# **CHAPTER 31**

# **KETOSIS IN DAIRY ANIMALS**

Illahi Bakhsh Marghazani<sup>\*</sup>, Zohaib Ahmed Bhutto, Nasrullah Bangulzai, Muhammad Umar, Abdul Hameed Baloch and Muhammad Jan

Faculty of Veterinary and Animal Sciences, Lasbela University of Agriculture, Water and Marine Sciences (LUAWMS), Uthal, Balochistan-Pakistan

\*Corresponding author: marghazani76@yahoo.com

## INTRODUCTION

Ketosis is also known as acetonemia characterized by the presence of ketone (acetone, acetoacetate and 3hydroxybutyrate) compounds inside the blood and these compounds are eliminated through urine and milk. Generally, ketosis is a metabolic disorder, but in dairy animals it is a lactation syndrome correlated with high milk yielding cows having negative energy balance. Ketosis occurring in the dairy animals is of two types i.e., clinical, and subclinical with signs and without signs respectively.

The transition period (three weeks before and after calving) is important one in high milk producing dairy cattle. During this period (late gestation and early lactation), there is need of more energy. National Research Council also recommended that sufficient nutrients should be included in the diet of animals particularly in the form of bypass nutrients (Marghazani 2012; Das et al. 2014; Gyanendra et al. 2019). To avoid from environment stress, more care should be practiced during last three weeks of pre-partum in dairy animals (Gerloff 2000). It is found that due to intake of energy deficient diet, the huge metabolic burden shifted on the liver due to import of nonesterified fatty acids (NEFA). NEFAs are liberated out from fat stores (triacylglycerol) due to negative energy balance or shortage of glucose. The more concentration of NEFAs into the liver led to increase in oxidation process that finally buoy up liver-connected diseases i.e., ketosis and hepatic lipidosis. Taken together, more acceleration in the gluconeogenesis contributes glucose utilization by the mammary glands (lactose synthesizes) that in turn cause stress in the hepatic tissue of early-lactating animals (Ringseis et al. 2015).

There are two main types of ketosis i.e., clinical with visible signs and subclinical without signs (Youssef et al. 2010). There are many factors involved in the establishment of ketosis during lactating period including body condition score and release of fat during calving (Busato et al. 2002). Ketosis is not considered as life threatening disease (death uncommon) but its negative impact on high yielding dairy cattle have been observed that includes decreased milk production, increased culling rate of early lactating animals, occurrence of displaced abomasum, chances of metritis, less fertility rate, and economical losses (Rajala-Schultz et al. 1999; Oetzel 2013; Steeneveld et al. 2020). More frequency in cases of ketosis, even at normal level imparts overall impact on economic losses to the dairy industry.

## **Classification of Ketosis**

Generally, there are two types of ketosis primary and secondary which differs on the basis of disease source (Herdt 2000). Most of evidence show the occurrence of ketosis in starting days of calving or during early lactating phase (McArt et al. 2012). The terms subclinical and clinical are also commonly used for ketosis classification. Clinical form of ketosis is manifested by increased ketones in biological fluids (blood, urine, milk) along with other symptoms including appetite, decreased weight, and dryness of dung. While the subclinical form of ketosis shows high level of ketones in the blood, milk, and urine, along with absence of major signs. The reason is loosed-type housing system in majority of dairy forms which causes difficulty in observing the signs in specific or individual animal. It was thought that clinicalor subclinical forms of ketosis should be according to the b-hydroxybutyrate (BHB) level inside the blood. Though, it was experienced after careful observation of the animals that increased ketonemia condition did not show clinical signs. Each animal tolerance to clinical ketosis were different and accordingly some animals show clinical signs whilst others may not show (Herdt 2000). It is long-established that term hyperketonemia is best for description of ketosis rather than its clinical or subclinical forms.

### **Biochemistry and Cycle of Ketones Production**

The production source of ketones is associated with fatty acids and amino acids from the fat reservoir tissues in the body and feed (Coelho et al. 2013; Kohlmeier 2015). The process of ketogenesis happens in mitochondria of hepatocytes. Adipokine signal triggering the release of fatty acid from the fat reservoirs results in increasing level of glucagon and epinephrine and decrease in the concentration of insulin. Such happening led to development of hypoglycemic and fasting states (Owen 2005). Coenzyme A is attached with fatty acids and transported to mitochondria. Fatty acids through betaoxidation splits 2 carbons from the acyl-CoA compound in each cycle for the formation of acetyl-CoA. This acetyl-CoA transported into citric acid cycle for the formation of citric acid, and then citric acid incorporated into the tricarboxylic acid cycle (TCA), finally high energy produced (Stryer 1995). These mechanisms are shown in flow diagram (Figure-2). Every cell of the body can metabolize the acetyl-CoA by TCA cycle,

How to cite this chapter: Marghazani IB, Bhutto ZA, Bangulzai N, Umar M, Baloch AH and Jan M, 2022. Ketosis in dairy animals. In: Abbas RZ, Khan A, Liu P and Saleemi MK (eds), Animal Health Perspectives, Unique Scientific Publishers, Faisalabad, Pakistan, Vol. I, pp: 232-237. <u>https://doi.org/10.47278/book.ahp/2022.31</u>

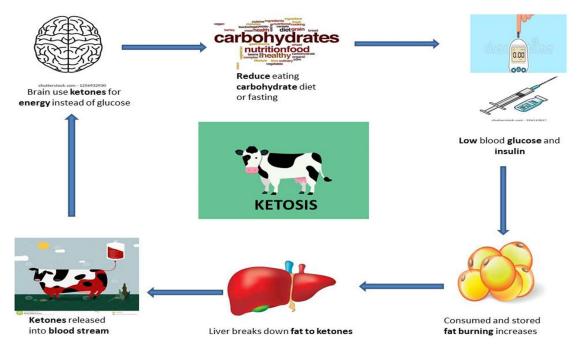


Fig. I: Factors leading to Ketosis in dairy animals (Zhang and Ametaj, 2020).

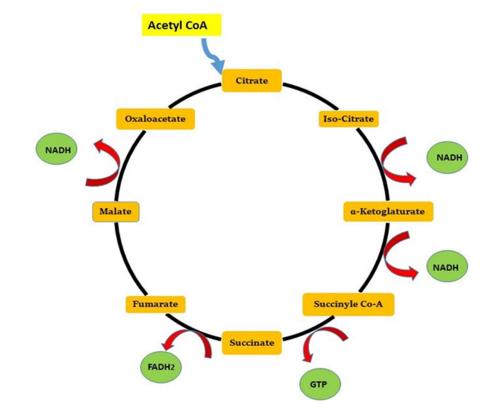


Fig. 2: Kreb's Citric Acid Cycle (Tri-Carboxlic Acid Cycle): (Martínez and Chandel 2020; Protasoni and Zeviani 2021)

but it is also involved in ketogenic process in the hepatocytes (Laffel 1999). During decreased level of glucose inside the blood, oxaloacetate intermediate compound of TCA cycle transported and is utilized in the production of glucose (gluconeogenesis). Oxaloacetate is used in gluconeogenesis prevent its condensation with acetyl-CoA, hence no entrance of acetyl-CoA into the TCA cycle. In nutshell, the energy was generated in the form of ketones from acetyl-CoA. In ketone

occurring process, acetyl-CoA 2 molecules converted into acetoacetyl-CoA by condensation with the help of thiolase. In short, acetoacetyl-CoA plus a new molecule of acetyl-CoA produced the hydroxy- $\beta$ -methylglutaryl (HMG)-CoA with the help of HMG-CoA synthase. HMG-CoA converted into acetoacetate (ketone body) with HMG-CoA lyase enzyme. D- $\beta$ -hydroxybutyrate type of ketone formed from acetoacetate through D- $\beta$ -hydroxybutyrate dehydrogenase action. Due to result of continuous degradation of acetoacetate, acetone (ketone) and  $CO_2$  produced. In the end, ketone compounds not used as fuel of energy by the liver but transported as energy source towards brain and other tissues of body. Primarily ketones not only produced from fatty acids, but also produced from ketogenic amino acids. These amino acids are converted into the intermediate compounds of citric acid cycle through deamination (Kohlmeier 2015).

### **Causes and Factors Involved in Ketosis**

In context of particular biochemical and physiological etiology of ketosis, no proven data is available till date. There is no single cause, while it is well established that misbalancing in nutrient availability, particularly low energy, is a crucial factor. It is proved that glucose deficiency is associated with the occurrence of ketosis during early lactation phase of dairy cattle. Because 60-85% glucose is utilized by the udder for the collecting lactose contents in milk (Reynolds 2005). Deficiency in ACTH or cortisone hormones is also one of the causes of ketosis that cause pathological changes in adrenal gland (Dressler et al. 2019). Another possible cause of ketosis is the deficiency of oxaloacetic acid (OAA) in hepatic tissues (Bach 1978).

Some risk factors are associated directly or indirectly with occurrence of ketosis. Decreased milk yield and production is affected by the presence of ketones inside the body fluids (Dohoo and Martin 1984). Reported reproductive pathological conditions also associated with ketosis (Dohoo and Martin 1984). Ketotic dairy animal were seen with reduction in conception rate during first artificial insemination shot. This may help in culling of dairy animal at early phase of milking (Oetzel 2013). High milk producing ketotic animals had chance of displaced abomasal disease (Koeck et al. 2013). Among feeding management factors, the dairy cows at calving should not be fatty enough, otherwise it may cause reduced feed intake during early lactation. It is found that body condition score of 2.5-3.0 on a 1-5 scale is optimal and any increases in it enhance risk of ketosis (Melendez et al. 2004). Due to occurrence of associated other diseases, ketosis thought to be gateway disease during starting milking phase. Prevention and treatment of ketosis could decrease the possible chance of associated diseases.

### **S**igns of Ketosis

Ketosis is related with the presence of ketones in fluids and characterized by hypoxia along with drop in milk yield. Decreased intake of diet by lactating animal is indication of first clinical sign during ketosis. In some cases, change in rumen motility (active to inactive) was also observed.

Other important clinical signs observed during ketosis were anorexia, decline in milk yield, body weight loss, poor body appearance, dry dung, and sometimes nervous signs. Nervous dysfunction signs are observed in nervous ketosis type which includes not normal licking, pica, absence of normal gait, incoordination, bellowing and aggression. While in cases of subclinical form of ketosis, less signs or no signs are found (Herdt 2019).

### Diagnosis

In ketosis, hypoglycemic condition appeared due to less carbohydrate diet intake by the dairy animals. As a result, body

produce one chemical named as Beta-hydroxybutyrate (BHB), which facilitate provision of energy when body is in need of sugar. The common approach related with diagnosis of ketosis in dairy animal is blood, milk, and urine testing. In normal circumstances, quantification of BHB level in biological fluid with the help of specialized meter indicates the occurrence of ketosis. The cut off value of BHB considered for non-ketotic cows is <1.2 mmol/L whilst level of BHB >1.2 mmol/L in cows are considered as ketogenic (Itle et al. 2015).

Numerous diagnostic tests (strips, kits, powders, and tablets) are used for the detection of ketosis which give results within seconds and minutes. Acetoacetate from urine (Ketostix strip, Bayer, Leverkusen, Germany) and BHB from milk or blood (Ketolac, Biolab, München, Germany) could be detected by using strips. These tests qualitatively confirm the presence of ketone in body fluids (Geishauser et al. 1998, Geishauser et al. 2000; Carrier et al. 2004; Oetzel 2007). The quantitative confirmatory methods of ketosis detection are using electronic BHB meter, digital devices, and high technological equipment (thin layer chromatography, high resolution gas chromatography and flame ionization detector/mass spectrometer (HRGC-FID/MS). With these techniques, fatty acids profile can be quantified inside the blood and milk and their connection with BHB level (BHB < 1.2 mmol/L). The fatty acids could be valued biomarker that guides in predicting the future occurrence of hyperketonemia (Zhang et al. 2012; Zhang et al. 2013; Li et al. 2014; Zeng and Cao 2018). Diagnosis before occurrence of ketosis may be helpful in the productive and profitable management of lactating animals. In advance, inclusion of various feed additivesor supplements and pharmaceutics could prevent the ketosis.

### **Prevention and Treatments**

Various factors are associated with ketosis including glucose utilization in the formation of lactose (Kronfeld 1972), excess fat on animal body during calving (Smith et al. 1997), less energy diet in post calving period (Dann et al. 2005), not normal hepatic function (Tendler et al. 2007), endocrine glands illness such as ACTH or glucocorticoids, more or less inclusion of proteins diet, mineral and vitamins deficits, and more inclusion of ketogenic diet. Ketosis can be prevented effectively by avoiding these along with few other factors.

One of the approaches for the prevention of ketosis is inclusion of forages (rich in carbohydrates) in the diet of high milk yielding cows during transition period. It will also prevent the occurrence of hypoglycemic cows (Vickers et al. 2013). Second approach, addition of feed additives (supplements, pharmaceutics, herbal) in the diet can also avoid the ketosis in dairy cows (Mammi et al. 2021).

There are shortcomings in well-planned treatment regimens of ketosis and in efficacious ketosis treatment. The most of focused studies were done on treatment of ketosis rather than improvement of production or yield of dairy animals. Variety of treatments had been practiced in dairy cattle industry, with variable outcomes. More treatments had been reported along with other existing diseases but specifically and common one treatment only for ketosis practiced in dairy animals is described hereinafter.

It was proved in 1930s that hypo-glycemic condition occurred in ketosis (McSherry et al. 1960). From that period, glucose/dextrose is used primarily for treating the ketosis. Dextrose treatment play role in correcting the physiology of the animal as glucose connected with the milk production further corrected the hypoglycemia and fat metabolism (Herdt 2000). It is believed that concentration of glucose is more in 500 ml bottle (50% dextrose). 50% dextrose will boost the glucose level inside the blood eight times more than normal level immediately after injection and normal after 2 hours (Sakai et al. 1996). Due to dextrose administration, the impact on BHB levels is short lived (<24 hours) and dextrose should be repeated for additional impacts (Wagner and Schimek 2010). It had been reported that continuous dextrose administration resulted in hyperglycemia and abomasum dysfunction by decreasing its motility (Holtenius et al. 2000; Zadnik 2003; Sahinduran and Albay 2006; Šamanc et al. 2009). Though, dextrose administration single treatment in connection with onward effects yet not well-known. Dextrose should be selected by the veterinary clinicians as secondary treatment ketonemia. Dextrose could provide reverse effect in animals which were hypoglycemic along with associated signs in nervous system. Later on, animals were administered with specific treatment regime for long period of time (Herdt and Emery 1992; Wagner and Schimek 2010).

Glucocorticoids is one of the corticosteroids that practiced in managing ketosis due to its impact on increasing level of glucose (Herdt and Emery 1992). It is proven that steroids having lessening effect on insulin whist increasing influence on breakdown of fat and protein reservoirs. Dexamethasone injection resulted in increased level of glucose and insulin hormone after 48 hours (Jorritsma et al. 2004). Corticosteroids were not recommended for long term usage in ketosis, due to less efficacious effects along with chances of side effects.

At early stage of lactation, dairy animals are inherently considered as insulin resistant (Bauman 2000). In characteristics, it is in-fact part of the complex mechanism of homeorhesis. During negative energy balance, it allows early lactating animals to produce increased amount of milk. Increased insulin resistance is manifested in animals affected by ketosis than other healthy animals (Sakai et al. 1996). In ketosis affected animals, insulin is used for treatment of ketosis because of its anabolic effect. This hormone lessens breakdown of fat, increases synthesis of fat and use of ketone bodies as energy sources. This interconnected metabolic activity decreases the level and significance of ketonemia. Literature shows limited evidence in favor of insulin as therapy.

It is found that vitamin  $B_{12}$  (cyanocobalamin) is involved in the process of gluconeogenesis, that's why it is administered in ketonemia. It is speculated that vitamin  $B_{12}$  have ability in stimulation of methylmalonyl-coenzyme (CoA) A mutase, which is main component of krebs cycle and gluconeogenesis (Kennedy et al. 1990, Gordon et al. 2017). The increasing level of this enzyme will produce more efficient energy for the cells. Butaphosphan is one of the compound and precursor of phosphorus production administered in ketosis for its role in gluconeogenesis (Rollin et al. 2010). Phosphorus involved in the phosphorylation of variety of compounds which occurs during gluconeogenesis. Butaphophan-cyanocobalamin pharmaceutical product could be administered in management of ketosis if its impact on milk yield and risk on disease should be validated.

During 1954, it was discovered that ketosis can be treated with propylene glycol (Johnson 1954; Maplesden 1954). Propylene glycol 100% (300 ml) administered orally once a day upto the period of one week (Gordon et al. 2013). In rumen it is

transported into the blood, or before converted into the propionate (Nielsen and Ingvartsen 2004). It is involved in the TCA cycle by boosting the oxidation of acetyle CoA, increasing the glucose formation and triggers the insulin (Studer 1993). Insulin level is increased after 15 minutes administration of propylene glycol, and hormone surge maintained into the blood for 2 hours (Studer 1993). Many veterinarians use the propylene glycol by incorporating it into the feed as feed additive (Nielsen and Ingvartsen 2004), so, ruminal environment slowly adjusted for long term till formation of propionate (Nielsen and Ingvartsen 2004). In relation with hepatic oxidation theory, this will result in reduced feed consumption, more fat utilization and prolonging the problem of ketosis, while the clinical significance of this has not been resolved (Allen et al. 2004).

Recently documented review article suggested that monensin is one of the ionotropic antibiotics popularized in developed countries (Europe, USA, and Canada) for the prevention of the ketosis in dairy cows. It produces impact by raising the level of propionate in rumen, by reducing the BHB level inside the blood and by improving the indicators of functional liver (Mammi et al. 2021).

The combined treatment regimens were practiced in resolving the ketosis. Implementation of multiple pharmaceutical products had showed good outcomes in comparison with usage of single product. One of the problems with usage of combined therapy is economically not sound. Taken together, in future research trials should be validated regarding application of each product individually. Additional research is needed for efficacious treatment plan of ketosis which should be economically sound and can be implemented in livestock sector.

#### **Economic Losses Due to Ketosis**

The economic losses associated with ketosis depend on individual and herd-level and also associated with other factors. According to one report, the cost for one case of clinical ketosis and subclinical ketosis in average way estimated as  $\notin$ 709 and  $\notin$ 150. Normal herd level ketosis rates (clinical plus subclinical) were  $\notin$ 3,613 for a default farm and  $\notin$ 7,371 yearly for a high-risked farm (Steeneveld et al. 2020). In another report, it is recorded that cost of ketosis is  $\notin$ 21/cow/year (Van Soest et al. 2019). In Canada, one case of subclinical ketosis treatment cost is \$203 (Gohary et al. 2016).

In earlier literature, it is also reported that the losses in dairy animals due to ketosis is about 2.2 to 3.1 lbs of daily milk (4.4 to 6.6%) (Dohoo and Martin 1984; Chapinal et al. 2012), 4.1 lbs daily decline (5.5%) in yield (Duffield et al. 2009), and 865 lbs during 305 days milking period (7%). At herd level, frequency of ketosis cases is about 25-60%. More frequency in cases of ketosis, even at normal level influences overall impact on economic losses to the dairy industry.

#### Conclusion

Ketosis is an important metabolic disease of early lactating dairy animals affecting production performance and consequent economic losses. This disease can be avoided by proper nutrition and good management of dairy cattle during transition period and at early stage of lactation. The application of quick detection tests can support in early management of problem and preventing more income losses.

- Allen MS et al., 2009. Board-invited review: The hepatic oxidation theory of the control of feed intake and its application to ruminants. Journal of Animal Science 87(10): 3317-3334.
- Bach A, 1978. Oxaloacetate deficiency in MCT-induced ketogenesis. Archives International es de Physiologie et de biochimie 86(5): 1133-1142.
- Bauman DE, 2000. Regulation of nutrient partitioning during lactation: homeostasis and homeorhesis revisited. Ruminant Physiology: Digestion, Metabolism, Growth and Reproduction 311-328.
- Busato A et al., 2002. Body condition scores in dairy cows: associations with metabolic and endocrine changes in healthy dairy cows. Journal of Veterinary Medicine Series A 49(9): 455-460.
- Carrier J et al., 2004. Evaluation and use of three cow side tests for detection of subclinical ketosis in early postpartum cows. Journal of Dairy Science 87(11): 3725-3735.
- Chapinal NME et al., 2012. The association of serum metabolites in the transition period with milk production and early-lactation reproductive performance. Journal of Dairy Science 95: 1301-1309.
- Coelho DAL et al., 2013. FGF21 mediates the lipid metabolism response to amino acid starvation. Journal of Lipid Research. 54:1786–1797.
- Dann HM et al., 2005. Prepartum intake, postpartum induction of ketosis, and periparturient disorders affect the metabolic status of dairy cows. Journal of Dairy Science 88(9): 3249-3264.
- Das LK et al., 2014. Metabolizable protein systems in ruminant nutrition: A review. Veterinary World 7(8): 622-629.
- Dohoo IR and Martin SW, 1984. Subclinical ketosis: prevalence and associations with production and disease. Canadian Journal of Comparative Medicine 48(1): 1.
- Dressler A et al., 2019. Efficacy and tolerability of the ketogenic diet versus high-dose adrenocorticotropic hormone for infantile spasms: A single-center parallel-cohort randomized controlled trial. Epilepsia 60 (3): 441-451.
- Duffield TF et al., 2009. Impact of hyperketonemia in early lactation dairy cows on health and production. Journal of Dairy Science 92: 571-580.
- Gyanendra SK et al., 2019. Effect of rumen-protected nutrients on feed intake, body weights, milk yield, and composition in Murrah buffaloes during early lactation. Tropical Animal Health and Production 51: 2297–2304.
- Geishauser T et al., 1998. Evaluation of five cow side tests for use with milk to detect subclinical ketosis in dairy cows. Journal of Dairy Science 81(2): 438-443.
- Gerloff BJ, 2000. Dry cow management for the prevention of ketosis and fatty liver in dairy cows. Veterinary Clinics of North America: Food Animal Practice 16(2): 283-292.
- Geishauser T et al., 2000. Evaluation of eight cow-side ketone tests in milk for detection of subclinical ketosis in dairy cows. Journal of Dairy Science 83(2): 296-299.
- Gohary K et al., 2016. The cost of a case of subclinical ketosis in Canadian dairy herds. The Canadian Veterinary Journal 57(7): 728.
- Gordon JL et al., 2017. Effects of a combination butaphosphan and cyanocobalamin product and insulin on ketosis resolution and milk production. Journal of Dairy Science 100(4): 2954-2966.

- Gordon JL et al., 2013. Ketosis treatment in lactating dairy cattle. Veterinary Clinics: Food Animal Practice 29(2): 433-445.
- Herdt TH, 2000. Ruminant adaptation to negative energy balance: Influences on the etiology of ketosis and fatty liver. Veterinary Clinics of North America: Food Animal Practice 16(2): 215-230.
- Herdt TH, 2019. Overview of ketosis in cattle. MERCK Veterinary Manual 12.
- Herdt TH and Emery RS, 1992. Therapy of diseases of ruminant intermediary metabolism. Veterinary Clinics of North America: Food Animal Practice, 8(1): 91-106.
- Holtenius K et al., 2000. The effect of the plasma glucose level on the abomasal function in dairy cows. Journal of Animal Science 78(7): 1930-1935.
- Itle AJ et al., 2015. Clinical ketosis and standing behavior in transition cows. Journal of Dairy Science 98(1): 128-134.
- Johnson RB, 1954. The treatment of ketosis with glycerol and propylene glycol. Cornell Veterinarian 44: 6-21.
- Jorritsma R et al. 2004. Effects of a single dose of dexamethasone-21-isonicotinate on the metabolism of heifers in early lactation. Veterinary Record 155 (17): 521-523.
- Kennedy DG et al., 1990. Methylmalonyl-CoA mutase (EC 5.4. 99.2) and methionine synthetase (EC 2.1. 1.13) in the tissues of cobalt-vitamin B12 deficient sheep. British Journal of Nutrition 64 (3): 721-732.
- Kohlmeier M, 2015. Nutrient metabolism: structures, functions, and genes. Academic Press.
- Koeck A et al., 2013. Genetic associations of ketosis and displaced abomasum with milk production traits in early first lactation of Canadian Holsteins. Journal of Dairy Science 96 (7): 4688-4696.
- Kronfeld DS, 1972. Glucose, milk production, and ketosis (No. COO-3400-16). Pennsylvania Univ., Kennett Square (USA). School of Veterinary Medicine.
- Laffel L, 1999. Ketone bodies: a review of physiology, pathophysiology and application of monitoring to diabetes. Diabetes/Metabolism Research and Reviews 15(6): 412-426.
- Li Y et al., 2014. Plasma metabolic profiling of dairy cows affected with clinical ketosis using LC/MS technology. Veterinary Quarterly 34 (3): 152-158.
- Maplesden DC, 1954. Propylene glycol in the treatment of ketosis. Canadian Journal of Comparative Medicine and Veterinary Science 18(8): 287
- Marghazani IB, 2012. Effect of protein supplements varying in ruminal degradability on milk production, composition and nutrient utilization in early lactating Sahiwal cows and Nili-Rari buffaloes. PhD thesis. Department of Animal Nutrition. University of Veterinary and Animal Sciences, Lahore, Pakistan.
- McArt JAA et al., 2012. Epidemiology of subclinical ketosis in early lactation dairy cattle. Journal of Dairy Science 95(9): 5056-5066.
- McSherry BJ, 1960. Ketosis in cattle-a review. Can Vet Journal 1(5): 208–13.
- Mammi LM et al., 2021. The Use of Monensin for Ketosis Prevention in Dairy Cows during the Transition Period: A Systematic Review. Animals 11(7): 1988.
- Martínez RI and Chandel NS, 2020. Mitochondrial TCA cycle metabolites control physiology and disease. Nature Communications 11(1): 1-11.

- Melendez P et al., 2004. Plasma mineral and energy metabolite concentrations in dairy cows fed an anionic prepartum diet that did or did not have retained fetal membranes after parturition. American Journal of Veterinary Research 65(8): 1071-1076.
- Nielsen NI and Ingvartsen KL, 2004. Propylene glycol for dairy cows: A review of the metabolism of propylene glycol and its effects on physiological parameters, feed intake, milk production and risk of ketosis. Animal Feed Science and Technology 115(3-4): 191-213.
- Oetzel GR, 2013. Understanding the impact of subclinical ketosis. University of Wisconsin: Madison 15-26.
- Oetzel GR, 2007. Herd level ketosis diagnosis and risk factors. In Preconference seminar C (Vol. 7, pp. 67-91).
- Owen OE, 2005. Ketone bodies as a fuel for the brain during starvation. Biochemistry and Molecular Biology Education 33(4): 246-251.
- Stryer L, 1995. Biochemistry (Fourth ed.). New York: W.H. Freeman and Company. pp. 510–778.
- Protasoni M and Zeviani M, 2021. Mitochondrial structure and bioenergetics in normal and disease conditions. International Journal of Molecular Sciences 22(2): 586.
- Rajala-Schultz PJ et al., 1999. Effects of milk fever, ketosis, and lameness on milk yield in dairy cows. Journal of Dairy Science 82 (2): 288-294.
- Reynolds C, 2005. Glucose balance in cattle. Florida Ruminant Nutrition Symposium pp. 143-154.
- Rollin E et al., 2010. The effect of injectable butaphosphan and cyanocobalamin on postpartum serum  $\beta$ -hydroxybutyrate, calcium, and phosphorus concentrations in dairy cattle. Journal of Dairy Science 93(3): 978-987.
- Ringseis R et al., 2015. Molecular insights into the mechanisms of liver-associated diseases in early-lactating dairy cows: hypothetical role of endoplasmic reticulum stress. Journal of Animal Physiology and Animal Nutrition 99(4): 626-645.
- Sakai T et al., 1996. Glucose and xylitol tolerance tests for ketotic and healthy dairy cows. Journal of Dairy Science 79(3): 372-377.
- Šamanc H et al., 2009. Glucose tolerance test in the assessment of endocrine pancreatic function in cows before and after surgical correction of left diplaced abomasum. Acta Veterinaria-Beograd 59 (5-6): 513-523.
- Sahinduran S and Albay MK, 2006. Haematological and biochemical profiles in right displacement of abomasum in cattle. Revue de Médecine Vétérinaire 157(7): 352-356.

- Smith TR et al., 1997. Metabolic characteristics of induced ketosis in normal and obese dairy cows. Journal of Dairy Science 80(8): 1569-1581.
- Steeneveld W et al., 2020. Estimating the combined costs of clinical and subclinical ketosis in dairy cows. PloS One 15 (4): e0230448.
- Studer VA et al., 1993. Effect of prepartum propylene glycol administration on peri-parturient fatty liver in dairy cows. Journal of Dairy Science 76(10): 2931-2939.
- Tendler D et al., 2007. The effect of a low-carbohydrate, ketogenic diet on nonalcoholic fatty liver disease: a pilot study. Digestive Diseases and Sciences 52(2): 589-593.
- Van Soest et al., 2019. Farm-specific failure costs of production disorders in European organic dairy herds. Preventive Veterinary Medicine 168: 19-29.
- Vickers LA et al., 2013. Feeding a higher forage diet prepartum decreases incidences of subclinical ketosis in transition dairy cows. Journal of Animal Science 91(2): 886-894.
- Wagner SA and Schimek DE, 2010. Evaluation of the effect of bolus administration of 50% dextrose solution on measures of electrolyte and energy balance in postpartum dairy cows. American Journal of Veterinary Research 71(9): 1074-1080.
- Youssef MA et al., 2010. Ketosis in buffalo (Bubalus bubalis): clinical findings and the associated oxidative stress level. Tropical Animal Health and Production 42(8): 1771-1777.
- Zadnik T, 2003. A comparative study of the hematobiochemical parameters between clinically healthy cows and cows with displacement of the abomasum. Acta Veterinaria 53 (5-6): 297-310.
- Zeng M and Cao H, 2018. Fast quantification of short chain fatty acids and ketone bodies by liquid chromatographytandem mass spectrometry after facile derivatization coupled with liquid-liquid extraction. Journal of Chromatography B 1083: 137-145.
- Zhang G and Ametaj BN, 2020. Ketosis an old story under a new approach. Dairy 1(1): 42-60.
- Zhang H et al., 2013. Plasma metabolomic profiling of dairy cows affected with ketosis using gas chromatography/mass spectrometry. BMC Veterinary Research 9(1): 1-13.
- Zhang Z et al., 2012. Detection of Subclinical Ketosis in Dairy Cows. Pakistan Veterinary Journal 32(2):156-160.