Cryptosporidiosis: A Zoonotic Challenge in Waterborne Disease Management

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Abstract

Cryptosporidiosis is a zoonotic emerging disease caused by the protozoan parasite of *Cryptosporidium*. *It* is spread by food and drink and characterized by profuse watery diarrhea. Right now, there aren't effective medications or vaccinations on the market for the management and treatment of Cryptosporidiosis. We must learn more about the disease's transmission and spread, as well as how to break the cycle of transmission, to avoid cryptosporidiosis in both humans and animals. Understanding cryptosporidiosis, including its infectious stage, pathophysiology, life cycle, epidemiology, prior outbreaks, infection source, transmission, and high-risk groups, is the main goal of this chapter. The facts presented in this review will be useful in addressing upcoming human and animal *Cryptosporidium* infections and lowering the incidence of the disease.

Keywords: Emerging protozoan, Gastrointestinal infection, Cryptosporidiosis, Epidemiology, Outbreaks.

Cite this Article as: Ismael SS and Alnakshabandi WMY, 2025. Cryptosporidiosis: A zoonotic challenge in waterborne disease management. In: Kun Li (ed), Protozoan Zoonoses: Advances in the Diagnosis, Prevention, and Treatment of Cryptosporidiosis and Toxoplasmosis. Unique Scientific Publishers, Faisalabad, Pakistan, pp: 9-15. <u>https://doi.org/10.47278/book.HH/2025.279</u>



A Publication of Unique Scientific Publishers Chapter No: 25-222 **Received:** 21-Feb-2025 **Revised:** 12-Apr-2025 **Accepted:** 15-May-2025

Introduction

Cryptosporidiosis is the most common and prevalent waterborne parasitic and zoonotic disease caused by a coccidian protozoon parasite *Cryptosporidium* (Helmy et al., 2017). *Cryptosporidium* spp. is considered the major source of waterborne parasitic infection outbreaks globally (Gharpure et al., 2019). About fifty-eight million cases of diarrhea among children are identified per year, and these occurrences are linked to protozoal infections. In particular, the World Health Organization's "Neglected Disease Initiative" involved waterborne diseases like *Giardia* and *Cryptosporidium* (Savioli et al., 2008). In immunocompetent people, Cryptosporidium infections cause acute, self-limiting gastroenteritis, but in immunocompromised people, they can cause chronic, sometimes fatal diarrheal illness. Because of their developing immune systems, neonates are particularly vulnerable to infections, which can be contracted by ingesting small amounts of the parasite's oocysts (Dadonaite et al., 2019). *Cryptosporidium* is responsible for children's diarrhea in developing countries, and the disease is more potent in immunocompromised patients (Mosier & Oberst, 2000). The final host is infected with cryptosporidiosis by ingestion of sporulated oocysts with contaminated foods, vegetables, grass, or drinking of contaminated water, or through direct contact with infected animals or people (CDC, 2017). This chapter outlined cryptosporidiosis, the most important species that infect humans, its life cycle, and sources of infection, clinical manifestation, diagnosis, and disease prevention.

1.1. Causative Agent

Cryptosporidium species are intracellular coccidian parasites under the phylum Sporozoa (Apicomplexa) (Fayer et al., 2000, Hijjawi et al., 2002, Pal et al., 2016). The pathogenic human species responsible for disease and outbreaks are *C. parvum*, *C. cuniculus*, *C. hominis, and C. meleagridis*, *C. parvum* is considered the most common and pathogenic. Species that infect animals are *C. andersoni*, *C. baileyi*, *C. ducismarci*, *C. fragile*, *C. galli*, *C. meleagridis*, *C. parvum*, *C. ubiquitum*, *C. serpentis*, *C. varanii*, and *C. xiaoi* (Gaibova et al., 2017). It is commonly reported that the most significant species, *C. parvum* can infect both humans and animals (Pal et al., 2007, Mor et al., 2018). At this time, about 60 distinct genotypes of *Cryptosporidium* with varying molecular sequences have been identified. Subtypes of *Cryptosporidium* are identified by the number of repeats in each strand. In certain subtypes, the trinucleotide repeats are followed immediately by short, repetitive sequences (R). Eleven subtype families (IIa–IIk) with a minimum of 78 subtypes have been identified in *C. parvum* (Fayer, 2008, Ahmed & Karanis, 2020).

1.2. Transmission of Cryptosporidiosis

Cryptosporidium oocysts are released from stools, that's accidentally consumed and can spread directly from animal to animal, from animal to human, or from human to animal (Desai et al., 2012; Ismael et al., 2024). Among the indirect transmissions are cross-contamination of food items, food components, drinking water, and other clothes and footwear items used in livestock farms or wildlife parks that have come into contact with an infected person's or animal's excrement. It can live and infect the intestinal epithelium in humans and many vertebrate animals. Also, spreads from the stool into the surrounding environment through soil and water sources like rivers, ponds, sewage, slurry, and wastewater, as well as through many water containers, especially public water supplies that are not properly treated (Hubalek, 2003, Efstratiou et al., 2017, Tomazic et al., 2018). Cryptosporidiosis outbreaks have been documented in both industrialized and developing nations (Shirley et al., 2012, Ursini et al., 2020). These outbreaks are primarily caused by the virus's low infectious dose, diverse animal reservoirs, and resistance to chlorination (Bouzid et al., 2013, Adeyemo et al., 2019).

1.3 Life cycle of Cryptosporidiosis

Humans are infected by consuming sporulated oocysts. After ingestion of sporulated oocysts, in the small intestine, the excystation occurs and the four sporozoites are released and invade the rims of the intestinal epithelial cells (Tzipori & Ward, 2002, Tzipori & Ward, 2008). Asexual reproduction began and two times schizogony occurs (Chalmers et al., 2011), the result of the first schizogony is eight merozoites are released and invade a healthy epithelial cell and the second schizogony takes place and form four merozoites (Bouzid et al., 2013). Then merozoites are released and invade another intestinal cell and sexual reproduction begins followed by the forming of male and female gametes, zygote, ookinete, and oocyst, two types of oocysts are produced thin-walled and thick-walled oocysts, the thin-walled type is responsible for the autoinfection and the thick-walled is a pass with stool and considered the source for infection as seen in Figure 1 (Jenkins et al., 2010, Khan et al., 2019).

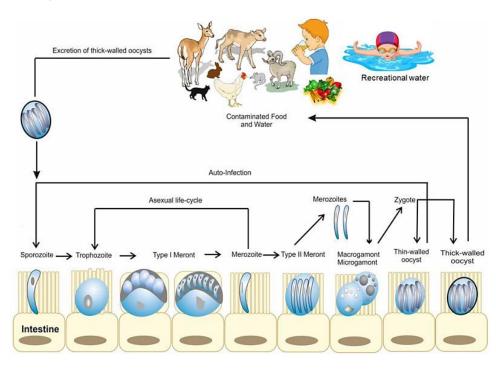


Fig. 1: Cryptosporidiosis life cycle (Etzold et al., 2014)

2.4. Prevalence of Cryptosporidiosis

An estimated 30 to 50 percent of deaths in young people worldwide are attributed to *Cryptosporidium* infections, which are also the second leading cause of diarrhea and pediatric mortality after rotavirus (Striepen, 2013). There have been 325 documented cases of parasitic protozoan disease up to 2007, with *C. parvum* accounting for 50.8% of these cases, with 93% of these cases occurring in North American and European nations. Of these, 30 percent of outbreaks were reported to have originated in Europe, with the UK responsible for 24 percent of all outbreaks worldwide (Karanis et al., 2007) Australia was the continent where Cryptosporidium outbreaks were most common, followed by North America and Europe (Baldursson & Karanis, 2011). North America accounted for 36% of the total number of publications, then followed by the European Union (31%), Asia (19%), South and Central America (7%), Oceania (5%), and Africa (3%), according to a recent study that demonstrated the scientific motivation provided to the surveillance of parasitic protozoan illness to fifty-three nations (1970 to 2016) (Efstratiou et al., 2017).

According to an epidemiological study from three African and three Asian sites, *Cryptosporidium* spp. was the second most common parasite that caused severe diarrhea and substantial morbidity in children aged 12 to 23 months (Kotloff et al., 2013). It is predicted that more than 2.9 million children under the age of 24 months in sub-Saharan Africa contract Cryptosporidium each year (Squire & Ryan, 2017). Sixteen water-borne protozoa outbreaks were documented in Latin American nations between 1979 and 2015; the most prevalent protozoa in these outbreaks were Cryptosporidium spp. (Rosado-García et al., 2017). Another study was done by Khan et al. (2019), which found that the primary source of infection in children was *C. parvum*. The findings showed that people who had diarrhea had a higher chance of contracting *Cryptosporidium* and that several environmental factors might potentially be important in the parasite's transmission.

The prevalence of cryptosporidiosis in Iraq differed from city to city and from year. The prevalence of infection in Iraq from 2002 to 2020. In 2002, the prevalence of infection among adults was 100% in Basrah City (Mahdi & Ali, 2002), was high in Diwaniya City at 35.74% (Mahdi and Ali, 2004), in Mosul City (Mukhtar and Sherefat, 2005), in Kut City, Iraq, a high prevalence was reported among children less than twelve years old at 33.99% (Rahi *et al.*, 2013), in Kirkuk City was low at 6.7% (Salman *et al.*, 2016), in Erbil City among pediatrics was 10.77% (Kanabe & Darogha, 2017). In Duhok City, a low prevalence rate was reported by Hussein & Meerkhan (2019), and a high prevalence rate was reported in both Baghdad City (Whaeeb *et al.*, 2020) and in Duhok City (Al-Saeed and Abdo, 2020), which were 44.5% and 67.0%, respectively; finally, in Duhok City, it was as high as usual in 2024, at 17.1% (Ismael *et al.*, 2024). Lack of knowledge and regular testing to identify this parasite may be the cause of the high prevalence of Cryptosporidiosis. Also, since many people are affected and there are related socioeconomic consequences, the effects of waterborne epidemics are rather substantial, and the oocysts are extremely resistant to both biological and chemical threats (Khan et al., 2019).

The economic and health consequences of cryptosporidiosis outbreaks linked to polluted water sources can be severe. An estimated 96.2 million Dollars was spent on the massive waterborne outbreak in Wisconsin, which impacted 403,000 people (Corso et al., 2003). According to estimates, a waterborne cryptosporidiosis outbreak in Sweden, where the attack rate was 45% of the country's 60, 000 inhabitants, resulted in 50,000 sick leave days (Ridderstedt et al., 2018). Public water supplies are frequently condemned, water boil notices are posted, and bottled water is provided when Cryptosporidium oocysts are found in them. Over 120,432 persons were impacted by a boil water order that was in effect for 158 days during a recent waterborne outbreak in Ireland, which cost over 19 million EUR (Chyzheuskaya et al., 2017).

2.5. Clinical Manifestations and Risked Population

In both immunocompetent and immunodeficient individuals, *Cryptosporidium* has been identified as a cause of gastrointestinal disorders (Rossle & Latif, 2013). Cryptosporidiosis is characterized by severe diarrhea that can sometimes be fatal. Watery diarrhea and dehydration are the most severe symptoms in immunocompromised individuals, but symptoms in immunocompetent individuals typically resolve on their own. Additionally, AIDS may be lethal for immunocompromised patients (Pal, 2007, Chukwuma, 2019).

Risk populations are poor nations, immunocompromised patients, and children. Due to a lack of resources and knowledge about illness prevention and control, the disease is more common in developing nations than in developed ones. Although cryptosporidiosis is rare in immunocompetent individuals, it causes 10% to 15% of cases of severe diarrhea in underdeveloped nations, especially in undernourished children under five. Outbreaks of cryptosporidiosis have been linked to tainted drinking water or swimming pools in several countries (Lebbad et al., 2021). People who are immunocompromised due to infection with HIV are more likely to develop Cryptosporidiosis, and most of the time, having the parasite is linked to diarrheal illness (Pozio and Morales, 2005). Malignant neoplasms, organ transplants, and primary immunodeficiency disorders increased the possibility of infection in these individuals; one and multiple species of *Cryptosporidium* were found in these patients (Hunter and Nichols, 2002).

1.6. Diagnosis

There are several techniques used for diagnosis as mentioned in Table 1. Cryptosporidiosis can be directly detected in fecal samples using a variety of techniques, such as microscopy identification of the oocysts, which counts the number of oocysts in the stools using sedimentation or flotation methods (Under a microscope, the detection of oocysts limit has been recorded to be as low as 50,000 to 500,000 oocysts per gram of stool). The most common methods for directly detecting *Cryptosporidium* oocysts are either unstaining microscopy or the modified Ziehl-Neelsen stain, in which the oocysts are colored purple on a blue backdrop (Chartier et al., 2013). Monoclonal antibodies that react with the oocyst wall antigen are also frequently utilized in immunofluorescent antibody-based (IFA) staining procedures. Compared to other conventional staining techniques, these are less expensive and have a high sensitivity (Garcia et al., 1983, Cacciò & Widmer, 2013).

The most effective techniques for screening a large number of samples, especially in epidemiological surveys, are thought to be serological approaches. Enzyme-linked immunoelectron transfer blots and enzyme-linked immunosorbent assays (ELISA) are among the serological testing. In comparison to immunofluorescence approaches, enzyme immunoassay techniques are more sensitive, quicker, simpler, and less expensive (Fayer & Xiao, 2007). It is also possible to utilize rapid immunochromatographic tests (Helmy et al., 2014, El-Missiry *et al.*, 2019), which use monoclonal antibodies to detect the proteins in the oocyst cell wall (Papini & Cardini, 2006).

Diagnostic Technique	Description	References	
Microscopy	Utilizes modified Ziehl-Neelsen staining to identify oocysts in stool samples.	Omoruyi et al., 2014	
Immunochromatography	Rapid test detecting Cryptosporidium antigens in fecal samples.	Khurana & Chaudhary, 2018	
Enzyme-Linked	Detects antigens in stool samples, offering higher sensitivity than microscopy.	Barwari & Ismael, 2011;	
Immunosorbent Assay (ELISA)	Abdou et al., 2022	
Polymerase Chain	Molecular method detecting Cryptosporidium DNA in various samples, including Costa et al., 2021, Haji &		
Reaction (PCR)	stool and biopsy.	Ismael, 2023	
Loop-Mediated Isothermal Simple and specific molecular technique for detecting Cryptosporidium DNA in Karakavuk et al., 2014			
Amplification (LAMP)	environmental samples.		
Histological Examination	Involves using stains like haematoxylin and eosin on biopsy samples to identify Khurana & Chaudhary, 2018		
	oocysts.		
Electron Microscopy	Direct visualization of oocysts; historically used but less common due to high	h Khurana & Chaudhary, 2018	
	costs and complexity.		

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1.7. Prevention of Cryptosporidiosis

The primary preventive measures include drinking water that has been treated, eating cooked meals when visiting places with contaminated food supplies, disinfecting swimming pools, preventing patients with diarrhea from swimming, washing hands after using restrooms or changing children's diapers, and using protection techniques for safe sex as shown in Figure 2. (Manyazewal et al., 2018, Firouzivand et al., 2021). Given the difficulty of treating cryptosporidiosis, lowering the oocyst count may help to lessen the infection's severity and enable young animals and children to build immunity through appropriate colostrum feeding. More care must be taken with personal hygiene when handling animals and children who have diarrhea (Innes et al., 2020). Immunocompromised individuals and young children are not recommended to handle animals that are diarrheal (Pal, 2007).

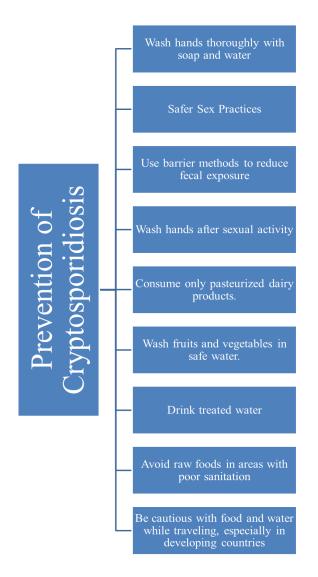


Fig. 2: Illustrated the prevention of Cryptosporidiosis

1.8. Treatment

According to Lima et al. (2011), the optimal treatment for cryptosporidiosis includes adequate hydration and electrolyte maintenance, anti-motility medicines, anti-parasitic medications, nutritional support, and immunosuppressive reversal. Nitazoxanide, on the other hand, may reduce the risk of death in malnourished infants and presumably reduce the severity of diarrhea (Amadi et al., 2002). Its effectiveness has also been verified among adults according to Rossignol et al. (2006). Also, it is important to treat infected domestic animals, due to zoonotic species (Innes et al., 2020).

Effective therapies for cryptosporidiosis were not found during the initial evaluation of the substances that were accessible. Several medications that were formerly thought to be useful have not worked in clinical studies (Cabada & White, 2010). Nitazoxanide is the only medication approved for the treatment of Cryptosporidium. In the 1980s, nitazoxanide was created by joining a thiazole ring. Nitazoxanide is a broad-spectrum anti-parasitic that has been used in controlled trials for giardiasis and cryptosporidiosis, as well as a deworming agent (Rossignol et al., 2001, Bailey & Erramouspe, 2004). Other medications such as bovine anti-cryptosporidium immunoglobulin, spiramycin, and azithromycin have also been shown to have some effect in case series (Cabada & White, 2010). Children with cryptosporidiosis appeared to respond more effectively to azithromycin than to two anthelminthic medications (Allam & Shehab, 2002). Additionally, it is frequently utilized in combination with paromomycin/nitazoxanide in weakened hosts, as some individuals have reduced parasite clearance and stool frequency (Legrand et al., 2011). In HIV/AIDS patients, paromomycin, which some anti-Cryptosporidium efficacy in immunocompetent individuals, is likewise ineffective. Restoring immunological conditions by a combination of antiretroviral therapy is necessary for symptom resolution. For patients with cryptosporidiosis, combination antiretroviral and antiparasitic therapy appears to be beneficial, particularly when used with a protease inhibitor-based treatment that appears to have some extra anti-parasitic action (Cabada & White, 2010).

Nanoparticles (NPs) have been generating a lot of attention lately in the fields of drug delivery and therapy (De Jong & Borm, 2008). The invention of nanoparticles into intelligent coating systems for therapeutic substances enables controlled release medication delivery to the intended locations. Furthermore, this targeted delivery increases patient compliance with lower dose frequency and lessens therapeutic side effects

(Rizvi & Saleh, 2018). The FDA of the United States has approved chitosan nanoparticles a naturally occurring cationic polysaccharide that is mucoadhesive and biocompatible, for use in medication delivery (Sorlier et al., 2001, Moawad et al., 2021). *In vitro*, it was established that chitosan nanoparticles conjugated with polyvinyl alcohol inhibit the attachment of *Cryptosporidium* sporozoites to enterocytes in cases of cryptosporidiosis (Luzardo Álvarez et al., 2012, Ahmed et al. 2019).

Conclusion

Cryptosporidium infections are prevalent in both people and animals. The two categories most at risk for infection are immunocompromised people and children under twelve years. The majority of *Cryptosporidium*-related foodborne outbreaks are zoonotic. Regular surveillance systems and the implementation of the One Health approach are necessary to stop disease outbreaks. In order to stop and/or lessen future epidemics globally, food safety and water cleanliness are essential. Considering the sources of infection (people and animals), the transmission pathways, the environmental survival of oocysts, and the risk factors can help prevent and control *Cryptosporidium* infections. There are currently no effective medications or vaccinations to cure or prevent infection in people or animals.

References

Abdou, N. M. I., Al-Azemi, M. S., Al-Sayegh, M. T., & Majeed, Q. A. H. (2022). Performance of diagnostic assays used to detect Cryptosporidium oocysts in faecal samples of cattle in Kuwait and genotyping of Cryptosporidium species. *BMC Veterinary Research*, 18(1), 336. https://doi.org/10.1186/s12917-022-03435-w

Abdulsadah A. Rahi, Magda A. Ali, Alaa H. Al-Charrakh. (2013). Prevalence of *Cryptosporidium parvum* among Children in Iraq. *American Journal of Life Sciences*, 1(6), 256-260. https://doi.org/10.11648/j.ajls.20130106.13

Adeyemo, F. E., Singh, G., Reddy, P., Bux, F., & Stenström, T. A. (2019). Efficiency of chlorine and UV in the inactivation of Cryptosporidium

and Giardia in wastewater. PloS oneOne, 14(5), e0216040. https://doi.org/10.1371/journal.pone.0216040

- Ahmed, S. A., & Karanis, P. (2020). Cryptosporidium and Cryptosporidiosis: The Perspective from the Gulf Countries. International journal Journal of Environmental Research and Public Health, 17(18), 6824. https://doi.org/10.3390/ijerph17186824
- Ahmed, S. A., El-Mahallawy, H. S., & Karanis, P. (2019). Inhibitory activity of chitosan nanoparticles against Cryptosporidium parvum oocysts. *Parasitology Research*, *118*(7), 2053–2063. https://doi.org/10.1007/s00436-019-06364-0
- Allam, A. F., & Shehab, A. Y. (2002). Efficacy of azithromycin, praziquantel and mirazid in treatment of cryptosporidiosis in school children. *Journal of the Egyptian Society of Parasitology*, 32(3), 969–978.
- Al-Saeed, A.T., Abdo, J.M., & Gorgess, R.G. (2020). Cryptosporidiosis in children in Duhok City / Kurdistan Region / Iraq. Journal of the Pakistan Medical AssociationJ. Pakistan Medicine Assoc., 70(7), 1251-1255.
- Amadi, B., Mwiya, M., Musuku, J., et al. Watuka, A., Sianongo, S., Ayoub, A., & Kelly, P. (2002). Effect of nitazoxanide on morbidity and mortality in Zambian children with cryptosporidiosis: a randomised controlled trial. *Lancet*, 360(9343), 1375–1380. https://doi.org/10.1016/S0140-6736(02)11401-2
- Bailey, J. M., & Erramouspe, J. (2004). Nitazoxanide treatment for giardiasis and cryptosporidiosis in children. *The Annals of Pharmacotherapy*, 38(4), 634–640. https://doi.org/10.1345/aph.1D451
- Baldursson, S., & Karanis, P. (2011). Waterborne transmission of protozoan parasites: review of worldwide outbreaks an update 2004-2010. *Water Research*, 45(20), 6603–6614. https://doi.org/10.1016/j.watres.2011.10.013
- Barwari, W. J., & Ismael, S. S. (2011). Detection of Pathogenic Strains of Entamoeba Histolytica in Children Using ELISA Technique in Duhok. *Journal of the University of Duhok, 8*(2), 109-119.
- Bouzid, M., Hunter, P. R., Chalmers, R. M., & Tyler, K. M. (2013). Cryptosporidium pathogenicity and virulence. *Clinical Microbiology Reviews*, 26(1), 115–134. https://doi.org/10.1128/CMR.00076-12
- Cabada, M. M., & White, A. C., Jr (2010). Treatment of cryptosporidiosis: do we know what we think we know?.know? *Current Opinion in Infectious Diseases*, 23(5), 494-499. https://doi.org/10.1097/QCO.ob013e32833de052
- Cacciò, S. M., & Widmer, G. (Eds.). (2013). Cryptosporidium: parasite and disease. Springer Science & Business Media. https://doi.org/10.1007/978-3-7091-1562-6
- Center for Disease Control and Prevention (CDC). (2017). Cryptosporidiosis Outbreaks United States, 2009–2017. https://www.cdc.gov/mmwr/volumes/68/wr/mm6825a3.htm
- Chalmers, R. M., Smith, R., Elwin, K., Clifton-Hadley, F. A., & Giles, M. (2011). Epidemiology of anthroponotic and zoonotic human cryptosporidiosis in England and Wales, 2004-2006. *Epidemiology and Infection*, 139(5), 700–712. https://doi.org/10.1017/S0950268810001688
- Chartier, C., Rieux, A., Delafosse, A., Lehebel, A., & Paraud, C. (2013). Detection of Cryptosporidium oocysts in fresh calf faeces: characteristics of two simple tests and evaluation of a semi-quantitative approach. *Veterinary Journal (London, England: 1997)*, *198*(1), 148–152. Https://doi.org/10.1016/j.tvjl.2013.06.011
- Chukwuma, C. (2019). Cryptosporidium: Public health problems and environmental indicators. Journal of Austin Medical Science, 4,1036.
- Chyzheuskaya, A., Cormican, M., Srivinas, R., et al. O'Donovan, D., Prendergast, M., O'Donoghue, C., & Morris, D. (2017). Economic Assessment of Waterborne Outbreak of Cryptosporidiosis. *Emerging Infectious Diseases*, 23(10), 1650–1656. https://doi.org/10.3201/eid2310.152037
- Corso, P. S., Kramer, M. H., Blair, K. A., Addiss, D. G., Davis, J. P., & Haddix, A. C. (2003). Cost of illness in the 1993 waterborne Cryptosporidium outbreak, Milwaukee, Wisconsin. *Emerging infectious Infectious diseasesDiseases*, *9*(4), 426–431. https://doi.org/10.3201/eid0904.020417
- Costa, D., Soulieux, L., Razakandrainibe, R., Basmaciyan, L., Gargala, G., Valot, S., Dalle, F., & Favennec, L. (2021). Comparative Performance of Eight PCR Methods to Detect Cryptosporidium Species. *Pathogens*, 10(6), 647. https://doi.org/10.3390/pathogens10060647
- Dadonaite, B., Ritchie, H., & Roser, M. (2019). Diarrheal Diseases. In Our World in Data. Current Urban Studies. 9, :3.
- De Jong, W. H., & Borm, P. J. (2008). Drug delivery and nanoparticles:applications and hazards. *International Journal of Nanomedicine*, 3(2), 133–149. https://doi.org/10.2147/ijn.s596
- De Jong, W. H., & Borm, P. J. (2008). Drug delivery and nanoparticles: applications and hazards. *International Journal of Nanomedicine*, 3(2), 133–149. https://doi.org/10.2147/ijn.s596
- Desai, N. T., Sarkar, R., & Kang, G. (2012). Cryptosporidiosis: An under-recognized public health problem. *Tropical Parasitology*, 2(2), 91–98. https://doi.org/10.4103/2229-5070.105173
- Efstratiou, A., Ongerth, J., & Karanis, P. (2017). Evolution of monitoring for Giardia and Cryptosporidium in water. *Water Research*, *123*, 96–112. https://doi.org/10.1016/j.watres.2017.06.042
- El-Adawy, Y. A., & Abdelwhab, E. M. (2017). A Comprehensive Review of Common Bacterial, Parasitic and Viral Zoonoses at the Human-Animal Interface in Egypt. *Pathogens (Basel, Switzerland)*, 6(3), 33. https://doi.org/10.3390/pathogens6030033
- El-Missiry A., Abd El-Hameed L., Saad G., El-Badry A., Helmy Y., & Shehata M. (2019). Molecular genetic characterization of human Cryptosporidium isolates and their respective demographic, environmental and clinical manifestations in Egyptian diarrheic patients. *Parasitologists United Journal*, 12,187–196.
- Etzold, M., Lendner, M., Daugschies, A., & Dyachenko, V. (2014). CDPKs of *Cryptosporidium* parvum--stage-specific expression in vitro. *Parasitology Research*, *11*3(7), 2525–2533. https://doi.org/10.1007/s00436-014-3902-0
- Fayer R., & Xiao L. (2007). Cryptosporidium and Cryptosporidiosis. (2nd ed). CRC Press; Boca Raton, FL, USA: Taylor & Francis Group; pp. 580.
- Fayer, R. (2008). General Biology. In: Fayer R., Xiao L., (Eds.), itors. *Cryptosporidium and Cryptosporidiosis*. CRC Press; Boca Raton, FL, USA: 2008. pp. 1–42

- Fayer, R., Morgan, U., & Upton, S. J. (2000). Epidemiology of *Cryptosporidium:* transmission, detection and identification. *International Journal of Parasitology*, 30(12-13), 1305–1322. https://doi.org/10.1016/s0020-7519(00)00135-1
- Firouzivand, Y., Garedagh, Y, Hassanzadeh Khanmiri H. (2021). Prevalence of *Cryptosporidium* in children under 13 years of age with acute diarrhea in Tabriz, Iran. *International Journal of Medical Parasitology and Epidemiology Sciences, International Journal Medicine Parasitology Epidemiolgical Science*, 2(1): 19-22. http://dx.doi.org/10.34172/ijmpes.2021.05
- Gaibova, H., Iskenderova, N., & Gurbanova, T. (2017). Review of the modern state of the emeridcoccides of terrestrial vertebrates of Azerbaijan. In: Proceedings of the XIX International Scientific Conference with Elements of the Scientific School of Young Scientists. Makhachkala, Russia: Biological diversity Caucasus and South Russia, pp. 423-5.
- Gharpure, R., Perez, A., Miller, A. D., et al. Wikswo, M. E., Silver, R., & Hlavsa, M. C. (2019). Cryptosporidiosis Outbreaks United States, 2009-2017. MMWR. Morbidity and Mortality Weekly Rreport, 68(25), 568–572. https://doi.org/10.15585/mmwr.mm6825a3
- Haji, H. M., & Ismael, S. S. (2023). Prevalence of Entamoeba histolytica and Giardia lamblia in Children in Duhok Province, Kurdistan Region, Iraq. Journal of Duhok University, 26(1), 274-280.
- Helmy, Y. A., Krücken, J., Nöckler, K., et al. von Samson-Himmelstjerna, G., & Zessin, K. H. (2014). Comparison between two commercially available serological tests and polymerase chain reaction in the diagnosis of *Cryptosporidium* in animals and diarrhoeicdiarrheic children. *Parasitology Research*, 113(1), 211–216. https://doi.org/10.1007/s00436-013-3645-3
- Hijjawi, N. S., Meloni, B. P., Ryan, U. M., Olson, M. E., & Thompson, R. C. (2002). Successful in vitro cultivation of Cryptosporidium andersoni: evidence for the existence of novel extracellular stages in the life cycle and implications for the classification of *Cryptosporidium. International Journal for Parasitology*, 32(14), 1719–1726. https://doi.org/10.1016/s0020-7519(02)00199-6
- Hubálek Z. (2003). Emerging human infectious diseases: anthroponoses, zoonoses, and sapronoses. *Emerging Infectious Diseases*, *9*(3), 403–404. https://doi.org/10.3201/eido903.020208
- Hunter, P. R., & Nichols, G. (2002). Epidemiology and clinical features of Cryptosporidium infection in immunocompromised patients. *Clinical Microbiology Reviews*, 15(1), 145–154. https://doi.org/10.1128/CMR.15.1.145-154.2002
- Hussein, J. N., & Meerkhan, A. A. (2019). The Incidence of Intestinal Parasites among Children in Hivi Pediatric Hospital, Duhok, Iraq. Science Journal of University of Zakho, 7(1), 1–4. https://doi.org/10.25271/sjuoz.2019.7.1.571
- Innes, E. A., Chalmers, R. M., Wells, B., & Pawlowic, M. C. (2020). A One Health Approach to Tackle Cryptosporidiosis. *Trends in Parasitology*, 36(3), 290-303. https://doi.org/10.1016/j.pt.2019.12.016
- Ismael, S., Abdullah, B. H., Sadiq, A. J., Ajaj, J. S., Ali, N. S., Omer, D. M., & Nori, N. Y. (2024). Prevalence of Intestinal Protozoan Parasites among Children attending the Hevi Pediatric Hospital in Duhok Province, Kurdistan Region, Iraq. Archives of Razi Institute, 79(3), 507-512.
- Jenkins, M. B., Eaglesham, B. S., Anthony, L. C., Kachlany, S. C., Bowman, D. D., & Ghiorse, W. C. (2010). Significance of wall structure, macromolecular composition, and surface polymers to the survival and transport of Cryptosporidium parvum oocysts. Applied and Environmental Microbiology, 76(6), 1926–1934. https://doi.org/10.1128/AEM.02295-09
- Kanabe, L.O., Darogha, S.N.R. (2017). Epidemiology of Cryptosporidiosis among diarrheic children of Raparin Pediatric hospital, Erbil Province-Kurdistan Region, Iraq. *Cihan University-Erbil Scientific Journal*, 2, 538
- Karakavuk, M., Can, H., Can, Ş., Karakavuk, T., Döşkaya, M., & Değirmenci Döşkaya, A. (2024). Development of a "Rapid-Crypto Colorimetric LAMP Test" to Detect Cryptosporidiosis in Feces of Newborns Calves. Acta Parasitologica, 69(1), 691–699. https://doi.org/10.1007/s11686-023-00791-x
- Karanis, P., Kourenti, C., & Smith, H. (2007). Waterborne transmission of protozoan parasites: a worldwide review of outbreaks and lessons learnt. *Journal of Water and Health*, 5(1), 1–38. https://doi.org/10.2166/wh.2006.002
- Khan, A., Shams, S., Khan, S., Khan, M. I., Khan, S., & Ali, A. (2019). Evaluation of prevalence and risk factors associated with Cryptosporidium infection in rural population of district Buner, Pakistan. *PloS One*, *14*(1), e0209188. https://doi.org/10.1371/journal.pone.0209188
- Khurana, S., & Chaudhary, P. (2018). Laboratory diagnosis of cryptosporidiosis. *Tropical Parasitology*, 8(1), 2–7. https://doi.org/10.4103/tp.TP_34_17
- Kotloff, K. L., Nataro, J. P., Blackwelder, W. C., et al. Nasrin, D., Farag, T. H., Panchalingam, S., Wu, Y., Sow, S. O., Sur, D., Breiman, R. F., Faruque, A. S., Zaidi, A. K., Saha, D., Alonso, P. L., Tamboura, B., Sanogo, D., Onwuchekwa, U., Manna, B., Ramamurthy, T., Kanungo, S., ... Levine, M. M. (2013). Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study. *Lancet (London, England)*, 382(9888), 209–222. https://doi.org/10.1016/S0140-6736(13)60844-2
- Lebbad, M., Winiecka-Krusnell, J., Stensvold, C. R., & Beser, J. (2021). High Diversity of *Cryptosporidium* Species and Subtypes Identified in Cryptosporidiosis Acquired in Sweden and Abroad. *Pathogens*, 10(5), 523. https://doi.org/10.3390/pathogens10050523
- Legrand, F., Grenouillet, F., Larosa, F., Dalle, F., Saas, P., Millon, L., Deconinck, E., & Rohrlich, P. S. (2011). Diagnosis and treatment of digestive cryptosporidiosis in allogeneic haematopoietic stem cell transplant recipients: a prospective single centre study. *Bone Marrow Transplantation*, 46(6), 858–862. https://doi.org/10.1038/bmt.2010.200
- Lima, A.A.M., Samie, A., & Guerrant, R.L. (2011). Cryptosporidiosis. In: Guerrant RL, Walker DH, Weller PF, eds. Tropical Infectious Diseases. Philadelphia, Pa: *Elsevier-Churchill Livingstone*, 640-663.
- Luzardo Álvarez, A., Blanco García, E., et alGuerrero Callejas, F., Gómez Couso, H., & Blanco Méndez, J. (2012). In vitro evaluation of the suppressive effect of chitosan/poly (vinyl alcohol) microspheres on attachment of C. parvum to enterocytic cells. *European journal Journal* of pharmaceutical Pharmaceutical sciencesSciences: official Journal of the European Federation for Pharmaceutical Sciences, 47(1), 215– 227. https://doi.org/10.1016/j.ejps.2012.06.002
- Mahdi, N. K., & Ali, N. H. (2002). Intestinal parasites, including Cryptosporidium species, in Iraqi patients with sickle-cell anaemia. Eastern

Mediterranean Health Journal, 8(2-3), 345-349.

- Mahdi, N. K., & Ali, N. H. (2004). Cryptosporidiosis and other intestinal parasitic infections in patients with chronic diarrhea. Saudi Medical Journal, 25(9), 1204–1207.
- Manyazewal, A., Francesca, S., Pal, M., et al. Gezahegn, M., Tesfaye, M., Lucy, M., Teklu, W., & Getachew, T. (2018). Prevalence, risk factors and molecular characterization of Cryptosporidium infection in cattle in Addis Ababa and its environs, Ethiopia. Veterinary Parasitology, Regional Studies and Reports, 13, 79–84. https://doi.org/10.1016/j.vprsr.2018.03.005
- Moawad, H. S. F., Hegab, M. H. A. E., Badawey, M. S. R., Ashoush, S. E., Ibrahim, S. M., & Ali, A. A. E. S. (2021). Assessment of chitosan nanoparticles in improving the efficacy of nitazoxanide on cryptosporidiosis in immunosuppressed and immunocompetent murine models. *Journal of parasitic Parasitic Diseases: official organ Organ of the Indian Society for Parasitology*, 45(3), 606–619. https://doi.org/10.1007/s12639-020-01337-y
- Mor, S.M., Ascolillo, L.R., Nakato, R., Ndeezi, G., Tumwine, J.K., Okwera, A., & Griffiths, J. K. et al. (2018). Expectoration of *Cryptosporidium* parasites in sputum of human immunodeficiency virus-positive and -negative adults. *American Journal of Tropical Medicine and Hygiene Am J Trop Med Hyg.*, 98(4),1086-90.
- Mosier, D. A., & Oberst, R. D. (2000). Cryptosporidiosis. A global challenge. Annals of the New York Academy of Sciences, 916, 102-111. https://doi.org/10.1111/j.1749-6632.2000.tb05279.x
- Omoruyi, B. E., Nwodo, U. U., Udem, C. S., & Okonkwo, F. O. (2014). Comparative Diagnostic Techniques for Cryptosporidium Infection. *Molecules*, 19(2), 2674-2683. https://doi.org/10.3390/molecules19022674
- Pal M, Tafese W, Tilahun G, Anberber M. (2016). Cryptosporidiosis: an emerging food and waterborne protozoan disease of global significance. *Beverage & Food World*, 2016; 43(1), :43-45.
- Pal, M. (2067). Zoonoses. 2nd Ed. Satyam publishers, Jaipurs, India.Pp.215-216.
- Papini, R.A., & Cardini, G. (2006). Evaluation of a rapid Cryptosporidium/Giardia immunochromatographic test for diagnosis of giardiasis in dogs. *Revue De Medecine Veterinaire*, 157, 490-493.
- Pozio, E., & Morales, M. A. (2005). The impact of HIV-protease inhibitors on opportunistic parasites. *Trends in Parasitology*, 21(2), 58-63. https://doi.org/10.1016/j.pt.2004.11.003
- Ridderstedt, F., Widerström, M., Lindh, J., & Lilja, M. (2018). Sick leave due to diarrhea caused by contamination of drinking water supply with Cryptosporidium hominis in Sweden: a retrospective study. *Journal of Water and Health*, *16*(5), 704–710. https://doi.org/10.2166/wh.2017.311
- Rizvi, S. A. A., & Saleh, A. M. (2018). Applications of nanoparticle systems in drug delivery technology. Saudi pharmaceutical Pharmaceutical journalJournal: SPJ: the official Official publication Publication of the Saudi Saudi Pharmaceutical Society, 26(1), 64–70. https://doi.org/10.1016/j.jsps.2017.10.012
- Rosado-García, F. M., Guerrero-Flórez, M., Karanis, G., Hinojosa, M. D. C., & Karanis, P. (2017). Water-borne protozoa parasites: The Latin American perspective. *International Journal of Hygiene and Environmental Health*, 220(5), 783–798. https://doi.org/10.1016/j.ijheh.2017.03.008
- Rossignol, J. F., Ayoub, A., & Ayers, M. S. (2001). Treatment of diarrhea caused by Cryptosporidium parvum: a prospective randomized, doubleblind, placebo-controlled study of Nitazoxanide. *The Journal of Infectious Diseases*, *184*(1), 103–106. https://doi.org/10.1086/321008
- Rossignol, J. F., Kabil, S. M., el-Gohary, Y., & Younis, A. M. (2006). Effect of nitazoxanide in diarrhea and enteritis caused by Cryptosporidium species. *Clinical gastroenterology Gastroenterology and Hepatology: the Official clinical Clinical Practice Journal of the American Gastroenterological Association*, 4(3), 320–324. https://doi.org/10.1016/j.cgh.2005.12.020
- Rossle, N. F., & Latif, B. (2013). Cryptosporidiosis as threatening health problem: A review. Asian Pacific Journal of Tropical Biomedicine, 3(11), 916–924. https://doi.org/10.1016/S2221-1691(13)60179-3
- Savioli, L., Smith, H., & Thompson, A. (2006). *Giardia* and *Cryptosporidium* join the 'Neglected Diseases Initiative'. *Trends in Parasitology*, 22(5), 203–208. https://doi.org/10.1016/j.pt.2006.02.015
- Shirley, D. A., Moonah, S. N., & Kotloff, K. L. (2012). Burden of disease from cryptosporidiosis. *Current Opinion in Infectious Diseases*, 25(5), 555–563. https://doi.org/10.1097/QCO.ob013e328357e569
- Sorlier, P., Denuzière, A., Viton, C., & Domard, A. (2001). Relation between the degree of acetylation and the electrostatic properties of chitin and chitosan. *Biomacromolecules*, 2(3), 765–772. https://doi.org/10.1021/bm015531+
- Squire, S. A., & Ryan, U. (2017). Cryptosporidium and Giardia in Africa: current and future challenges. Parasites & Vectors, 10(1), 195. https://doi.org/10.1186/s13071-017-2111-y
- Striepen, B. (2013). Parasitic infections: Time to tackle cryptosporidiosis. Nature, 503(7475), 189-191. https://doi.org/10.1038/503189a
- Tomazic M, Garro C, Schnittger L. (2018). Cryptosporidium. In: Florin-Christensen M, Schnittger L, (Eds.), Parasitic Protozoa of Farm Animals and Pets. Cham, Switzerland,: Springer.
- Tzipori, S., & Ward, H. (2002). Cryptosporidiosis: biology, pathogenesis and disease. *Microbes and Infection*, 4(10), 1047–1058. https://doi.org/10.1016/s1286-4579(02)01629-5
- Tzipori, S., & Widmer, G. (2008). A hundred-year retrospective on cryptosporidiosis. *Trends in Parasitology*, 24(4), 184–189. https://doi.org/10.1016/j.pt.2008.01.002
- Ursini, T., Moro, L., Requena-Méndez, A. *et al.* (2020). A review of outbreaks of cryptosporidiosis due to unpasteurized milk. *Infection, 48*, 659–663 (2020). https://doi.org/10.1007/s15010-020-01426-
- Whaeeb, S.T., Alsadoon, Z., Altaee, M.N.K., Alshakir, H.S.S., Salih, H.S., Lattef, F.A., & Kadhim, R.S. (2020). The comparison between male and female of infection *Cryptosporidium* in Baghdad. *International Journal of Pharmaceutical Sciences and Research*, *12*(4), 2530-2532