resistance to the antibiotic's chloramphenicol, ampicillin, tetracycline, florfenicol, streptomycin, and sulfonamides using a gene identified in *Salmonella* genomic island I (Al-Saeed et al., 2023, Abdullah et al., 2024).

Concern about antimicrobial resistance in salmonella (1,4,[5],12: -:-) has grown over the last 20 years, especially in multidrug-resistant isolates (MDR) (Vestergaard et al., 2016, Proroga et al., 2019, Qin et al., 2022). These MDR strains have an association with the rising prevalence of human salmonellosis (Mu et al., 2022, Wei et al., 2023). There are two dominant clonal lineages of salmonella multidrug resistance (1,4,[5],12: i:-:) are a European clone with an ASSuT (ampicillin, streptomycin, sulfonamides, and tetracycline) resistance pattern (Zeng et al., 2021; Keestra-Gounder et al., 2015). Salmonella (1,4,[5],12: i:-) isolates mostly from clinical sources had a high MDR rate (86.0%) in a meta-analysis (Hopkins et al., 2010; Nadimpalli et al., 2019). *Salmonella* (1,4,[5],12: i:-) isolates have also been found to be resistant to colistin, quinolones, and extended-spectrum β -lactamases (ESBLs) (García et al., 2011, Casas et al., 2016). Cleaning and sanitation practices, routine food handler screening and diagnosis, routine animal monitoring for potential carriers, and treatment of carriers and symptomatic individuals are some of these safeguards (Mancini et al., 2021, AlFaleh et al., 2023). An animal Salmonellosis needs a regular check for Salmonella infection should be conducted throughout all stages of the animal production chain (Ismael et al., 2023).

Conclusion

This chapter illustrated how Salmonella can occasionally contaminate food along the farm-to-fork chain and showed how healthy hens can harbor harmful and resistant forms of the bacteria. *Salmonella enterica* serovar is a newly discovered pathogen that can cause foodborne illnesses, especially when it contaminates foods that come from animals. New compounds and therapeutic targets that can treat *Salmonella* infections and prevent the emergence of antibiotic resistance may be found with the aid of modern technology and molecular techniques. It is essential to increase the monitoring of food isolates obtained from animals in order to curtail and control the transmission of them throughout the food chain.

References

- Abdullah, B. H., Ismael, S. S., & Qasim, N. A. (2024). Antimicrobial Susceptibility of Proteus mirabilis Isolated from Urinary Tract Infections in Duhok City, Iraq, Using VITEK2 System. *European Journal of Medical and Health Research*, 2(4), 75-79. https://doi.org/10.59324/ejmhr.2024.2(4).09
- AlFaleh, F. A., Ismael, S. S., Aguilar-Marcelino, L., Silva, F. E. M., Ashraf, T., Abbas, R. Z., & Qamar, W. (2023). Use of nanoparticles, a modern means of drug delivery, against cryptosporidiosis. *Journal of Advanced Veterinary and Animal Research*, 10(4), 704–719. https://doi.org/10.5455/javar.2023.j726
- Al-Saeed, F. A., Ismael Bamarni, S. S., Iqbal, K. J., Faruk, A. Z., Mahmood, S., Şahin, T., ... & Riaz, R. (2023). In vitro anthelmintic efficacy of *Haloxylon salicornicum* leaves extract using adult Heamonchus contortus worms. *Pakistan Veterinary Journal*, 43(1), 91-96. https://doi.org/10.29261/pakvetj/2022.091
- Bakhshandeh, B., Sorboni, S. G., Haghighi, D. M., Ahmadi, F., Dehghani, Z., & Badiei, A. (2022). New analytical methods using carbon-based nanomaterials for detection of Salmonella species as a major food poisoning organism in water and soil resources. *Chemosphere*, 287(3), 132243. https://doi.org/10.1016/j.chemosphere.2021.132243
- Baptista, D., Borsoi, A., Reischak, D., Nascimento, A., Montesino, L., Camillo, S., Abreu, D., & Pereira, V. (2023). Salmonella Serovars Isolated from Poultry Breeding Flocks under the Brazilian Official Control Programme Between 2016 and 2018. *Brazilian Journal of Poultry Science*, 25, 1. https://doi.org/10.1590/1806-9061-2022-1646
- Casas, M.R., Camargo, C.H., Soares, F.B., da Silveira, W.D., & Fernandes, S.A. (2016). Presence of plasmid-mediated quinolone resistance determinants and mutations in gyrase and topoisomerase in Salmonella enterica isolates with resistance and reduced susceptibility to ciprofloxacin. *Diagnostic Microbiology and Infectious Disease*, *85*(1), 85-9.
- Center for Disease Control and Prevention (CDC). Drug-Resistant Nontyphoidal Salmonella. (2023). https://www.cdc.gov/drugresistance/pdf/threats-report/nt-salmonella-508.pdf
- Chen, C., Tsen, H.Y., Lin, C., Yu, B., & Chen, C. (2012). Oral administration of a combination of select lactic acid bacteria strains to reduce the Salmonella invasion and inflammation of broiler chicks. *Poultry Science*, *91*(*9*), 2139-47. https://doi.org/10.3382/ps.2012-02237
- Chlebicz, A., & Śliżewska, K. (2018). Campylobacteriosis, Salmonellosis, Yersiniosis, and Listeriosis as Zoonotic Foodborne Diseases: A Review. International Journal of Environmental Research and Public Health, 15(5), 863. https://doi.org/10.3390/ijerph15050863
- Choi, S., Choi, E., Cho, Y.J., Nam, D.L., & Jangwoo, L.E. (2019). The *Salmonella* virulence protein MgtC promotes phosphate uptake inside macrophages. *Nature Communications, 10*, 3326 https://doi.org/10.1038/s41467-019-11318-2

- Chong, A., Lee, S., Yang, Y. A., & Song, J. (2017). The Role of Typhoid Toxin in Salmonella Typhi Virulence Typhi Virulence (P). The Yale Journal of Biology and Medicine, 90(2), 283–290.
- Cilia, G., Turchi, B., Fratini, F., Bilei, S., Bossù, T., De Marchis, M. L., Cerri, D., Pacini, M. I., & Bertelloni, F. (2021). Prevalence, Virulence and Antimicrobial Susceptibility of *Salmonella* spp., *Yersinia enterocolitica* and *Listeria monocytogenes* in European Wild Boar (*Sus scrofa*) Hunted in Tuscany (Central Italy). *Pathogens (Basel, Switzerland)*, 10(2), 93. https://doi.org/10.3390/pathogens10020093
- Dos Santos, A. M. P., Ferrari, R. G., & Conte-Junior, C. A. (2020). Type three secretion system in Salmonella Typhimurium: the key to infection. *Genes & Genomics*, 42(5), 495–506. https://doi.org/10.1007/s13258-020-00918-8
- Dróżdż, M., Małaszczuk, M., Paluch, E., & Pawlak, A. (2021). Zoonotic potential and prevalence of *Salmonella* serovars isolated from pets. *Infection Ecology & Epidemiology*, 11(1), 1975530. https://doi.org/10.1080/20008686.2021.1975530
- Ehuwa, O., Jaiswal, A. K., & Jaiswal, S. (2021). Salmonella, Food Safety and Food Handling Practices. Foods (Basel, Switzerland), 10(5), 907. https://doi.org/10.3390/foods10050907
- Fang, L., Shen, H., Tang, Y., & Fang, W. (2015). Superoxide dismutase of Streptococcus suis serotype 2 plays a role in anti-autophagic response by scavenging reactive oxygen species in infected macrophages. *Veterinary Microbiology*, 176(3-4), 328–336. https://doi.org/10.1016/j.vetmic.2015.02.006
- Felmy, B., Songhet, P., Slack, E. M., Müller, A. J., Kremer, M., Van Maele, L., Cayet, D., Heikenwalder, M., Sirard, J. C., & Hardt, W. D. (2013). NADPH oxidase deficient mice develop colitis and bacteremia upon infection with normally avirulent, TTSS-1- and TTSS-2-deficient Salmonella Typhimurium. PloS One, 8(10), e77204. https://doi.org/10.1371/journal.pone.0077204
- Ford, L., Buuck, S., Eisenstein, T., Cote, A., McCormic, Z. D., Kremer-Caldwell, S., Kissler, B., Forstner, M., Sorenson, A., Wise, M. E., Smith, K., Medus, C., Griffin, P. M., & Robyn, M. (2023). Salmonella Outbreaks Associated with Not Ready-to-Eat Breaded, Stuffed Chicken Products - United States, 1998-2022. Morbidity and Mortality Weekly Report, 72(18), 484–487. https://doi.org/10.15585/mmwr.mm7218a2
- Galán J. E. (2016). Typhoid toxin provides a window into typhoid fever and the biology of Salmonella Typhi. *Proceedings of the National Academy of Sciences of the United States of America*, 113(23), 6338–6344. https://doi.org/10.1073/pnas.1606335113
- Galán J. E. (2021). Salmonella Typhimurium and inflammation: a pathogen-centric affair. *Nature Reviews Microbiology*, 19(11), 716–725. https://doi.org/10.1038/s41579-021-00561-4
- García, P., Guerra, B., Bances, M., Mendoza, M. C., & Rodicio, M. R. (2011). IncA/C plasmids mediate antimicrobial resistance linked to virulence genes in the Spanish clone of the emerging Salmonella enterica serotype 4,[5],12:i:-. *The Journal of Antimicrobial Chemotherapy*, 66(3), 543–549. https://doi.org/10.1093/jac/dkq481
- Giannelli, C., Cappelletti, E., Di Benedetto, R., Pippi, F., Arcuri, M., Di Cioccio, V., Martin, L. B., Saul, A., & Micoli, F. (2017). Determination of free polysaccharide in Vi glycoconjugate vaccine against typhoid fever. *Journal of Pharmaceutical and Biomedical Analysis*, 139, 143–147. https://doi.org/10.1016/j.jpba.2017.02.042
- Gil Molino, M., García Sánchez, A., Risco Pérez, D., Gonçalves Blanco, P., Quesada Molina, A., Rey Pérez, J., & Fernández Llario, P. (2019). Prevalence of Salmonella spp. in tonsils, mandibular lymph nodes and faeces of wild boar from Spain and genetic relationship between isolates. *Transboundary and Emerging Diseases*, *66*(3), 1218–1226. https://doi.org/10.1111/tbed.13140
- Gonzalez-Escobedo, G., & Gunn, J. S. (2013). Gallbladder epithelium as a niche for chronic Salmonella carriage. *Infection and Immunity*, 81(8), 2920–2930. https://doi.org/10.1128/IAI.00258-13
- Grassl, G.A., & Finlay, B.B. (2008). Pathogenesis of enteric Salmonella infections. Current Opinion in Gastroenterology, 24, 22-26.
- Griffith, R.W., Carlson, S.A., & Krull, A.C. (2019). Salmonellosis. Diseases of Swine, Eleventh Edition, 912–925. https://doi.org/10.1002/9781119350927.ch59
- Grzymajło, K., Ugorski, M., Kolenda, R., Kędzierska, A., & Wieliczko, A. (2013). FimH adhesin from host unrestricted Salmonella Enteritidis binds to different glycoprotein ligands expressed by enterocytes from sheep, pig and cattle than FimH adhesins from host restricted Salmonella Abortus-ovis, Salmonella Choleraesuis and Salmonella Dublin. Veterinary Microbiology, 166(3-4), 550–557. https://doi.org/10.1016/j.vetmic.2013.07.004
- Haiko, J., & Westerlund-Wikström, B. (2013). The role of the bacterial flagellum in adhesion and virulence. *Biology*, 2(4), 1242–1267. https://doi.org/10.3390/biology2041242
- Hailu, W., Helmy, Y. A., Carney-Knisely, G., Kauffman, M., Fraga, D., & Rajashekara, G. (2021). Prevalence and Antimicrobial Resistance Profiles of Foodborne Pathogens Isolated from Dairy Cattle and Poultry Manure Amended Farms in Northeastern Ohio, the United States. Antibiotics (Basel, Switzerland), 10(12), 1450. https://doi.org/10.3390/antibiotics10121450
- Helmy, Y. A., El-Adawy, H., & Abdelwhab, E. M. (2017). A Comprehensive Review of Common Bacterial, Parasitic and Viral Zoonoses at the Human-Animal Interface in Egypt. *Pathogens (Basel, Switzerland)*, 6(3), 33. https://doi.org/10.3390/pathogens6030033
- Hiyoshi, H., Tiffany, C. R., Bronner, D. N., & Bäumler, A. J. (2018). Typhoidal Salmonella serovars: ecological opportunity and the evolution of a new pathovar. *FEMS Microbiology Reviews*, *42*(4), 527–541. https://doi.org/10.1093/femsre/fuy024
- Hoelzer, K., Moreno Switt, A. I., & Wiedmann, M. (2011). Animal contact as a source of human non-typhoidal salmonellosis. *Veterinary Research*, 42(1), 34. https://doi.org/10.1186/1297-9716-42-34
- Hoffmann, S., Batz, M. B., & Morris, J. G., Jr (2012). Annual cost of illness and quality-adjusted life year losses in the United States due to 14 foodborne pathogens. *Journal of Food Protection*, 75(7), 1292–1302. https://doi.org/10.4315/0362-028X.JFP-11-417
- Hohmann E. L. (2001). Nontyphoidal salmonellosis. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*, 32(2), 263–269. https://doi.org/10.1086/318457
- Hopkins, K. L., Kirchner, M., Guerra, B., Granier, S. A., Jakubczak, A., Threlfall, E. J., & Mevius, D. J. (2010). Multiresistant Salmonella enterica serovar 4,[5], 12:i:- in Europe: a new pandemic strain?. *Euro surveillance: bulletin Europeen sur les Maladies Transmissibles = European Communicable Disease Bulletin*, 15(22), 19580.

- Horstmann, J. A., Zschieschang, E., Truschel, T., de Diego, J., Lunelli, M., Rohde, M., May, T., Strowig, T., Stradal, T., Kolbe, M., & Erhardt, M. (2017). Flagellin phase-dependent swimming on epithelial cell surfaces contributes to productive Salmonella gut colonisation. *Cellular Microbiology*, 19(8), 10.1111/cmi.12739. https://doi.org/10.1111/cmi.12739
- Ismael, S. S., Abdullah, B. H., Hasan, S. H., Sadiq, A. J., & Omer, D. M. (2023). How Using Probiotics in Poultry Can Fight the Spread of Antibiotic Resistance to Humans? *Al-Qadisiyah Journal for Agriculture Sciences*, 13(2), 105-109. https://doi.org/10.33794/qjas.2023.143760.1143
- Ismael, S.S., Alnakshabandie, W.M.Y., Sulaiman, SA., & Jalal, R.B. (2024). Methicillin resistant *Staphylococcus aureus* (MRSA). *Biological Times*, 3(4): 14-15.
- Jibril, A. H., Okeke, I. N., Dalsgaard, A., & Olsen, J. E. (2021). Association between antimicrobial usage and resistance in Salmonella from poultry farms in Nigeria. *BMC Veterinary Research*, *17*(1), 234. https://doi.org/10.1186/s12917-021-02938-2
- Kader, A., Simm, R., Gerstel, U., Morr, M., & Römling, U. (2006). Hierarchical involvement of various GGDEF domain proteins in rdar morphotype development of Salmonella enterica serovar Typhimurium. *Molecular Microbiology*, 60(3), 602–616. https://doi.org/10.1111/j.1365-2958.2006.05123.x
- Keestra-Gounder, A. M., Tsolis, R. M., & Bäumler, A. J. (2015). Now you see me, now you don't: the interaction of Salmonella with innate immune receptors. *Nature Reviews Microbiology*, *13*(4), 206–216. https://doi.org/10.1038/nrmicr03428
- Kolenda, R., Ugorski, M., & Grzymajlo, K. (2019). Everything You Always Wanted to Know About Salmonella Type 1 Fimbriae, but Were Afraid to Ask. Frontiers in Microbiology, 10, 1017. https://doi.org/10.3389/fmicb.2019.01017
- Kong, Q., Yang, J., Liu, Q., Alamuri, P., Roland, K. L., & Curtiss, R. (2011). Effect of deletion of genes involved in lipopolysaccharide core and Oantigen synthesis on virulence and immunogenicity of Salmonella enterica serovar typhimurium. *Infection and Immunity*, 79(10), 4227– 4239. https://doi.org/10.1128/IAI.05398-11
- Kosarewicz, A., Königsmaier, L., & Marlovits, T. C. (2012). The blueprint of the type-3 injectisome. *Philosophical transactions of the Royal* Society of London. Series B, Biological Sciences, 367(1592), 1140–1154. https://doi.org/10.1098/rstb.2011.0205
- Krishnakumar, R., Kim, B., Mollo, E.A., Imlay, J.A., & Slauch, J.M. (2007). Structural Properties of Periplasmic SodCI That Correlate with Virulence in Salmonella enterica Serovar Typhimurium. *Journal of Bacteriology*, 189, 4343 - 4352. https://doi.org/10.1128/JB.00010-07
- Kuvandik, C., Karaoglan, I., Namiduru, M., & Baydar, I. (2009). Predictive value of clinical and laboratory findings in the diagnosis of the enteric fever. *The New Microbiologica*, 32(1), 25–30.
- Lamichhane, B., Mawad, A. M. M., Saleh, M., Kelley, W. G., Harrington, P. J., 2nd, Lovestad, C. W., Amezcua, J., Sarhan, M. M., El Zowalaty, M. E., Ramadan, H., Morgan, M., & Helmy, Y. A. (2024). Salmonellosis: An Overview of Epidemiology, Pathogenesis, and Innovative Approaches to Mitigate the Antimicrobial Resistant Infections. *Antibiotics (Basel, Switzerland)*, *13*(1), *76*. https://doi.org/10.3390/antibiotics13010076
- Lamichhane, B., Mawad, A. M. M., Saleh, M., Kelley, W. G., Harrington, P. J., Lovestad, C. W., Amezcua, J., Sarhan, M. M., El Zowalaty, M. E., Ramadan, H., Morgan, M., & Helmy, Y. A. (2024). Salmonellosis: An Overview of Epidemiology, Pathogenesis, and Innovative Approaches to Mitigate the Antimicrobial Resistant Infections. *Antibiotics (Basel, Switzerland)*, 13(1), 76. https://doi.org/10.3390/antibiotics13010076
- LaRock, D. L., Chaudhary, A., & Miller, S. I. (2015). Salmonella's interactions with host processes. *Nature Reviews Microbiology*, 13(4), 191–205. https://doi.org/10.1038/nrmicr03420
- Li, X., Bleumink-Pluym, N. M. C., Luijkx, Y. M. C. A., Wubbolts, R. W., van Putten, J. P. M., & Strijbis, K. (2019). MUC1 is a receptor for the Salmonella SiiE adhesin that enables apical invasion into enterocytes. *PLoS Pathogens*, 15(2), e1007566. https://doi.org/10.1371/journal.ppat.1007566
- Majowicz, S. E., Musto, J., Scallan, E., Angulo, F. J., Kirk, M., O'Brien, S. J., Jones, T. F., Fazil, A., Hoekstra, R. M., & International Collaboration on Enteric Disease 'Burden of Illness' Studies (2010). The global burden of nontyphoidal Salmonella gastroenteritis. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*, 50(6), 882–889. https://doi.org/10.1086/650733
- McGhie, E. J., Brawn, L. C., Hume, P. J., Humphreys, D., & Koronakis, V. (2009). Salmonella takes control: effector-driven manipulation of the host. *Current Opinion in Microbiology*, *12*(1), 117–124. https://doi.org/10.1016/j.mib.2008.12.001
- Mellor, K. C., Blackwell, G. A., Cawthraw, S. A., Mensah, N. E., Reid, S. W. J., Thomson, N. R., Petrovska, L., & Mather, A. E. (2022). Contrasting long-term dynamics of antimicrobial resistance and virulence plasmids in *Salmonella* Typhimurium from animals. *Microbial Genomics*, 8(8), mgen000826. https://doi.org/10.1099/mgen.0.000826
- Mu, Y., Li, R., Du, P., Zhang, P., Li, Y., Cui, S., Fanning, S., & (2022). Bai, L. Genomic epidemiology of ST34 monophasic salmonella enterica serovar typhimurium from clinical patients from 2008 to 2017 in Henan, China. *Engineering*, *15*, 34-44.
- Nadimpalli, M., Fabre, L., Yith, V., Sem, N., Gouali, M., Delarocque-Astagneau, E., Sreng, N., Le Hello, S., & BIRDY study group (2019). CTX-M-55-type ESBL-producing Salmonella enterica are emerging among retail meats in Phnom Penh, Cambodia. *The Journal of Antimicrobial Chemotherapy*, 74(2), 342–348. https://doi.org/10.1093/jac/dky451
- Needham, B. D., & Trent, M. S. (2013). Fortifying the barrier: the impact of lipid A remodelling on bacterial pathogenesis. *Nature Reviews Microbiology*, 11(7), 467–481. https://doi.org/10.1038/nrmicr03047
- Olnood, C. G., Beski, S. S. M., Choct, M., & Iji, P. A. (2015). Use of *Lactobacillus johnsonii* in broilers challenged with *Salmonella sofia*. *Animal Nutrition (Zhongguo xu mu shou yi xue hui)*, 1(3), 203–212. https://doi.org/10.1016/j.aninu.2015.07.001
- Patel, T. A., Armstrong, M., Morris-Jones, S. D., Wright, S. G., & Doherty, T. (2010). Imported enteric fever: case series from the hospital for tropical diseases, London, United Kingdom. *The American Journal of Tropical Medicine and Hygiene*, 82(6), 1121–1126. https://doi.org/10.4269/ajtmh.2010.10-0007
- Pradhan, D., & Negi, V. D. (2019). Repeated in-vitro and in-vivo exposure leads to genetic alteration, adaptations, and hypervirulence in Salmonella. *Microbial Pathogenesis*, *136*, 103654. https://doi.org/10.1016/j.micpath.2019.103654
- Proroga, Y. T. R., Mancusi, A., Peruzy, M. F., Carullo, M. R., Montone, A. M. I., Fulgione, A., & Capuano, F. (2019). Characterization of Salmonella

Typhimurium and its monophasic variant 1,4, [5],12:i:- isolated from different sources. Folia Microbiologica, 64(6), 711–718. https://doi.org/10.1007/s12223-019-00683-6

- Pulford, C.V., Perez-Sepulveda, B.M., Canals, R. *et al.* (2021). Stepwise evolution of *Salmonella Typhimurium* ST313 causing bloodstream infection in Africa. *Nature Microbiology*, *6*, 327–338. https://doi.org/10.1038/s41564-020-00836-1
- Qin, X., Yang, M., Cai, H., Liu, Y., Gorris, L., Aslam, M. Z., Jia, K., Sun, T., Wang, X., & Dong, Q. (2022). Antibiotic Resistance of *Salmonella* Typhimurium Monophasic Variant 1,4,[5],1 2:i:-in China: A Systematic Review and Meta-Analysis. *Antibiotics (Basel, Switzerland)*, 11(4), 532. https://doi.org/10.3390/antibiotics11040532
- Reddy, E. A., Shaw, A. V., & Crump, J. A. (2010). Community-acquired bloodstream infections in Africa: a systematic review and metaanalysis. *The Lancet Infectious Diseases*, 10(6), 417–432. https://doi.org/10.1016/S1473-3099(10)70072-4
- Rehman, T., Yin, L., Latif, M. B., Chen, J., Wang, K., Guo, H., & Ouyang, P. (2019). Adhesive mechanism of different Salmonella fimbrial adhesins. *Microbial Pathogenesis*, 137, 103748. https://doi.org/10.1016/j.micpath.2019.103748
- Richards, S. M., Strandberg, K. L., & Gunn, J. S. (2010). Salmonella-regulated lipopolysaccharide modifications. *Sub-cellular Biochemistry*, 53, 101–122. https://doi.org/10.1007/978-90-481-9078-2_5
- Roche, S. M., Holbert, S., Trotereau, J., Schaeffer, S., Georgeault, S., Virlogeux-Payant, I., & Velge, P. (2018). Salmonella Typhimurium Invalidated for the Three Currently Known Invasion Factors Keeps Its Ability to Invade Several Cell Models. Frontiers in Cellular and Infection Microbiology, 8, 273. https://doi.org/10.3389/fcimb.2018.00273
- Rodríguez-Beltrán, J., DelaFuente, J., León-Sampedro, R., MacLean, R. C., & San Millán, Á. (2021). Beyond horizontal gene transfer: the role of plasmids in bacterial evolution. *Nature Reviews Microbiology*, 19(6), 347–359. https://doi.org/10.1038/s41579-020-00497-1
- Römling U. (2005). Characterization of the rdar morphotype, a multicellular behaviour in Enterobacteriaceae. *Cellular and Molecular Life Sciences: CMLS*, 62(11), 1234–1246. https://doi.org/10.1007/s00018-005-4557-x
- Sanad, Y.M., Johnson, K., Park, S.H., Han, J., Deck, J., Foley, S.L., Kenney, B., Ricke, S., & Nayak, R. (2016). Molecular characterization of Salmonella enterica serovars isolated from a turkey production facility in the absence of selective antimicrobial pressure. *Foodborne Pathogens and Disease*, 13(2), 80-7. https://doi.org/10.1089/fpd.2015.2002
- Sánchez-Vargas, F. M., Abu-El-Haija, M. A., & Gómez-Duarte, O. G. (2011). Salmonella infections: an update on epidemiology, management, and prevention. *Travel Medicine and Infectious Disease*, 9(6), 263–277. https://doi.org/10.1016/j.tmaid.2011.11.001
- Scallan, E., Hoekstra, R. M., Angulo, F. J., Tauxe, R. V., Widdowson, M. A., Roy, S. L., Jones, J. L., & Griffin, P. M. (2011). Foodborne illness acquired in the United States--major pathogens. *Emerging Infectious Diseases*, 17(1), 7–15. https://doi.org/10.3201/eid1701.p11101
- Schempp, C. M., Schauer, F., Huhn, C. K., Venhoff, N., & Finzel, S. (2019). Skin inflammation associated with arthritis, synovitis and enthesitis. Part 2: rheumatoid arthritis, reactive arthritis, Reiter's syndrome, Lyme borreliosis, dermatomyositis and lupus erythematosus. *Journal der Deutschen Dermatologischen Gesellschaft = Journal of the German Society of Dermatology: JDDG*, 17(2), 167–181. https://doi.org/10.1111/ddg.13761
- Sears, K. T., Galen, J. E., & Tennant, S. M. (2021). Advances in the development of Salmonella-based vaccine strategies for protection against Salmonellosis in humans. *Journal of Applied Microbiology*, 131(6), 2640–2658. https://doi.org/10.1111/jam.15055
- Sellin, M. E., Müller, A. A., Felmy, B., Dolowschiak, T., Diard, M., Tardivel, A., Maslowski, K. M., & Hardt, W. D. (2014). Epithelium-intrinsic NAIP/NLRC4 inflammasome drives infected enterocyte expulsion to restrict Salmonella replication in the intestinal mucosa. *Cell Host & Microbe*, *16*(2), 237–248. https://doi.org/10.1016/j.chom.2014.07.001
- Sengupta, M., & Austin, S. (2011). Prevalence and significance of plasmid maintenance functions in the virulence plasmids of pathogenic bacteria. *Infection and Immunity*, 79(7), 2502–2509. https://doi.org/10.1128/IAI.00127-11
- Sokaribo, A. S., Hansen, E. G., McCarthy, M., Desin, T. S., Waldner, L. L., MacKenzie, K. D., Mutwiri, G., Jr, Herman, N. J., Herman, D. J., Wang, Y., & White, A. P. (2020). Metabolic Activation of CsgD in the Regulation of Salmonella Biofilms. *Microorganisms*, 8(7), 964. https://doi.org/10.3390/microorganisms8070964
- Somorin, Y.M., Odeyemi, O.A., Ateba, C.N. (2021). *Salmonella* is the most common foodborne pathogen in African food exports to the European Union: Analysis of the Rapid Alert System for Food and Feed (1999–2019). *Food Control*, *123*, 107849.
- Srikanth, C. V., Mercado-Lubo, R., Hallstrom, K., & McCormick, B. A. (2011). Salmonella effector proteins and host-cell responses. *Cellular and Molecular Life Sciences: CMLS*, 68(22), 3687–3697. https://doi.org/10.1007/s00018-011-0841-0

Stoycheva, M. V., & Murdjeva, M. A. (2006). Antimicrobial therapy of salmonelloses--current state and perspectives. *Folia Medica*, 48(1), 5–10. Tang, B., Elbediwi, M., Nambiar, R.B., Yang, H., Lin, J., & Yue, M. (2022). Genomic Characterization of Antimicrobial-Resistant Salmonella

- enterica in Duck, Chicken, and Pig Farms and Retail Markets in Eastern China. Microbiology Spectrum, 10, e01257-22. Thakur, R., Suri, C. R., & Rishi, P. (2022). Contribution of typhoid toxin in the pathogenesis of Salmonella Typhi. *Microbial Pathogenesis*, 164,
- 105444. https://doi.org/10.1016/j.micpath.2022.105444
- Uchiya, K. I., Kamimura, Y., Jusakon, A., & Nikai, T. (2019). Salmonella Fimbrial Protein FimH Is Involved in Expression of Proinflammatory Cytokines in a Toll-Like Receptor 4-Dependent Manner. *Infection and Immunity*, 87(3), e00881-18. https://doi.org/10.1128/IAI.00881-18
- Van Immerseel F., Russell J.B., Flythe M.D., Gantois I., Timbermont L., Pasmans F., Haesebrouck F., & Ducatelle R. (2006). The use of organic acids to combat Salmonella in poultry: A mechanistic explanation of the efficacy. Avian Pathology, 35, 182–188. https://doi.org/10.1080/03079450600711045
- van Panhuis, W. G., Grefenstette, J., Jung, S. Y., Chok, N. S., Cross, A., Eng, H., Lee, B. Y., Zadorozhny, V., & Burke, D. S. (2013). Contagious diseases in the United States from 1888 to the present. *The New England Journal of Medicine*, 369(22), 2152–2158. https://doi.org/10.1056/NEJMms1215400
- Vestergaard, M., Paulander, W., Marvig, R. L., Clasen, J., Jochumsen, N., Molin, S., Jelsbak, L., Ingmer, H., & Folkesson, A. (2016). Antibiotic combination therapy can select for broad-spectrum multidrug resistance in Pseudomonas aeruginosa. *International Journal of*

Antimicrobial Agents, 47(1), 48-55. https://doi.org/10.1016/j.ijantimicag.2015.09.014

- Villanueva, J.A., Crooks, A.L., Nagy, T.A., Quintana, J.L.J., Dalebroux, Z.D., & Detweiler, C.S. (2022). Salmonella enterica Infections Are Disrupted by Two Small Molecules That Accumulate within Phagosomes and Differentially Damage Bacterial Inner Membranes. *mBio*, *13*, e01790-22.
- Wang, F., Deng, L., Huang, F., Wang, Z., Lu, Q., & Xu, C. (2020). Flagellar Motility Is Critical for *Salmonella enterica* Serovar Typhimurium Biofilm Development. *Frontiers in Microbiology*, *11*, 1695. https://doi.org/10.3389/fmicb.2020.01695
- Wei, X., Long, L., You, L., Wang, M., Wang, D., Liu, C., Li, S., & Wang, J. (2023). Serotype distribution, trend of multidrug resistance and prevalence of β-lactamase resistance genes in human Salmonella isolates from clinical specimens in Guizhou, China. *PloS One*, 18(4), e0282254. https://doi.org/10.1371/journal.pone.0282254
- Wibisono, F.M., Wibisono, F.J., Effendi, M.H., Plumeriastuti, H., Hidayatullah, A.R., Hartadi, E.B., & Sofiana, E.D. (2020). A Review of Salmonellosis on Poultry Farms: Public Health Importance. *Systematic Reviews in Pharmacy*, *11*, 481-486.
- World Health Organization (WHO). Salmonella (Non-Typhoidal). (2023).: https://www.who.int/news-room/fact-sheets/detail/salmonella-(non-typhoidal)
- Zeng, X., Lv, S., Qu, C., Lan, L., Tan, D., Li, X., Bai, L. (2021). Serotypes, antibiotic resistance, and molecular characterization of non-typhoidal Salmonella isolated from diarrheic patients in Guangxi Zhuang Autonomous Region, China, 2014–2017. *Food Control, 120,* 107478.
- Zuo, L., Zhou, L., Wu, C., Wang, Y., Li, Y., Huang, R., & Wu, S. (2020). Salmonella spvC Gene Inhibits Pyroptosis and Intestinal Inflammation to Aggravate Systemic Infection in Mice. Frontiers in Microbiology, 11, 562491. https://doi.org/10.3389/fmicb.2020.562491