

CHAPTER 27

MECHANISM RESEARCH AND TREATMENT OF REPEAT BREEDING SYNDROME IN COWS

Pei Nie¹, Muhammd Jamil Ahmad², Chao Chen², Xinxin Zhang², Haimiao Lv², Pei Liu³, Chengxi Jiang³, Vincent Latigo³ and Liguang Yang^{2*}

¹College of Veterinary Medicine, Hunan Agricultural University, Changsha 410128, China

²College of Animal Science and Technology, Huazhong Agricultural University, Wuhan 430070, China

³College of Animal Science and Technology, Jiangxi Agricultural University, Nanchang 330045, China

*Corresponding author: ylg@mail.hzau.edu.cn

INTRODUCTION

Repeat breeding syndrome (RBS) is a critical reproductive disorder in repeated breeder cows, with pregnancy losses after three or more artificial inseminations but no anatomical or infectious abnormalities (Gunther 1981). The incidence of RBS in cow was reported to be 25% in Spain and Australia, 22% in USA and United Kingdom and 10% in Sweden (Bartlett et al. 1986; Eddy 1994; Gustafsson et al. 2002; García-Ispuerto et al. 2007; Canu et al. 2010). RBS is one of the most important factors affecting economic success in the dairy and beef industries, such as increasing intervals to conception, wasteful semen and insemination costs, increased veterinary examination and treatment costs, and additional maintenance charges (Casida 1961; Bartlett et al. 1986; Lafi et al. 1992; Perez-Marin et al. 2012). RBS contributes to declining fertility worldwide, which poses a severe threat to the breeding of cows. Unfortunately, RBS remains unmanageable mainly due to the lack in obvious factors. This chapter is focused on the mechanism research and clinical treatment of repeat breeding syndrome.

Mechanism Research

The physiological cause of repeat breeding syndrome is multifactorial. Nevertheless, many researchers prefer to focus on the following mechanisms: fertilization failure and early embryo death.

Failure of Fertilization

Failure of fertilization may result from inferior oocyte quality of female or unqualified artificial insemination (AI), including inopportune insemination. Poor oocyte quality can be caused by the following factors: chromosomal abnormalities, heat stress, and hormone disorders (Wiltbank et al. 2006; Sartori et al. 2010; Roberto et al. 2013; Sood et al. 2017). Plenty of research shows that repeat breeding cows have poorer oocyte quality and less quantity, which delays cytoplasmic maturation and obstructs fertilization (Kurykin et al. 2011; Kafi et al. 2017). During the research study in heifers directed by two scientists, seventy-four percent of the embryos generated by

superovulation on day 7 after insemination were morphologically normal, and merely 28% were generated by the superovulated repeat breeding group (Gustafsson et al. 1983). A comparison of the amount in gene expression of repeat breeding cow and healthy cumulus-oocyte complex showed 178 genes were distinctly expressed. There were lower expression levels of ANXA1, Lactoferrin, HEM45, LOX-1, and GSTA4 gene in repeat breeding Holstein Friesian heifers, which provides reliable data for seeking the underlying candidate marker genes for RBS (Roberto et al. 2013). Heat stress, one of the major problems for cow fertility, has a harmful impact on the maturation of oocytes and the sustainable development of predatory embryos (Ferreira et al. 2011; Walsh et al. 2011; Silva et al. 2013). For instance, at the embryo development towards the blastocysts period, heat stress can increase the apoptotic index and disturb the expression of some genes in both *in vitro* fertilized and parthenogenetic embryos (Roberta et al. 2016). Hormone disorders is another main factor contributing to the failure of fertilization (Ariane et al. 2011). During the estrous cycle, hormonal asynchrony was found in repeat breeding heifers, with progesterone (P₄) concentration of approximately 0.5-1.0 nmol/L and lower plasma P₄ concentrations after AI (Albihn et al. 1991; Wiltbank et al. 2006). In addition, delayed LH peak, prolonged duration of estrus, and delayed ovulation were considered the evident clinical symptoms in some reports (Bage et al. 2002; Sood et al. 2017). Finally, fertilization failure can also be caused by unsuccessful AI, including in-accurate oestrus detection, incorrect time and unbefitting skill of AI, obstruction of fallopian tubes in females and inferior semen quality in males (Ayalon et al. 1984; Sartori et al. 2010; Yusuf et al. 2010).

Death of Early Embryo

Early embryo death, a major source of repeat breeding syndrome, is mainly generated by inferior oocyte quality and an inadequate uterine environment. The embryonic mortality of ruminants ranges between 20 and 50% (Humblot 2001; Diskin et al. 2008). The highest early embryo death rate is in the first week of a pregnant lactating cow (Walsh et al. 2011). Firstly, nutrition plays a vital role in the cytoplasmic maturation of

How to cite this chapter: Nie P, Ahmad MJ, Chen C, Zhang X, Lv H, Liu P, Jiang C, Latigo V and Yang L, 2022. Mechanism research and treatment of repeat breeding syndrome in cows. In: Abbas RZ, Khan A, Liu P and Saleemi MK (eds), *Animal Health Perspectives*, Unique Scientific Publishers, Faisalabad, Pakistan, Vol. I, pp: 208-213. <https://doi.org/10.47278/book.ahp/2022.27>



oocytes in cows. Accumulation of cytoplasmic lipid deposits and their misdistribution have deleterious effects on oocyte maturation and quality (Adamiak et al. 2006; Leroy et al. 2008). A mass of oocytes with lipid droplets was an inescapable threat to their quality, mature capacities, and cryo-tolerances (Abe et al. 2002). It was reported that the reproductive performance of animals has a connection with their weight. As the body score surpasses 3.5, the cow has delayed initial oestrus, late conception and additional services to be pregnant (Crow et al. 2008). The body condition is inextricably linked to the daily feeding management, and several studies show deleterious influence on oocyte quality and blastocyst rate by immoderate intake (Humblot et al. 1998; Boland et al. 2001; Webb et al. 2004). Secondly, an inadequate uterine environment resulting from endometritis or other diseases can reduce embryo death by growth factors, bacterial toxins, and inflammatory mediators, including endometrial epidermal growth factor, prostaglandins, and reactive oxygen species, and cytokines (Sheldon et al. 2006; Wagener et al. 2017). Cytological endometritis has been reported to have deleterious effects on repeat breeder cows (Salasel et al. 2010; Janowski et al. 2013; Pothmann et al. 2015; Pascottini et al. 2016).

Treatment

Hormonal Treatments

Hormonal treatments, such as progesterone (P_4), gonadotropin-releasing hormone (GnRH), human chorionic gonadotropin (HCG), or insulin, have been widely used to improve the rate of conception for repeat breeding cows (Young et al. 1988; Thuemmel et al. 1992; Selvaraju et al. 2002; Kharche et al. 2007). During early pregnancy in the repeat breeding cows and heifers, the administration of P_4 (75 mg/d for cows, 40 mg/d for heifers, intramuscular injection on daily days 6-10 after AI) may increase plasma P_4 enough to reduce peripheral circulation demand for P_4 from the ovarian circulation, thus decreasing embryonic mortality due to insufficient P_4 . However, it did not affect the conception rate (Thuemmel et al. 1992). Supplementation with PRID (1.55g of P_4) between days 5 and 19 after AI-enhanced the likelihood of pregnancy in 1-2 parity repeat breeding cows. It is worth mentioning that the proportion of abortions tended to be lower in PRID cows. However, there are some contradictory results regarding the advantage of GnRH therapeutic effects (Perry et al. 2009).

On the one hand, some reports indicated that the GnRH or synthetic analogues could improve the pregnancy rate in crossbred cows. There are several preparations of GnRH, such as gonadorelin, buserelin, cystorelin, and fertirelin, and the dose used for them ranges from 10 to 500 μ g in clinical treatments for several decades (Chenault et al. 1990; Stevenson et al. 1990; Ahuja et al. 2005). A study showed that there is a dose-effect relationship in RBS crossbred cow bred, which was previously exceeded 5 times reproductive procedure, particularly in the times of first service and conception rate (Kharche et al. 2007). On the other hand, it was shown that a dose of GnRH at the time of AI did not influence conception rates in beef cattle with obvious behavior of estrus (Heuwieser et al. 2011). Insulin and IGF-I can function as significant accommodators of follicular growth, reproductive hormone fluctuation, oocyte maturation, and embryonic development. Cows in the insulin treatment group were

administrated subcutaneously with a long-acting purified form of bovine insulin (0.2 IU/kg/day on days 8, 9 and 10 of the estrous cycle). The results showed that P_4 concentration increased after insulin injection. However, the pregnancy rate had no first service pregnancy rate, and the overall pregnancy rate did not differ between the insulin injection group and the control group (Selvaraju et al. 2002).

Regulation of Fatty Acids by Diet Therapy

Fatty acids (FA) were not only closely related to the formation and metabolism of many important chemical compounds (hormone, ketosteroid and cholesterol) in reproductive health but also were essential factors for the structure and function of some organelle in germ cells (Abayasekara et al. 1999; Jump et al. 1999; Cheng et al. 2004; Leroy et al. 2005). Dietotherapy of FA can improve milking and reproductive performance, speculated by the possibility of harm reduction in the negative energy balance of high-producing dairy cows. Studies were conducted on diets with some individual fatty acids or FA groups. The results showed that ω -3 PUFAs could positively influence reproductive status, for instance, follicular development, persistent corpus luteum, and high fertility (Staples et al. 1998; Mattos et al. 2004). Results from dietary studies showed that ω -3 added to the ration of ruminants can prolong the presence of corpus luteum by reducing PGF_{2a} and increasing embryonic survival (Diskin et al. 2008; Gulliver et al. 2012). Ambrose and his colleagues verified that dietary supplementation of ALA (alpha-linolenic acid, a type of ω -3) was provided for Holstein cows for twenty-eight successive days before artificial inseminations caused larger preovulatory follicles and fewer embryonic losses (Ambrose et al. 2006). During the non-breeding season, a more significant proportion of saturated FA in cow oocytes could account for a smaller conception rate (Zeron et al. 2001). Feeding fish oil is rich in ω -3 (a blend of eicosapentaenoic acid-EPA and docosahexaenoic acid-DHA, two types of ω -3), from -2 to +2 weeks of breeding, and the experimental results indicated that the group of repeat breeding cows who achieved pregnancies fed by fish oil in the synchronization of estrus and artificial insemination, possessed a greater diameter of the preovulatory follicle and higher expression of mRNA of interferon-stimulated gene transcripts (Aamir et al. 2019).

Acupuncture Therapy

Acupuncture, a simple, safe and effective technique, has been used successfully to treat reproductive disorders in cows, pigs, and women (Fung 1984; Lin et al. 1988; West 2000; Lin et al. 2001). As an alternative treatment, it is recognized gradually by the Oriental. Eighteen repeat breeding cows and heifers were treated by aquapuncture, in which 5 and 10 ml of 50% glucose solution were injected at Shenpeng and Baihui acupoints, respectively. The pregnancy rates after the treatment were 77.7 and 66.6% based on P_4 concentration and rectal palpation, respectively. However, the final rate obtained from the actual delivery of the fetus was 44.4%, which had a similar consequence to GnRH treatments (Lin et al. 2002).

Embryo Transfer Protocol

Embryo Transfer (ET) is diffusely applied to enhance the amount of excellent production performance animals and build

superiority gene population. Many studies have investigated the function of embryo transfer for increasing breeding efficiency in repeat breeding cows (Tanabe et al. 1985; Son et al. 2007; Block et al. 2010; Yaginuma et al. 2019). Rodrigues and his colleagues used the conception rate to compare RBS Holstein cows by AI with those by ET. The conception rate was higher in the group by ET (41.7%) than that by AI (17.9%) (Rodrigues et al. 2007). However, there was no remarkable distinction in the pregnancy rates on two months between health and RBS (82% vs. 70%) (Tanabe et al. 1985). Timed artificial insemination superovulated donors and timed embryo transfer in embryo recipients is a choice to avoid oestrus detection. In a study, conception rates were enhanced subsequent CIDR TAI or TET, compared with the rates subsequent AI after a single injection of PGF_{2α} in the synchronization in RBS cows. There was a higher conception rate in the ET than in the appropriate AI group or the TAI (53.8% vs. 18.5% vs. 7.7%) group (Son et al. 2007). In a word, the embryo transfer protocol provides a capacity for mitigating the threat of inferior quality of oocyte and embryo and remedying the imperfect functionality of the uterine in repeat breeding cows.

Vaginal Treatment

Seminal plasma (SP) and milk osteopontin (mOPN) in the vagina are new approaches in recent years. The normalized endometrial Epidermal growth factor (EGF) profile and preserved fertility in RBS cows were studied (Dagvajiamts et al. 2020; Hay et al. 2022). A series of research have manifested there is a likelihood that SP enhances female reproductive performance by regulating some factors of uterine function and ameliorating prenatary environment in mice, sows, horses, cows, and humans (VWS et al. 1989; Gooneratne et al. 1989; Tremellen et al. 2000; Alghamdi et al. 2004; Robertson 2007; Dagvajiamts et al. 2020). EGF is a critical factor in controlling uterine function and embryo development, which is regulated by estrogen in mice and rats. The healthy cows have two peaks at days 2-4 and 13-14 of the estrus cycle with EGF concentrations of their uterine endometrium, and the RBS cows are deficient in these peaks relatively (Katagiri et al. 2004; Katagiri et al. 2013; Katagiri et al. 2016; Kafi et al. 2017). In a recent study, 5% SP with PBS and only PBS were infused into the vagina of two RBS cows' groups at day 3 of their estrous cycle, respectively (31 vs. 36). The results showed that about 60% of RBS cows had normalized two maximum values of endometrial EGF concentrations again and produced more pregnancies than the controls (44.4 % vs 19.4%) (Dagvajiamts et al. 2020). However, the mechanism of EGF concentrations in the uterus altered by SP infusion into the vagina remains unknown. Nowadays, studies have shown that SP improved females' fertility, which is likely associated with sperm motility and function or the direct mode of action on uteri found in other varieties (Katila et al. 2012; Bromfield 2016). OPN, a glycosylated phosphoprotein, was firstly separated from the bovine diaphyseal bone and widely detected in a variety of epithelial cells and seminal plasma, blood, milk, urine, oviduct, uterus and placenta (Franzen et al. 1985; Cancel et al. 1997; Sodek et al. 2000; Johnson et al. 2003; Tsuji et al. 2007; Goncalves et al. 2008; Schack et al. 2009; Dudemaine et al. 2014). Cow OPN has 262 amino acids and displays a molecular weight of 16- 60 kDa (Kumura et al. 2004; Christensen et al. 2016; Dagvajiamts et al. 2020). mOPN, a bountiful origin of OPN in cows, could promote sperm capacitation, cleavage,

blastulation and embryonic development *in vitro* experiments (Killian et al. 1993; Christensen et al. 2016). Hay and his team separated mOPN from cow milk, identified 3 major protein bands of 31, 37 and 61 kDa and compared three vaginal treatments (mOPN vs. SP vs. PBS only) to normalize EFG profiles and conception rate of RBS cows. The result shows that mOPN regained the normal endometrial EGF profile (56.1% vs 58.1% vs 23.8%) and improved the conception rate (43.5% vs 40% vs 23.8%) in RBS cows. Besides, there are approximate levels between seminal plasma OPN and milk OPN (Hay et al. 2022).

Conclusion

In summary, researchers on mechanism research and treatments of repeat breeding syndrome generally pay more attention to endocrinology and genomics nowadays. We can utilize multi-omics in all kinds of fluids of RBS and healthy cows in the future. In addition, a high-throughput mass spectrometric discovery approach can be used to unravel new information and potential candidate proteins for repeat breeding syndrome. Finally, the complexity of exosome biology of RBS deserves more investigation, which is beneficial for discovering new treatments.

REFERENCES

- Aamir ST et al., 2019. Effect of dietary n-3 polyunsaturated rich fish oil supplementation on ovarian function and interferon stimulated genes in the repeat breeding cow, *Animal Reproduction Science* 211:106230.
- Abayasekara DRE et al.,1999. Effects of altering dietary fatty acid composition on prostaglandin synthesis and fertility. *Prostaglandins Leukotrienes & Essential Fatty Acids* 61:275-287.
- Abe H et al., 2002. Accumulation of cytoplasmic lipid droplets in bovine embryos and cryotolerance of embryos developed in different culture systems using serum-free or serum-containing media. *Molecular Reproduction and Development* 61: 57-66.
- Adamiak S et al., 2006. Body composition, dietary carbohydrates and fatty acids determine post-fertilisation development of bovine oocytes in vitro. *Reproduction* 131:247-258.
- Ahuja C et al., 2005. Pregnancy rate following GnRH + PGF2 treatment of low body condition, anestrus Bos taurus by Bos indicus crossbred cows during the summer months in a tropical environment. *Animal Reproduction Science* 87: 203-213.
- Albiñ H et al., 1991. Embryonic ability to prolong the interestrous interval in virgin and repeat breeding heifers. *Animal Reproduction Science* 26:193-210.
- Alghamdi AS et al., 2004. Equine seminal plasma reduces sperm binding to polymorphonuclear neutrophils (PMNs) and improves the fertility of fresh semen inseminated into inflamed uteri. *Reproduction* 127: 593-600.
- Ambrose DJ et al., 2006. Lower pregnancy losses in lactating dairy cows fed a diet enriched in α-linolenic acid. *Journal of Dairy Science* 89: 3066-3074.
- Ariane BH et al., 2011. Effect of reproductive disorders and parity on repeat breeder status and culling of dairy cows in Quebec. *Canadian Journal of Veterinary Research* 75:147-151.

- Ayalon N et al., 1984. The repeat breeder problem. *Vlaams Diergeneeskundig Tijdschrift* 53: 230-239.
- Bage R et al., 2002. Repeat breeding in dairy heifers: follicular dynamics and estrous cycle characteristics in relation to sexual hormone patterns. *Theriogenology* 9:2257-2269.
- Bartlett PC et al., 1986. Repeated insemination in Michigan Holstein-Friesian cattle: Incidence, descriptive epidemiology and estimated economic impact. *Theriogenology* 26:309-322.
- Block J et al., 2010. Efficacy of in vitro embryo transfer in lactating dairy cows using fresh or vitrified embryos produced in a novel embryo culture medium. *Journal of Dairy Science* 93:5234-5242.
- Boland MP et al., 2001. Effect of nutrition on endocrine parameters, ovarian physiology, and oocyte and embryo development. *Theriogenology* 55: 1323-1340.
- Bromfield JJ, 2016. A role for seminal plasma in modulating pregnancy outcomes in domestic species. *Reproduction* 152: 223-232.
- Cancel AM et al., 1997. Osteopontin is the 55-kilodalton fertility-associated protein in Holstein bull seminal plasma. *Biology of Reproduction* 57: 1293-1301.
- Canu et al., 2010. Predisposition to repeat breeding in UK cattle and success of artificial insemination alone or in combination with embryo transfer. *Veterinary Record* 167: 44-57.
- Perez-Marin CC et al., 2012. Clinical approach to the repeat breeder cow syndrome. In: Perez-Marin CC (ed), *A Bird's-Eye View of Veterinary Medicine*, IntechOpen Limited, London, pp: 337-362.
- Casida LE, 1961. Present status of the repeat-breeder cow problem. *Journal of Dairy Science* 44: 2323-2329.
- Chenault JR et al., 1990. LH and FSH response of Holstein heifers to fertyrelin actate, gonadorelin and buserlin. *Theriogenology* 34: 81-84.
- Cheng Z et al., 2004. Alteration of prostaglandin production and agonist responsiveness by n-6 polyunsaturated fatty acids in endometrial cells from late-gestation ewes. *Journal of Endocrinology* 182: 249-256.
- Crow MA et al., 2008. Resumption of ovarian cyclicity in postpartum beef and dairy cows. *Reproduction in Domestic Animals* 43: 20-28.
- Dagvajiamts B et al., 2020. Effect of seminal plasma infusion into the vagina on the normalization of endometrial epidermal growth factor concentrations and fertility in repeat breeder dairy cows. *Journal of Reproduction and Development* 66: 149-154.
- Diskin MG et al., 2008. Embryonic and early foetal losses in cattle and other ruminants. *Reproduction in Domestic Animals* 43: 260-267.
- Dudemaine PL et al., 2014. Genetic variations in the SPPI promoter affect gene expression and the level of osteopontin secretion into bovine milk. *Animal Genetic* 45: 629-640.
- Eddy RG, 1994. The repeat breeder cow. *Cattle Practice* 2: 39-45.
- Ferreira RM et al., 2011. The low fertility of repeat breeder cows during summer heat stress is related to a low oocyte competence to develop into blastocysts. *Journal of Dairy Science* 94: 2383-2392.
- Franzen A et al., 1985. Isolation and characterization of two sialoproteins present only in bone calcified matrix. *Biochemical Journal* 232: 715-724.
- Fung HP, 1984. The treatment of sterility in cow with electroacupuncture. In: *Proc. 13th World Congress on Disease of Cattle*. Cape Town, South Africa, pp: 844-846.
- García-Ispuerto et al., 2007. Santolaria. Factors affecting the fertility of high producing dairy herds in northeastern Spain. *Theriogenology* 67: 632-638.
- Christensen B et al., 2016. Structure, function and nutritional potential of milk osteopontin. *International Dairy Journal* 57: 1-6.
- Goncalves RF et al., 2008. Pre-treatment of cattle semen or oocytes with purified milk osteopontin affects in vitro fertilization and embryo development. *Animal Reproduction Science* 108: 375-383.
- Gooneratne AD et al., 1989. Effect of intrauterine infusion of sperm antigens on gilt fertility. *Theriogenology* 31:1221-1226.
- Gulliver CE et al., 2012. The role of omega-3 polyunsaturated fatty acids in reproduction of sheep and cattle. *Animal Reproduction Science* 131:9-22.
- Gunther JD, 1981. Classification and clinical management of the repeat breeding cow. *Compendium on Continuing Education for the Practicing Veterinarian* 2:154-158.
- Gustafsson H et al., 1983. Reciprocal embryo transfer between repeat breeder and virgin heifers-an experimental model. *Acta Veterinaria Scandinavica* 24:59-64.
- Gustafsson H et al., 2002. Emanuelson U. Characterisation of the repeat breeding syndrome in Swedish dairy cattle. *Acta Veterinaria Scandinavica* 43:115-125.
- Hay MK et al., 2022. Effects of milk osteopontin on the endometrial epidermal growth factor profile and restoration of fertility in repeat breeder dairy cows. *Theriogenology* 184:26-33.
- Heuwieser W et al., 2011. Efficacy of carprofen on conception rates in lactating dairy cows after subcutaneous or intrauterine administration at the time of breeding. *Journal of Dairy Science* 94:146-151.
- Humblot P et al., 1998. Effects of high energy supply and metabolic status on superovulatory response and embryo production in dairy heifers. *Theriogenology* 49:378.
- Humblot P, 2001. Use of pregnancy specific proteins and progesterone assays to monitor pregnancy and determine the timing, frequencies and sources of embryonic mortality in ruminants. *Theriogenology* 56:1417-1433.
- Janowski T et al., 2013. Prevalence of subclinical endometritis in repeat breeding cows and mRNA expression of tumor necrosis factor and inducible nitric oxide synthase in the endometrium of repeat breeding cows with and without subclinical endometritis. *Polish Journal of Veterinary Science* 16:693-699.
- Johnson GA et al., 2003. Osteopontin:roles in implantation and placentation. *Biology of Reproduction* 69:1458-1471.
- Jump DB et al., 1999. Regulation of gene expression by dietary fat. *Annual. Review of Nutrition* 19:63-90.
- Kafi M et al., 2017. Inherent inferior quality of follicular fluid in repeat breeder heifers as evidenced by low rates of in vitro production of bovine embryos. *Theriogenology* 102:29-34.
- Katagiri S et al., 2004. Changes in EGF concentrations during estrous cycle in bovine endometrium and their alterations in repeat breeder cows. *Theriogenology* 62:103-112.
- Katagiri S et al., 2013. Low incidence of an altered endometrial epidermal growth factor (EGF) profile in repeat breeder Holstein heifers and differential effect of parity on the EGF profile between fertile Holstein (dairy) and Japanese Black

- (beef) cattle. *Journal of Reproduction and Development* 59:575-579.
- Katagiri S et al., 2016. Endometrial epidermal growth factor profile and its abnormalities in dairy cows. *Journal of Reproduction and Development* 62(5): 465-470.
- Katila T et al., 2012. Post-mating inflammatory responses of the uterus. *Reproduction in Domestic Animals* 47:31-41.
- Kharche SD et al., 2007. Dose dependent effect of GnRH analogue on pregnancy rate of repeat breeder crossbred cows. *Animal Reproduction Science* 99:196-201.
- Killian GJ et al., 1993. Fertility-associated proteins in Holstein bull seminal plasma. *Biology of Reproduction* 49:1202-1207.
- Kumura H et al., 2004. Susceptibility of bovine osteopontin to chymosin. *Journal of Dairy Research* 71: 500-504.
- Kurykin J et al., 2011. Morphological quality of oocytes and blood plasma metabolites in repeat breeding and early lactation dairy cows. *Reproduction in Domestic Animal* 46: 253-260.
- Lafi SQ et al., 1992. Epidemiological and economic study of the repeat breeder syndrome in Michigan dairy cattle. I. Epidemiological modeling. *Preventive Veterinary Medicine* 14: 87-98.
- Leroy JL et al., 2008. Reduced fertility in high-yielding dairy cows: are the oocyte and embryo in danger? Part II. Mechanisms linking nutrition and reduced oocyte and embryo quality in high-yielding dairy cows. *Reproduction in Domestic Animals* 43: 623-632.
- Leroy JLMR et al., 2005. Non-esterified fatty acids in follicular fluid of dairy cows and their effect on developmental capacity of bovine oocytes in vitro. *Reproduction* 130: 485-495.
- Lin JH et al., 1988. Effects of electroacupuncture and gonadotropin releasing hormone treatments on hormonal changes in anoestrous sows. *American Journal of Chinese Medicine* 16: 117-126.
- Lin JH et al., 2001. Acupuncture for animal reproductive disorders. In: A.M. Schoen *Veterinary Acupuncture*, 2nd ed. Mosby Inc., USA, pp: 261-267.
- Lin JH et al., 2002. Aquapuncture Therapy of Repeat Breeding in Dairy Cattle. *American Journal of Chinese Medicine* 30: 397-404.
- Mattos R et al., 2004. The effects of feeding fish oil on uterine secretion of PGF $_{2\alpha}$, milk composition, and metabolic status of periparturient Holstein cows. *Journal of Dairy Science* 87: 921-932.
- WS O et al., 1989. Effects of male accessory sex gland secretions on early embryonic development in the golden hamster. *Journal of Reproduction and Fertility* 84: 341-344.
- Pascottini OB et al., 2016. Prevalence of cytological endometritis and effect on pregnancy outcomes at the time of insemination in nulliparous dairy heifers. *Journal of Dairy Science* 99: 9051-9056.
- Perry GA et al., 2009. GnRH treatment at artificial insemination in beef cattle fails to increase plasma progesterone concentrations or pregnancy rates. *Theriogenology* 71: 775-779.
- Pothmann H et al., 2015. The prevalence of subclinical endometritis and intrauterine infections in repeat breeder cows. *Theriogenology* 83: 1249-1253.
- Roberta MF et al., 2016. The Infertility of Repeat-Breeder Cows During Summer Is Associated with Decreased Mitochondrial DNA and Increased Expression of Mitochondrial and Apoptotic Genes in Oocytes. *Biology of Reproduction* 94(3): 1-10.
- Roberto P et al., 2013. Differential gene expression in cumulus oocyte complexes collected by ovum pick up from repeat breeder and normally fertile Holstein Friesian heifers. *Animal Reproduction Science* 14: 26-33.
- Sood P et al., 2017. Preovulatory follicle characteristics and oocyte competence in repeat breeder dairy cows. *Journal of Dairy Science* 100: 9372-9381.
- Robertson SA, 2007. Seminal fluid signaling in the female reproductive tract: lessons from rodents and pigs. *Journal of Animal Science* 85: 36-44.
- Rodrigues CA et al., 2007. Comparison of pregnancy rates after artificial insemination or embryo transfer in high producing repeat breeder Holstein cows. 21st Annual meeting of the Brazilian Embryo Technology Society, Costa do Sauípe. *Acta Scientiae Veterinariae* 35:1255.
- Salasel B et al., 2010. Prevalence, risk factors for and impact of subclinical endometritis in repeat breeder dairy cows. *Theriogenology* 74: 1271-1278.
- Sartori R et al., 2010. Factors affecting fertilisation and early embryo quality in single and superovulated dairy cattle. *Reproduction Fertility and development* 22:151-158.
- Schack L et al. 2009. Considerable variation in the concentration of osteopontin in human milk, bovine milk, and infant formulas. *Journal of Dairy Science* 92:5378-5385.
- Selvaraju S et al., 2002. Fertility responses and hormonal profiles in repeat breeding cows treated with insulin. *Animal Reproduction Science* 73:141-149.
- Sheldon et al., 2006. Defining postpartum uterine disease in cattle. *Theriogenology* 65:1516-1530.
- Silva CF et al., 2013. Effects of heat stress on development, quality and survival of *Bos indicus* and *Bos Taurus* embryos produced in vitro. *Theriogenology* 79:351-357.
- Sodek J et al., 2000. Osteopontin. *Critical Reviews in Oral Biology and Medicine* 11:279-303.
- Son DS et al., 2007. A CIDR-based timed embryo transfer protocol increases the pregnancy rate of lactating repeat breeder dairy cows. *Journal of Reproduction and Development* 53:1313-1318.
- Staples CR et al., 1998. Influence of supplemental fats on reproductive tissues and performance of lactating cows. *Journal of Dairy Science* 81:856-871.
- Stevenson JS et al., 1990. Double insemination and gonadotropin releasing hormone treatment of repeat breeding dairy cattle. *Journal of Dairy Science* 73:1766-1772.
- Tanabe TY et al., 1985. Comparative fertility of normal and repeat-breeding cows as embryo recipients. *Theriogenology* 23:687-696.
- Thuemmel AE et al., 1992. Effect of progesterone supplementation in repeat-breeder cattle on conception and plasma progesterone. *Journal of Endocrinological Investigation* 15:393-396.
- Tremellen KP et al., 2000. The effect of intercourse on pregnancy rates during assisted human reproduction. *Human Reproduction* 15: 2653-2658.
- Tsuji H et al., 2007. Urinary concentration of osteopontin and association with urinary supersaturation and crystal formation. *International Journal of Urology* 14: 630-634.
- Wagener K et al., 2017. A review of the ongoing discussion

- about definition, diagnosis and pathomechanism of subclinical endometritis in dairy cows. *Theriogenology* 94:21-30.
- Walsh SW et al., 2011. A review of the causes of poor fertility in high milk producing dairy cows. *Animal Reproduction Science* 123:127-138.
- Webb R et al., 2004, Control of follicular growth: Local interactions and nutritional influences. *Journal of Animal Science* 82:63-74.
- West Z, 2000. *Acupuncture in Pregnancy and Childbirth*. Churchill Livingstone, New York, USA.
- Wiltbank MC et al., 2006. Changes in reproductive physiology of lactating dairy cows due to elevated steroid metabolism. *Theriogenology* 65: 17-29.
- Yaginuma H et al., 2019. Improvement of fertility in repeat breeder dairy cattle by embryo transfer following artificial insemination: possibility of interferon tau replenishment effect. *Journal of Reproduction and Development* 65: 223-229.
- Young AJ et al., 1988. Effect of GnRH and hCG at time of insemination of repeat breeders cows. *Journal of Dairy Science* 71:137.
- Yusuf M et al., 2010. Reproductive performance of repeat breeders in dairy herds. *Theriogenology* 73:1220-1229.
- Zeron Y et al., 2001. Seasonal changes in bovine fertility: relation to developmental competence of oocytes, membrane properties and fatty acid composition of follicles. *Reproduction* 121:447-454