From Bats to Humans: Transmission and Pathogenesis of Ebola Virus

Alisha Munir^{1,*}, Ruhaba Khan² and Azzah Khadim Hussain³

¹Department of Zoology, Wildlife and Fisheries, University of Agriculture, Faisalabad, Pakistan ²Department of Biochemistry, University of Wah, Islamabad, Pakistan ³Department of Pharmacy, University of Central Punjab, Lahore, Pakistan *Corresponding author: alishamunir46@gmail.com

Abstract

Ebola virus disease (EVD) is a highly lethal zoonotic disease with a mortality rates of 90% caused by Ebola viruses (EBOV) belonging to *Filoviridae* family and *Ebolavirus* genus. The disease was identified in 1976 in Zaire, close to the Ebola River which is today part of the Democratic Republic of Congo (DRC). Since that time numerous outbreak have been observed in various regions globally, particularly in Africa, resulting in the discovery of 5 distinct viral strains that can infect human and primates. Fruit bats are thought to be the main reservoirs for the Ebola virus. The virus spreads by contact with infected animal's bushmeat and body fluids, as well as direct person-to-person transmission through bodily fluids, contaminated utensils and equipment, damaged skin, and mucous membranes. EVD pathogenesis involves fast viral multiplication and immunological evasion, resulting in extensive infection of immune and endothelial cells. This leads to systemic inflammation, vascular damage, and a reduced immune response, all of which result in severe symptoms such as bleeding, multiple organ malfunction, and, in many cases, death. This chapter discusses Ebola virus virology, reservoirs, transmission, pathogenesis, clinical symptoms, diagnosis, treatment, vaccination, and preventative methods.

Keywords: Ebola virus, Ebola virus disease, Fruit bats, Zoonotic disease

Cite this Article as: Munir A, Khan R and Hussain AK, 2025. From bats to humans: transmission and pathogenesis of ebola virus. In: Abbas RZ, Akhtar T and Jamil M (eds), Pathways of Infection: Zoonoses and Environmental Disease Transmission. Unique Scientific Publishers, Faisalabad, Pakistan, pp: 39-45. <u>https://doi.org/10.47278/book.HH/2025.26</u>



A Publication of Unique Scientific Publishers **Chapter No:** 25-006

Received: 19-Jan-2025 Revised: 21-March-2025 Accepted: 05-Apr-2025

Introduction

Ebola Virus (EBOV) is an emerging and re-emerging zoonotic disease that leads to severe Ebola hemorrhagic fever (EHF) in humans (Malik et al., 2023) and has a high mortality rate of 57% to 90% (Moghadam et al., 2015; Ghosh et al., 2021). It has a single-stranded RNA genome with a negative sense and belongs to the Filoviridae family (Hussein, 2013; Burk et al., 2016). It was called "Ebola" after the River Ebola near Yambuku, which served as the epicenter of the 1976 EBOV outbreak (Barrette et al., 2009). The first outbreak was discovered in Nizara (Sudan), at the end of June 1976, though the mode of transmission was not identified (Ascenzi et al., 2008). Later, in September of the same year, another outbreak took place near the village of Yambuku in the Democratic Republic of Congo (DRC, previously Zaire). The outbreak resulted in over 500 cases with a fatality rate of 53% in Sudan and 88% in Zaire (Ahmad et al., 2015). The latest epidemic in West Africa, occurring from 2013 to 2016, resulted in approximately 28,000 confirmed cases along with deaths of around 11,000 people, highlighting the high fatality rate of this disease (Garske et al., 2017). Ebola virus disease (EVD) affects humans as well as non-human primates (Feldmann & Feldmann, 2013; Weingart et al., 2013). It is classified as a category agent, which requires biosafety level 4 (BSL 4) precautions when working with suspected materials. Disease manifestation begins with fever as well as stomach ache, then vomiting, intravascular coagulation, diarrhea, bleeding and failure of multiple organs. In extreme situation the patient can experience hypovolemic shocks which results in mortality according to Mendoza et al. (2016). EBOV is spread by direct contact with the blood and body fluids of an infected animal or human and contact with their dead body (Passi et al., 2015). EBOV may have emerged from fruit bats. For instance, the 2007 EVD outbreak in Luebo, was connected to the ingestion of newly killed bat (Leroy et al., 2009). Likewise, the 2014 West Africa outbreak emerged with a zoonotic transmission from fruit bats to a 2-year-old Guinean kid (Marí Sa'ez, et al., 2015).

2. Ebola Virology

Ebola virus's family *Filoviridae*, which belongs to the *Mononegavirales* order, has three genera: *Cuevavirus*, *Ebolavirus*, and *Marburgvirus*. Within the *Ebolavirus* genus, five species have been discovered, each with its own virus: The species include *Ebola virus* (EBOV previously classified as *Zaire ebolavirus*), *Bundibugyo ebolavirus* (BDBV), *Sudan ebolavirus* (SUDV), *Reston ebolavirus* (RESTV), and *Tai ebolaforestvirus* (TAFV) (Feldmann & Geisbert, 2011). EBOVs are 50 - 80 nm in diameter and 10 000 - 14 000 nm in length. The Virions differ in structure; they are of cylindrical structure, branched and looped structure. However, all, members of the family *filoviridae* retain a unique thread like filamentous morphology (Majid et al., 2016). Ebola virus (EBOV) possess a large, un-fragmented, negative-sense single strand RNA genome of around 19kb, comprising seven genes arranged sequentially. They include 3' leader, nucleoprotein (NP), virion protein (VP35), matrix protein VP40, glycoprotein (GP), VP30, VP24 genes and the RNA dependent RNA polymerase (L), and at the last part the 5' trailer (Figure 1). Each gene

gives only one protein product excluding the GP gene which coded for three different glycoproteins (Mbala-Kingebeni et al., 2021). Sequence differences exist between all five EBOV species including positioning and number of gene overlaps, SUDV, EBOV, RESTV TAFV and BDBV all contain different sequences (Zheng et al., 2015; Goldstein et al., 2019).



3. Viral Reservoir

Fruit bats (*Myonycteristorquata, Epomopsfranqueti,* and *Hypsignathusmonstrosus*) from the family *Pteropodidae* are thought to be natural reservoirs for the EBOV. Bats have been confirmed to be natural reservoirs of Ebola virus (EBOV) with historical connections to outbreaks since 1976 and 1979, and Marburg virus diseases since 1975 and 1980. Experimental studies showed that out of 19 vertebrate and 24 plant species treated with Ebola virus, only bats exhibited no clinical indications of disease, supporting their role as reservoirs. A survey conducted between 2002 and 2003 involving 1,030 animals, which includes 679 bats from Congo and Gabon, detected EBOV RNA in 13 fruit bats (Gebretadik et al., 2015). Potential or accidental intermediate hosts comprise nonhuman primates, rodents, porcupines, antelopes, pigs, foxes, cats and duikers. Unlike bats, these animals normally get deadly and acute diseases once infected. They may carry the Ebola virus asymptotically and spread the virus through their body fluids, and in turn infect humans who come across them while hunting or butchering bushmeat (Osterholm et al., 2015; Singh et al., 2017; Laing et al., 2018).

4. Transmission

4.1. Transmission from Animals

EBOV outbreaks are usually initiated by direct contact with infected animals and their bodily fluids (for instance, when in hunting, butchering, or consuming infected animals) (Weingart et al., 2013). Members of the fruit bat family *Pteropodidae* are believed to be natural hosts of EBOV. People become infected with the virus either through contact with bats or other animals who are already infected (Figure 2) (Asare et al., 2023). Marí Sa'ez et al. (2015) have pointed out that the first contact of EVD in West Africa from 2014 to 2016 was linked to contacts with bats. Fruits and vegetables may become contaminated with the blood of infected animals, which people consume raw: eating infected bat meat is a major route of oral transmission of the EBOV, especially in African countries (Pandey et al., 2015; Caron et al., 2018; Munster et al., 2018). Apart from bats, cases of EVD have been observed in individuals who have interacted with intermediate hosts of EBOV such as infected chimpanzees, forest antelopes, gorillas, monkeys, and porcupines, both alive and dead, in Congo, Cote d'Ivoire, and Gabon (Jain et al., 2023). For instance, in 1996, in Mayebout, Gabon, a deceased chimpanzee discovered in the forest was slaughtered and consumed by 19 individuals and all of those individuals fell seriously sick within a short period. Since then, multiple similar incidents have occurred due to human interaction with infected chimpanzees or gorillas through hunting activities (Nkoghe et al., 2005). Pigs infected with EVD have significantly higher levels of virus in their lungs, than in their bloodstream. As a result, they can spread the disease by releasing infectious particles in the air or on to surfaces when they cough or sneeze (Funk & Kumar, 2015; Meyer et al., 2018).

4.2. Human to Human Transmission

Transmission among humans primarily occurs via physical contact with the bodily fluids of people who have fallen sick with EVD or who have passed away from it, especially when proper personal protective equipment (PPE) is not used (Figure 2) (Asare et al., 2023). Those at high risk include caregivers providing hands-on medical care and individuals preparing bodies for burial (Reichler et al., 2018). For instance, a meta-analysis containing nine researches of household transmission of EBOV, found that the secondary attack rates was 47.9% for individuals offering patient care, while it was 2.1% for those family members who had close physical interactions without offering patient care (Dean et al., 2016). Cleansing ritual of EBOV victims during funerals has significantly contributed to the spread of the virus in previous outbreaks,

including the West Africa epidemic. For Instance, a funeral ceremony in Guinea in late 2014 was associated with 85 consequent cases of EBOV disease (Victory et al., 2015). In the initial stages of the West African outbreak, hundreds of African doctors along with nurses contracted EVD while providing patient care without proper personal protective equipment. A historical investigation of within-household transmission during the West African outbreak discovered that virus propagated more frequently in bigger families. Additionally, transmissions were more common in elderly patients along with those individuals with severe illness, with an approximately 18% of secondary attack rate (Glynn et al., 2018). Lab workers in any biosafety level 4 facilities studying EBOV may face the risk of accidental infection. It can also be transmitted by the potential use of Ebola Viruses as biological weapons (Borio et al., 2002; Green, 2014).



Fig. 2: Transmission of Ebola virus disease.

4.3. Transmission through Different Body Fluids

The type of bodily fluid an individual encounters along with the viral load it carries impacts their risk of disease. The most common mode of Transmission is physical contact with unprotected mucous membranes or damaged skin exposed to virus-carrying bodily fluids from someone exhibiting disease symptoms (Figure 2) (Asare et al., 2023). Ebola virus transmission within hospital wards during the newly documented EBOV disease epidemic in 1976 in the DRC was associated with the usage of un-sterilized needles (Judson et al., 2015). The World Health Organization identifies blood, vomitus, and feces as highly infectious bodily fluids. Also, the Ebola virus has been found in saliva, semen, aqueous humor, urine, breast milk, and vaginal fluid (Kreuels et al., 2014; Nordenstedt et al., 2016). Reverse-transcription Polymerase chain reaction tests revealed viral RNA in sweat as well as in tears, indicating that these fluids may contain the pathogenic EBOV. It also transmits by close interaction with the patient's skin, although the danger of EVD from this kind of exposure is considered lesser compared to contact with bodily fluids or blood. The virus present over the skin may originate from viral replication inside the epidermal as well as dermal layers, contamination by blood as well as other bodily fluids, or a combination of both. The amount of virus present in the fluid also determines the risk of EBOV transmission. In the initial stages of disease, the viral load inside the blood might be relatively low, however it can quickly increase, potentially exceeding 10^8 RNA copies/mL of serum in critically sick also dying patients (Towner et al., 2004). Epidemiological researches indicated that household members were at the highest likelihood of disease when they came into direct exposure with ill relatives especially their bodily fluids in the final stages of disease or when assisting in the preparation of a dead body for burial (Glynn et al., 2018).

4.4. Persistent Risks of Transmission after Recovery

Even when EBOV or its viral RNA is no longer detectable in the bloodstream of an EVD recovered patient it can still be present in certain bodily fluids (Figure 2) (Asare et al., 2023). Men who have recovered from EVD can spread the virus via their semen. For instance: In followup tests with arround 40 survivors from the 1995 epidemic in Kikwit, Congo, RT-PCR identified viral RNA sequences in the male patient's semen for a period of three months. Additionally, an infectious virus was isolated from one individual's semen 82 days following the commencement of the disease (Bray et al., 2015). A Research conducted on patient samples gathered at the time of Sudan ebolavirus disease outbreak in 2000 in Gulu, Uganda, identified the presence of virus in a patient's breast milk long after the virus had become undetectable in the blood (Bausch et al., 2007). Two infants who were breastfed by infected moms succumbed to EVD. In a similar case, during the West Africa outbreak, a 9-month old baby succumbed to EVD, despite his mother's good health. Viral RNA was later discovered in her breast milk, however the blood as well as urine tests returned negative results (Sissoko et al., 2017).

4.5. Transmission through Contaminated Surfaces

Ebola virus (EBOV) can spread though exposure to contaminated surfaces or objects. Using infected needles as well as medical equipment

without sanitizing them first increases the risk of EBOV transmission. The virus can remain pathogenic for several hours to days on surfaces like clothing, utensils, doorknobs, electrical switches, furniture, bedding, and other objects that may become infected through bodily fluids (Osterholm et al., 2015). While there is no definitive evidence confirming transmission through contact with contaminated surfaces, proper environmental cleaning can significantly reduce or mitigate this potential risk (Chertow et al., 2022).

4.6. Airborne Transmission

Airborne or aerosol transmission of a virus happens as tiny, virus-containing droplets vaporize before landing on surfaces, leaving infectious droplet nuclei capable of traveling over long distances. These tiny droplets that can produce such droplet nuclei are commonly referred to as aerosols. More broadly, aerosols are defined as any solid particles or small liquid that are suspended in air (Tellier, 2009). Currently, there is no evidence of reported cases of EBOV transmission from individual to individual via the respiratory route. Nonetheless, laboratory investigation demonstrated that EBOV, when released as a tiny-particle aerosol can infect nonhuman primates and rodents (Zumbrun et al., 2012). A research using the Makona virus strain from the 2013-2016 West African outbreak showed that even minimal quantities of aerosolized virus proved fatal to cynomolgus macaques with the time until mortality depending on the dosage. As a result, health-care professionals may face a danger of EVD when subjected to aerosols produced throughout surgical treatments (Prasad et al., 2023).

5. Pathogenesis

Laboratory studies using non-human primates have yielded information on the pathogenesis of the Ebola virus disease (Kreuels et al., 2014). Ebola virus enters the body through mucous membranes, skin breaks or mother to child transmission (parenteral infection). It infects various cell types, particularly dendritic cells and macrophages, where it replicates, resulting in cell necrosis (Bray & Geisbert, 2005). The virus disseminates throughout the body by inhibiting type I interferon responses and replicating in the lymph nodes. It enters the bloodstream and infects macrophages and dendritic cells in various organs including the spleen, liver, and thymus along with other lymphoid tissues (Figure 3). It can also infect endothelial cells, epithelial cells, adrenal cortical cells, hepatocytes, and fibroblasts. Fatal infections result from multiple necrotic lesions in tissues like the liver and spleen (Basler, 2005).

Patients then experience diarrhoea and vomiting, leading to severe volume depletion, hypotension and shock. It remains unclear whether the gastrointestinal dysfunction directly induced by viral infection of the gastrointestinal tract (GIT) or as a result of circulating cytokines (Kortepeter et al., 2011). The viral infection triggers a systemic inflammatory syndrome through promoting the secretion of cytokines, chemokines, along with other pro-inflammatory substances by various cells like macrophages (Mahanty & Bray, 2004).

Infected macrophages release tumor necrosis factor (TNF)-alpha, interleukin (IL)-1beta, IL-6, and macrophage chemotactic protein (MCP)-1, nitric oxide (NO), which contribute to coagulation disorders in EVD via the host inflammatory reaction (Figure 3). Viral-infected macrophages produce tissue factor (TF), activating the extrinsic coagulation pathway, while pro-inflammatory cytokines further stimulate macrophages for the production of TF51. This dual mechanism leads to rapid and severe coagulopathy in EVD, with EBOV infected monkeys exhibiting elevated D-dimers within 24 hours. Furthermore, decreased dendritic cell activities and T lymphocyte apoptosis in infected persons hinder the adaptive immune responses, leading to severe illness (Bray, 2005).

EBOV infection disrupts antigen-specific immune reactions primarily by replicating within dendritic cells, crucial for commencing adaptive immunity (Figure 3). In vitro researches indicate that infected dentritic cells do not mature properly and cannot provide antigens to naive lymphocytes, which may explain the absence of antibody formation in patients who die from Ebola virus disease (Sanchez et al., 2004). The viral infection impairs Adaptive immune responses through apoptosis stimulated by inflammatory substances as well as a lack of support signals from dendritic cells, a process also seen in septic shock (Geisbert et al., 2003).

6. Clinical Manifestations and Diagnoses

The duration from Ebola virus infection along with the appearance of symptoms is 2-21 days, on an average of 8 to10 days. Humans become infectious only after they begin to show symptoms. Initial symptoms include fever, sore throat, fatigue, severe headache, muscle pain, as well as weakness. This is accompanied by diarrhoea, vomiting, stomach discomfort, and rash, signs of damaged liver and kidney function, as well as bleeding. Laboratory tests reveal low counts of white blood cells as well as platelets, along with increased levels of liver enzyme (Xu et al., 2016).

Differentiating Ebola from other pathogenic diseases like fever, meningitis, typhoid, and malaria is challenging. As stated by World Health Organization, confirming EVD involves several methods, such as electron microscopy, reverse transcriptase polymerase chain reaction (RT-PCR) test, serum neutralization test, antigen capture detection tests, virus isolation by cell culture, and antibody-capture enzyme-linked immunosorbent assay (ELISA) (Wambani et al., 2016).

7. Treatment

Today, US FDA has approved only two drugs for the management of the Ebola virus disease. The first one is Inmazeb—a pharmaceutical preparation containing three monoclonal antibodies such as odesivimab, maftivimab, as well as atoltivimab; it acts on virus glycoproteins and thereby interferes with their direct connection with cells of a host organism. The second is ansuvimab (mAb114) which is a kind of monoclonal antibody that prevents the virus from adapting to the receptors of the cells in order to easily enter into the cell (Sivanandy et al., 2022). These medicines have been critically shown to raise the survival rates to the higher degree. Furthermore, individualized supportive care is recognised to be significant in sustaining organ functions in the patients. This includes maintainance of adequate water and electrolyte balance by the use of oral or intravenous fluids, treatment of symptoms of the underlying cause and the use of analgesics for pain relief. Facilitating the fluid balance and providing intensive care to such patients can significantly improve the survival chances (Dhama et al., 2015).

8. Prevention

To prevent Ebola virus disease and its spread, people traveling to EVD-endemic areas should avoid contact with or consumption of wild animals. Those returning from such areas should monitor their body temperature for 21 days. Healthcare workers should use personal protective equipment (PPE) correctly when caring for suspected or confirmed Ebola patients. Blood examination should be done using safety needles. If exposed to blood or body fluids, the exposure site must be thoroughly cleaned with soap and water, and body temperature of the individual should be monitored for a duration of 21 days. Patients should be kept in a negative pressure chamber or a solitary room with its own facilities. Isolation can be removed after two negative test results within a 48-hour period. In the case of death from Ebola virus disease, bodies should be burned within 24 hours (Tseng & Chan, 2015). Strict implementation of disease control measures is crucial to prevent the transmission of the virus when infected.



Fig. 3: The pathogenesis of Ebola virus disease.

Conclusion

Ebola virus disease is a deadly zoonotic disease with a high mortality rate. The transmission and pathogenesis of the Ebola virus from bats to humans emphasize the complex interplay between wildlife and human health. Understanding the ecological dynamics that facilitate spillover events is crucial for predicting and preventing future outbreaks. The evidence linking bats as potential reservoirs highlights the need for targeted surveillance and research to identify and mitigate risks associated with zoonotic transmission. Continued investigation into the virus's behavior, host interactions, and immune evasion mechanisms will enhance our understanding of its pathogenesis and inform the development of effective therapeutic and preventive strategies. Awareness of these issues is crucial for managing future epidemics in underprivileged and remote areas where deadly infectious diseases such as Ebola could re-emerge. A multidisciplinary approach, integrating veterinary and public health efforts, is essential for safeguarding both human and animal population from threat posed by Ebola Virus.

References

Ahmad, Z., Din, N. U., Ahmad, A., Imran, S., Pervez, S., Ahmed, R., & Kayani, N. (2015). Rhabdomyosarcoma-An epidemiological and histopathologic study of 277 cases from a major tertiary care center in Karachi, Pakistan. Asian Pacific Journal of Cancer Prevention, 16(2), 757-760.

Asare Fenteng, E., Ossei, P. P. S., Ayibor, W. G., & Narh-Bedu, T. (2023). Beyond survival: unraveling the dynamics of Ebola virus resurgence in Sub-Saharan Africa and the remarkable journey of survivors. *Frontiers in Virology*, *3*, 1227314.

- Ascenzi, P., Bocedi, A., Heptonstall, J., Capobianchi, M. R., Di Caro, A., Mastrangelo, E., Bolognesi, M., & Ippolito, G. (2008). Ebolavirus and Marburgvirus: insight the Filoviridae family. *Molecular Aspects of Medicine*, 29(3), 151-185.
- Barrette, R. W., Metwally, S. A., Rowland, J. M., Xu, L., Zaki, S. R., Nichol, S. T., & McIntosh, M. T. (2009). Discovery of swine as a host for the Reston ebolavirus. *Science*, *325*(5937), 204-206.
- Basler, C. F. (2005). Interferon antagonists encoded by emerging RNA viruses. In *Modulation of host gene expression and innate immunity by viruses*, 197-220. Dordrecht: Springer Netherlands.
- Bausch, D. G., Towner, J. S., Dowell, S. F., Kaducu, F., Lukwiya, M., Sanchez, A., & Rollin, P. E. (2007). Assessment of the risk of Ebola virus transmission from bodily fluids and fomites. *The Journal of Infectious Diseases*, *196*(Supplement_2), S142-S147.
- Borio, L., Inglesby, T., Peters, C. J., Schmaljohn, A. L., Hughes, J. M., Jahrling, P. B., & Working Group on Civilian Biodefense. (2002). Hemorrhagic fever viruses as biological weapons: medical and public health management. *Jama*, *287*(18), 2391-2405.
- Bray, M. (2005). Pathogenesis of viral hemorrhagic fever. Current Opinion in Immunology, 17(4), 399-403.
- Bray, M., & Geisbert, T. W. (2005). Ebola virus: the role of macrophages and dendritic cells in the pathogenesis of Ebola hemorrhagic fever. The International Journal of Biochemistry & Cell Biology, 37(8), 1560-1566.
- Burk, R., Bollinger, L., Johnson, J. C., Wada, J., Radoshitzky, S. R., Palacios, G., Bavari, S., Jahrling, P. B., & Kuhn, J. H. (2016). Neglected filoviruses. *FEMS Microbiology Reviews*, 40(4), 494-519.
- Caron, A., Bourgarel, M., Cappelle, J., Liégeois, F., De Nys, H. M., & Roger, F. (2018). Ebola Virus Maintenance: If Not (Only) Bats, What Else?. Viruses, 10(10), 549.
- Chertow, D. S., Bray, M., & Palmore, T. N. (2022). Treatment and prevention of Ebola virus disease. UpToDate.
- Dean, N. E., Halloran, M. E., Yang, Y., & Longini, I. M. (2016). Transmissibility and pathogenicity of Ebola virus: a systematic review and metaanalysis of household secondary attack rate and asymptomatic infection. *Clinical Infectious Diseases*, *62*(10), 1277-1286.
- Dhama, K., Malik, Y. S., Malik, S. V. S., & Singh, R. K. (2015). Ebola from emergence to epidemic: the virus and the disease, global preparedness and perspectives. *The Journal of Infection in Developing Countries*, *9*(05), 441-455.
- Feldmann, F., & Feldmann, H. (2013). Ebola: facing a new transboundary animal disease?. In *Vaccines and Diagnostics for Transboundary Animal Diseases 135*, 201-209. Karger Publishers.
- Feldmann, H., & Geisbert, T. W. (2011). Ebola haemorrhagic fever. Lancet (London, England), 377(9768), 849-862.
- Funk, D. J., & Kumar, A. (2015). Ebola virus disease: an update for anesthesiologists and intensivists. *Canadian Journal of Anaesthesia*, 62(1), 80-91.
- Garske, T., Cori, A., Ariyarajah, A., Blake, I. M., Dorigatti, I., Eckmanns, T., & Donnelly, C. A. (2017). Heterogeneities in the case fatality ratio in the West African Ebola outbreak 2013–2016. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 372(1721), 20160308.
- Gebretadik, F. A., Seifu, M. F., & Gelaw, B. K. (2015). Review on Ebola virus disease: its outbreak and current status. *Epidemiology (sunnyvale)*, 5(4), 1000204.
- Geisbert, T. W., Young, H. A., Jahrling, P. B., Davis, K. J., Larsen, T., Kagan, E., & Hensley, L. E. (2003). Pathogenesis of Ebola hemorrhagic fever in primate models: evidence that hemorrhage is not a direct effect of virus-induced cytolysis of endothelial cells. *The American Journal of Pathology*, 163(6), 2371-2382.
- Ghosh, S., Saha, A., Samanta, S., & Saha, R. P. (2021). Genome structure and genetic diversity in the Ebola virus. *Current Opinion in Pharmacology*, 60, 83-90.
- Glynn, J. R., Bower, H., Johnson, S., Turay, C., Sesay, D., Mansaray, S. H., & Checchi, F. (2018). Variability in intrahousehold transmission of Ebola virus, and estimation of the household secondary attack rate. *The Journal of Infectious Diseases*, 217(2), 232-237.
- Goldstein, T., Anthony, S. J., Gbakima, A., Bird, B., Bangura, J., Tremeau-Bravard, A., & Mazet, J. (2019). The discovery of a new Ebolavirus, Bombali virus, adds further support for bats as hosts of Ebolaviruses. *International Journal of Infectious Diseases*, 79, 4-5.
- Green A. (2014). Ebola emergency meeting establishes new control centre. *Lancet (London, England)*, 384(9938), 118.
- Hussein H. A. (2023). Brief review on ebola virus disease and one health approach. *Heliyon*, *9*(8), 19036.
- Jain, S., Khaiboullina, S., Martynova, E., Morzunov, S., & Baranwal, M. (2023). Epidemiology of Ebolaviruses from an Etiological Perspective. *Pathogens (Basel, Switzerland)*, 12(2), 248.
- Judson, S., Prescott, J., & Munster, V. (2015). Understanding ebola virus transmission. Viruses, 7(2), 511-521.
- Kortepeter, M. G., Bausch, D. G., & Bray, M. (2011). Basic clinical and laboratory features of filoviral hemorrhagic fever. *The Journal of Infectious Diseases*, 204(3), S810-S816.
- Kreuels, B., Wichmann, D., Emmerich, P., Schmidt-Chanasit, J., de Heer, G., Kluge, S., & Schmiedel, S. (2014). A case of severe Ebola virus infection complicated by gram-negative septicemia. *New England Journal of Medicine*, *371*(25), 2394-2401.
- Laing, E. D., Mendenhall, I. H., Linster, M., Low, D. H., Chen, Y., Yan, L., & Smith, G. J. (2018). Serologic evidence of fruit bat exposure to filoviruses, Singapore, 2011–2016. *Emerging Infectious Diseases*, 24(1), 122.
- Leroy, E. M., Epelboin, A., Mondonge, V., Pourrut, X., Gonzalez, J. P., Muyembe-Tamfum, J. J., & Formenty, P. (2009). Human Ebola outbreak resulting from direct exposure to fruit bats in Luebo, Democratic Republic of Congo, 2007. Vector-borne and Zoonotic Diseases, 9(6), 723-728.
- Mahanty, S., & Bray, M. (2004). Pathogenesis of filoviral haemorrhagic fevers. The Lancet. Infectious Diseases, 4(8), 487-498.
- Majid, M. U., Tahir, M. S., Ali, Q., Rao, A. Q., Rashid, B., Ali, A., Nasir, I. A. & Husnain, T. (2016). Nature and history of Ebola virus: an overview. *Archives of Neuroscience*, 3(3).
- Malik, S., Kishore, S., Nag, S., Dhasmana, A., Preetam, S., Mitra, O., & Sah, R. (2023). Ebola virus disease vaccines: development, current perspectives & challenges. *Vaccines*, *11*(2), 268.
- Marí Saéz, A., Weiss, S., Nowak, K., Lapeyre, V., Zimmermann, F., Düx, A., & Leendertz, F. H. (2015). Investigating the zoonotic origin of the

West African Ebola epidemic. EMBO Molecular Medicine, 7(1), 17-23.

- Mbala-Kingebeni, P., Pratt, C., Mutafali-Ruffin, M., Pauthner, M. G., Bile, F., Nkuba-Ndaye, A., & Muyembe Tamfum, J. J. (2021). Ebola virus transmission initiated by relapse of systemic Ebola virus disease. *New England Journal of Medicine*, *384*(13), 1240-1247.
- Mendoza, E. J., Qiu, X., & Kobinger, G. P. (2016). Progression of Ebola Therapeutics during the 2014-2015 Outbreak. Trends in Molecular Medicine, 22(2), 164-173.
- Meyer, M., Huang, E., Yuzhakov, O., Ramanathan, P., Ciaramella, G., & Bukreyev, A. (2018). Modified mRNA-Based Vaccines Elicit Robust Immune Responses and Protect Guinea Pigs from Ebola Virus Disease. *The Journal of Infectious Diseases*, *217*(3), 451-455.
- Bray, M., Chertow, D. S., Hirsch, M. S., & Mitty, J. (2015). Epidemiology and pathogenesis of Ebola virus disease. UpToDate, Waltham, MA.
- Moghadam, S. R. J., Omidi, N., Bayrami, S., Moghadam, S. J., & SeyedAlinaghi, S. (2015). Ebola viral disease: a review literature. Asian Pacific Journal of Tropical Biomedicine, 5(4), 260-267.
- Munster, V. J., Bausch, D. G., De Wit, E., Fischer, R., Kobinger, G., Muñoz-Fontela, C., & Mombouli, J. V. (2018). Outbreaks in a rapidly changing Central Africa–lessons from Ebola. New England Journal of Medicine, 379(13), 1198-1201.
- Nkoghe, D., Formenty, P., Leroy, E. M., Nnegue, S., Edou, S. Y., Ba, J. I., & Mve, M. T. (2005). Multiple Ebola virus haemorrhagic fever outbreaks in Gabon, from October 2001 to April 2002. *Bulletin de la Societe de Pathologie Exotique (1990)*, *98*(3), 224-229.
- Nordenstedt, H., Bah, E. I., de la Vega, M. A., Barry, M., N'Faly, M., Barry, M., & Ingelbeen, B. (2016). Ebola virus in breast milk in an Ebola virus–positive mother with twin babies, Guinea, 2015. *Emerging Infectious Diseases*, *22*(4), 759.
- Osterholm, M. T., Moore, K. A., Kelley, N. S., Brosseau, L. M., Wong, G., Murphy, F. A., & Kobinger, G. P. (2015). Transmission of Ebola viruses: what we know and what we do not know. *MBio*, *6*(2), 10-1128.
- Pandey, S., Chauhan, V. D., Parmar, B. C., & Nayak, J. B. (2015). Ebola Disease: An Emerging Zoonosis. Journal of Foodborne and Zoonotic Diseases, 3(2), 19-22.
- Passi, D., Sharma, S., Dutta, S. R., Dudeja, P., & Sharma, V. (2015). Ebola virus disease (the killer virus): another threat to humans and bioterrorism: brief review and recent updates. *Journal of Clinical and Diagnostic Research*, *9*(6), LE01.
- Prasad, A. N., Fenton, K. A., Agans, K. N., Borisevich, V., Woolsey, C., Comer, J. E., & Geisbert, T. W. (2023). Pathogenesis of aerosolized Ebola virus variant Makona in nonhuman primates. *The Journal of Infectious Diseases*, 228(7), S604-S616.
- Reichler, M. R., Bangura, J., Bruden, D., Keimbe, C., Duffy, N., Thomas, H., & Hennessy, T. (2018). Household transmission of Ebola virus: risks and preventive factors, Freetown, Sierra Leone, 2015. *The Journal of Infectious Diseases*, 218(5), 757-767.
- Sanchez, A., Lukwiya, M., Bausch, D., Mahanty, S., Sanchez, A. J., Wagoner, K. D., & Rollin, P. E. (2004). Analysis of human peripheral blood samples from fatal and nonfatal cases of Ebola (Sudan) hemorrhagic fever: cellular responses, virus load, and nitric oxide levels. *Journal* of Virology, 78(19), 10370-10377.
- Singh, R. K., Dhama, K., Malik, Y. S., Ramakrishnan, M. A., Karthik, K., Khandia, R., & Joshi, S. K. (2017). Ebola virus–epidemiology, diagnosis, and control: threat to humans, lessons learnt, and preparedness plans–an update on its 40 year's journey. *Veterinary Quarterly*, *37*(1), 98-135.
- Sissoko, D., Keïta, M., Diallo, B., Aliabadi, N., Fitter, D. L., Dahl, B. A., & Duraffour, S. (2017). Ebola virus persistence in breast milk after no reported illness: a likely source of virus transmission from mother to child. *Clinical Infectious Diseases*, *64*(4), 513-516.
- Sivanandy, P., Jun, P. H., Man, L. W., Wei, N. S., Mun, N. F. K., Yii, C. A. J., & Ying, C. C. X. (2022). A systematic review of Ebola virus disease outbreaks and an analysis of the efficacy and safety of newer drugs approved for the treatment of Ebola virus disease by the US Food and Drug Administration from 2016 to 2020. *Journal of Infection and Public Health*, 15(3), 285-292.
- Tellier, R. (2009). Aerosol transmission of influenza A virus: a review of new studies. Journal of the Royal Society Interface, 6(6), S783-S790.
- Towner, J. S., Rollin, P. E., Bausch, D. G., Sanchez, A., Crary, S. M., Vincent, M., & Nichol, S. T. (2004). Rapid diagnosis of Ebola hemorrhagic fever by reverse transcription-PCR in an outbreak setting and assessment of patient viral load as a predictor of outcome. *Journal of Virology*, 78(8), 4330-4341.
- Tseng, C. P., & Chan, Y. J. (2015). Overview of Ebola virus disease in 2014. Journal of the Chinese Medical Association, 78(1), 51-55.
- Victory, K. R., Coronado, F., Ifono, S. O., Soropogui, T., Dahl, B. A., & Centers for Disease Control and Prevention (CDC). (2015). Ebola transmission linked to a single traditional funeral ceremony–Kissidougou, Guinea, December, 2014–January 2015. Morbidity and Mortality Weekly Report, 64(14), 386-8.
- Wambani, R. J., Ogola, P. E., Arika, W. M., Rachuonyo, H. O., & Burugu, M. W. (2016). Ebola virus disease: a biological and epidemiological perspective of a virulent virus. *Journal of Infectious Diseases and Diagnosis*, *1*(103), 2.
- Weingart, H. M., Nfon, C., & Kobinger, G. (2013). Review of Ebola virus infections in domestic animals. Vaccines and Diagnostics for Transboundary Animal Diseases, 135, 211-218.
- Xu, Z., Jin, B., Teng, G., Rong, Y., Sun, L., Zhang, J., & Chen, H. (2016). Epidemiologic characteristics, clinical manifestations, and risk factors of 139 patients with Ebola virus disease in western Sierra Leone. *American Journal of Infection Control*, 44(11), 1285-1290.
- Zheng, H., Yin, C., Hoang, T., He, R. L., Yang, J., & Yau, S. S. T. (2015). Ebolavirus classification based on natural vectors. DNA and Cell Biology, 34(6), 418-428.
- Zumbrun, E. E., Abdeltawab, N. F., Bloomfield, H. A., Chance, T. B., Nichols, D. K., Harrison, P. E., & Nalca, A. (2012). Development of a murine model for aerosolized ebolavirus infection using a panel of recombinant inbred mice. *Viruses*, 4(12), 3468-3493