

Babesiosis in Cattle

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INTRODUCTION

Cattle is an important dairy and meat producing animal playing an important role in the economy (Saunsoy 1995; Suarez and Noh 2011; Suarez et al. 2018). *Babesia* is a protozoan parasite belonging to the genus piroplasmida, causes a deadly disease in livestock and farm animals and is transmitted by the ticks. Because the illness has direct economic effects like decreased milk output, loss of body weight, and animal death, it poses major issues for both animal life and farm economies (Menshaw 2020). It also exerts secondary costs associated with treatment and prevention (Guswanto et al. 2017). The several regions of Africa, Australia, America, and Asia, particularly India, have a great impact on the cattle industry (Bock et al. 2004; Bal et al. 2016; Hashem et al. 2018). It affects and spreads in tropical as well as subtropical countries (Beugnet and Moreau 2015; Rozej-Bielicka et al. 2015). It causes lack of appetite, fever, anemia, ceasing rumination, and increases in heart and respiratory rates. In later stages, it may lead to hemoglobinuria, a yellowish mucous membrane, and the death of animal (Wagner et al. 2002; Zintl et al. 2003; Demeke et al. 2018; Mezouaghi et al. 2019). According to (Silva et al. 2010), the Ixodidae tick can transmit the babesiosis infection to several animal species. *Babesia* (*B.*) *bovis* and *B. bigemina* are the two most important babesia species in cattle (Zintl et al. 2013). *B. divergens*, is one of the main babesia species that causes bovine babesiosis, and raised concerns among international health authorities (OIE). *Rhipicephalus* and *Ixodes* tick species can transmit babesiosis to cattle depending on the disease's type (Jabbar et al. 2015). *B. bovis* and *B. bigemina* can be transmitted by number of vectors including *Rhipicephalus* (*R.*) *microplus*, *R. annulatus*, and *R. geigy*, whereas *R. decoloratus* and *R. evertsi* can only be transmitted by *B. bigemina*. *Ixodes* (*I.*) *ricinus* typically transmits *B. divergens* (Bock et al. 2004; Gohil et al. 2013).

Etiology and Morphology

Babesiosis is also known by the various other names i.e., Piroplasmosis, Texas fever, and Red water fever (Sahinduran, 2012). The genus *Babesia* includes the two main species which are *B. bovis* and *B. bigemina* Belonging to the phylum Apicomplexa and class Sporozoa (Allsopp et al. 1994; Radostits et al. 2006). Furthermore, the taxonomical classification of *Babesia* species was based on the phylogenetic analysis of 18s rRNA (Criado-Fornelio et al. 2003). Babesiosis in bovine is caused by several species of babesia i.e., *B. bovis*, *B. bigemina*, and *B. divergens* are the three most prevalent pathogenic species (Kaandorp 2004; Radostits et al. 2007; Fakhar et al. 2012). *B. bovis* infection can result in more serious illness than *B. bigemina* (Gubbels et al. 1999). The parasite *B. bovis* is located in the core of the RBCs. Its dimensions are 1.1-1.5 x 0.5-1.0 μ m. While *B. bigemina* is longer than other species and can be seen in pairs. It has a pear-like form. It is 1-1.5 μ m wide and 3-3.5 μ m long (Soulsby 1986; El Sawalhy 1999). According to (Jerram and Willshire 2019) and (Alvarez et al. 2019), *B. divergens* has a small, thin, and obtuse angle (Fig. 1). Moreover, *B. major*, *B. ovata*, *B. occultans*, and *B. jakimovi* can also infect the cattle (Menshaw 2020).

Life Cycle of Bovine Babesiosis

All species belonging to the genus *Babesia* have shown same life cycle stages with minor differences. Some species showed transovarial transmission (*Babesia* spp. sensu stricto) while other may be transmitted through transstadial route (*B. microti*) (Saad et al. 2015). Their life cycle can be completed in three main stages:

- Gametogony: fusion and formation of gametes occur in the gut of the ticks.
- Sporogony: It is asexual reproduction taking place in the salivary gland of tick
- Merogony: It take place in the vertebrates (Fig. 2) (Otify 2011; Abdela and Jilo 2016). Binary fission is the way of multiplication inside the red blood cells, and causing considerable pleomorphism followed by the gametocyte formation. The conjugation of gametocyte take place in the tick gut followed by the multiplication and migration to the different tissues such as salivary glands. Furthermore, the continuous development occurs in the salivary glands. The transovarial transmission may happen at this stage (Gray et al. 2010). The host will be infected when the larvae sucks the blood. The larvae transform in to the nymph after molting which is then converted in to adult. Host may get the



Fig. 1: Babesia parasites inside red blood cells

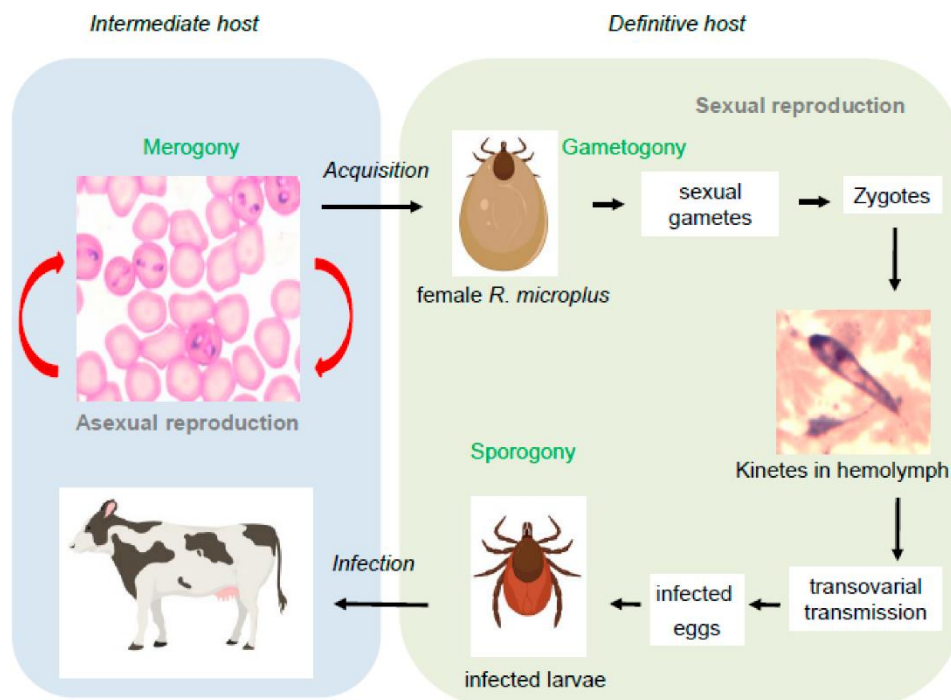


Fig. 2: Babesia Species life cycle (Gallego-Lopez et al. 2019)

infection, when vector takes a blood meal (Uilenberg 2006; Simuunza 2009; Lefevre et al. 2010; Mandal 2012; Schnittger et al. 2012; Ozubek et al. 2020).

Host Range

Out of hundred types of *Babesia* spp., only eighteen species can cause infection in domestic animals (Suarez and Noh 2011). Babesiosis mainly affects cattle, goats, sheep, horses, dogs, cats and human (Hamsho et al. 2015; Gray et al. 2019).

B. bovis and *B. bigemina* have recently been detected in deer. The primary host for *Babesia* spp. is cattle while all other animals are considered of little epidemiological distribution (CFSPH, 2008).

Geographic Distribution

Babesiosis in cattle is present across the world due to presence of vector. However, tropical and subtropical locations frequently experience it (CFSPH, 2008). The

highest prevalence of babesiosis is found in areas where ticks vector is present excessively. They are especially important in Australia, Africa, Asia, and the United States. Even while *B. bovis* typically inhabits the same habitats as *B. bigemina*, only a small number of other tick species have the ability to transmit both species. Additionally, the regional distribution of these ticks varies with the area. For instance, the two tick species can serve as a biological vector, *B. bigemina* is widely distributed in Africa (Spickler et al. 2010; Pohl 2013).

Risk Factors

Host Factors

Host factors which mainly affect the presence of disease include breed, age and immune status of the animals (Jabbar et al. 2015).

- Regarding the age of the host, the infection rate among young animals is low due to innate resistance, which is boosted by maternal antibodies passed on to calves via colostrum. This resistance gradually deteriorates, leaving the animal vulnerable to disease (Fadly 2012).
- Regarding breed, *Bos taurus* is more susceptible to babesia infection than *Bos indicus* (Radostits et al., 2007). Besides that, native breeds have higher resistance to babesiosis than foreign breeds. Because tick populations have been exposed to nature for a long time, they have developed either an innate ability or an innate resistance to progress a good immune system to the tick (Wodaje et al. 2019).
- In endemic areas, young animals can acquire passive immunity from dams via colostrum and often suffer the transient infections with mild symptoms. This infection is enough to activate active immunity and make the host a carrier for a long time. Active immunity is in charge of the carrier's persistence and premunity. These animals can be infected naturally or through chemotherapy and still have a strong immune system (Taylor et al. 2007). According to susceptibility to *B. bovis* infection, *Bos taurus* were classified into three phenotypes: 1- susceptible animals which may experience clinical signs leading to death, 2- animals having mild clinical signs, and 3- animals that are resistant and having few clinical signs (Benavides and Sacco 2007).

Pathogen Factor

Pathogenicity varies greatly depending on the strain. Because of the wide variety of strains, *B. bovis* is typically more virulent than *B. bigemina* and *B. divergens* (CFSPH 2008). Through rapid antigenic variation, various blood parasites can keep the host immune system alive (Bock et al. 2004).

Environmental Factors

The prevalence of clinical babesiosis can be varied according to seasonal variation, which also influenced by the peak of

tick population. The largest prevalence occurring directly after the summit of the population of the tick. Regarding weather conditions, temperature is the most crucial factor affecting on the activity of the tick. Increase in temperature can cause the increase of the disease happenings (Menshawy et al. 2018). Cattle infection reaches the top in the summer season (El Moghazy et al. 2014; El-Bahy et al. 2018). Main economic losses happen in those places where marginal occurrence of disease is present because the population of the tick is mostly variable according to the conditions of environment (Radostits et al. 2007; Demessie and Derso 2015).

Transmission

Babesia species are biologically transmitted by vectors via transovarian transmission (first generation) and transsarial transmission (transmission of infection from egg until the adult) (Demessie and Derso 2015; Enbiyale et al. 2018). Babesiosis can be transmitted to cattle by a biological tick vector (*Boophilus* spp.). *Boophilus* ticks can transmit both *B. bigemina* and *B. bovis*, with nymphs and adults transmitting *B. bigemina* but only tick larvae transmitting *B. bovis* (Esmail et al. 2015). It is also mechanically transmitted by infected needles and syringes, blood transfusion, and surgical instruments (Menshawy 2020). *R. micropuls* (formerly *Boophilus micropuls*) and *R. annulatus* are tick vectors of *B. bigemina* (formerly *Boophilus annulatus*). Competent vectors include *R. decoloratus*, *R. geigy*, and *R. evertsi*. *R. micropuls* and *R. annulatus* are tick vectors of *B. bovis*, and *R. geigy* can also act as its competent vector (Bock et al. 2004; De Vos and Potgieter 2004; Yadhav et al. 2015). Transplacental transmission of babesia species in cattle has also been demonstrated (De Vos and Potgieter 2004; Spickler and Anna Rovid 2016).

The Babesia species can develops and distribute throughout the organs of the ticks, infecting the salivary glands or eggs. When infected tick bites a cattle, it transferred the infection to the final host (Government and State agencies bord 2013).

Pathogenesis

There are two principal mechanisms of producing acute disease by babesia which are hemolysis and circulatory disturbance (Carlton and McGavin 1995). Sporozoites enter the host directly after tick bite and infect the erythrocytes. Within the body of the host, sporozoites will then progress into piroplasm inside the infected RBCs. This will produce 2 or 4 daughter cells and they will then leave the host cell to infect other RBCs (Hunfeldt et al. 2008). They will invade other erythrocytes and can cause intravascular and extravascular hemolysis (Carlton and McGavin 1995). The rapid division of the parasite in the cells can cause rapid destruction and then haemoglobinaemia, hemoglobinuria, and fever. This can be very acute and cause death in a few days. During this process, the PCV falls to less than 20% and this will cause anemia. Clinical signs can be detected during

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the stage of parasitemia. At this stage, up to 45% of the red cells are infected according to *Babesia* species (Urquhart et al. 1996). Hemolysis also involves the release of many pharmacologically active agents (ex: proteolytic enzyme), which affect microcirculation (vasodilation, increased permeability) leading to hypotension and edema, and affect blood (viscosity, coagulation and adherence) leading to ischemia (congestion and degeneration change in tissue/organ) (Ahmed 2002). The main consequence of the disease is anemia due to hemolysis. The secondary mechanism is electrolyte imbalance. Liver and kidney degeneration are caused by lack of oxygen and perhaps by immune pathologic reaction. The kidney tubule epithelium damage will lead to impair ion exchange, which will result in hydrogen ion retention and cause acidosis (Enbiyale et al. 2018).

Clinical Signs

Incubation period ranges between eight and fifteen days in natural infection. Before the onset of other clinical signs fever ($>40^{\circ}\text{C}$) usually appears (OIE 2010). The clinical signs are different according to the age and species of the animals, parasite strain, immunological status, concurrent infection with other pathogens, and genetic factors in the dose of the inoculated parasites. Most cases have been detected in animals less than 9 months of age usually staying asymptomatic (Anon 2008).

Babesiosis clinical signs include emaciation, ataxia, loss of appetite, stop rumination, loss of body weight, progressive hemolytic anemia, jaundice (Icterus), yellowish color of conjunctival as well as vaginal mucous membranes in more advanced cases; hemoglobinuria, problems in the heart and respiratory rates, and a decrease in milk yield. In some cases, fever during an infection causes abortion in cattle. Patients experience general circulatory shock and, in some cases, nervous symptoms due to the sequestration of the infected RBCs in cerebral capillaries (Zintl et al. 2003; Khan et al. 2004; Akande et al. 2010; Chaudhry et al. 2010; Rashid et al. 2010; Terkawi et al. 2011; Onoja et al. 2013; El Moghazy et al. 2014; Bhat et al. 2015; Masih et al. 2021).

Dark red urine is one of the clinical signs of babesia (Yadav et al. 2004). The main clinical signs of *B. bigemina* are fever, hemoglobinuria, and anemia (Zintl et al. 2013).

Diagnosis

Detection of active cases of babesiosis is based mainly on several diagnostic techniques as follow:

Microscopic Examination

The conventional model of babesiosis examination is a direct examination under a microscope. It is used to identify the agent in the infected host. This is accomplished by examining

thick and thin films and then staining them with Giemsa or Romanowsky stain. Thick films can detect parasites as few as one parasite out of 106 RBCs (Kahn 2005). Microscopic examination is still the most cost-effective and time-efficient technique for identifying *Babesia* parasites (Hamoda et al. 2014). Giemsa-stained thin blood smears are the traditional and gold standard for identification (Nayel et al. 2012) and serve as an ideal method for species differentiation. It is adequate for detecting acute infections but has lower effects in cases of low parasitemia in carriers (Criado-Fornelio et al. 2009; Bal et al. 2016; Shang et al. 2016; Masih et al. 2021).

Serological Examinations

To detect antibodies in subclinical cases and avoid the drawbacks of microscopic examination, the Indirect Fluorescent Antibody Test (IFAT) and Enzyme-Linked Immunosorbent Assay (ELISA) are used (El-Fayomy et al. 2013). These tests have low sensitivity and frequently fail to distinguish between chronic and acute infections (Mahmoud et al. 2016). These tests produce false-positive and false-negative results due to cross-reactive antibodies (Esmaeil et al. 2015). Another point to consider is that antibodies persist even months after infection, implying that no active infection exists. As a result, these will be unable to reveal the precise prevalence at a given time (Abdel Aziz et al. 2014). The most common test for detecting antibodies in babesia species is IFAT (Chaudhry et al. 2010). Anonymous (2008) described a complement fixation (CF) test for detecting antibodies to *B. bovis* and *B. bigemina*.

Molecular Diagnosis

Molecular diagnosis is used to identify nucleic acids which is considered as an indirect identification. However, both sensitivity and specificity are very high (Mosqueda et al. 2012). The most sensitive and specific technique for the detection of babesiosis is (PCR) Polymerase chain reaction (Vannier and Krause 2009; AbouLaila et al. 2010) and useful for the detection of infection in the early stage. It has been reported that the PCR technique is much more sensitive than microscopy for the identification of babesiosis. It is an important test for confirmation in some cases for regulatory testing (Shams et al. 2013; Sharma et al. 2016; Bal et al. 2016).

Differential Diagnosis

Like many other infectious diseases, babesiosis also causes fever and anemia. Anaplasmosis, theileriosis, trypanosomiasis, leptospirosis, rapeseed poisoning, and chronic copper poisoning can be counted as a differential diagnosis of babesiosis. Rabies and other encephalitis's can also be considered in cattle with CNS signs (Spickler and Anna Rovid 2016).

Treatment

The successful treatment of babesiosis is dependent on the use of effective drugs and early detection (Vial and Gorenflot 2006). Trypan blue, which was first used against *B. bigemina* but has no effect on *B. bovis*, was one of the most effective drugs used to treat bovine babesiosis. It is rarely used because it discolors the flesh of animals. In the tropics, diminazene aceturate is currently used as a babesiacide. It has been withdrawn from the market in Europe for marketing reasons (Sayin et al. 1997). Imidocarb, which is primarily used in animals, is another effective drug for treating babesiosis. This drug can also be used to prevent babesiosis and anaplasmosis. Imidocarb can linger in tissues for a long time (Hashem et al. 2018) However, acridine and quinuronium derivatives can be used as effective drugs as well. Many European countries used the babesiacides quinuronium sulfate, amicarbalide, diminazene aceturate, and imidocarb dipropionate against bovine babesiosis for several years, but quinuronium sulfate and amicarbilide were withdrawn due to manufacturing safety issues (Vial and Gorenflot 2006). The combination of imidocarb dipropionate and oxytetracycline is the most effective treatment for Babesiosis in small ruminants (Ijaz et al. 2013). Beside this, in severe cases, supportive therapy is also required (Zintl et al. 2013). Vitamin E can also be used as a supportive therapy because it reduces the oxidative effect of babesia by increasing antioxidant activity (Abdel Hamid et al. 2014).

Prevention and Control

In the world, several countries have not completely controlled bovine babesiosis, despite the availability of live attenuated vaccine (De Vos and Bock 2000; Florin-Christensen et al. 2014). This can confirm the quick action for crucial vaccines to prevent the development of acute disease as well as parasite distribution into non-endemic areas. Bovine babesiosis control is currently under threat because of climate changes that act on vector development and expansion (Dantas-Torres 2015; Sonenshine 2018).

Control of this disease is created by accurate diagnosis, perfect treatment, and prevention of babesiosis (Mylonakis 2001). Animals after recovering from infection remain immunized. The parasite can persist in the peripheral blood for several years in *B. bovis* cases and for many months in *B. bigemina*, and no signs are apparent during this carrier state, so the animal should be monitored and treated after infection to prevent the distribution of disease to other animals (El Sawalhy 1999). Prevention and control of babesiosis can actively be maintained by the following methods: immunization, chemoprophylaxis, and vector control (Suarez and Noh 2011; ILRAD 1991). The combination of these three methods is also a choice. Tick control by vaccination has been stated as a useful way in Australia (Lightowlers 2013). A research has reported that using

combined chemotherapeutics is more effective for parasite elimination and results in decreasing the risk of drug resistance (Pritchard et al. 2013). The advantages of mixing of the chemotherapeutics include highly effectiveness, reduction in the dose (which may lead to reduced side effects) and lowering of drug resistance. According to the US reports, Babesiosis can be controlled and eradicated by eliminating the host tick(s). This will be done by using acaricides every two to three weeks. In those countries where eradication is not applicable, tick control can reduce the incidence of disease (APHIS 2010). Chemotherapy is another important method for controlling bovine babesiosis, either in the field or to control artificially induced infections. Chemotherapy is critical in some parts of the world to eradicate and prevent babesiosis. Infected animals should be treated with antiparasitic drugs as soon as possible in countries where the disease is endemic. The success of disease treatment is dependent on early diagnosis and proper administration of the drug of choice (Fernandez and White 2010; Georgiou et al. 2015). Use of living attenuated vaccine is the cornerstone to control and prevent babesiosis in many countries like Argentina, Israel, and Australia. However, this live vaccine is not cheap to produce and has many limitations (Brown et al. 2006; Florin-Christensen et al. 2014; Costamagna et al. 2016; Aranda et al. 2017; Suarez et al. 2018). Vaccines are provided in frozen form. Live babesia vaccines are not completely safe. A single dose can immunize animals against babesiosis over life (Saad et al. 2015).

Immunization of the animals in a prophylactic way has been stated as the most efficient way to decrease losses happened by bovine babesiosis. Live attenuated vaccine from the *B. bovis* or *B. bigemina* strain is used to immunize cattle in many countries. These vaccines are important due to having safety issues such as the potential effect for virulence in adult animals, contamination possibly occurring with other etiological agents, and blood protein hypersensitivity reactions (OIE 2015).

Conclusion

Babesiosis is a severe disease not only in cattle and other domestic and wild animals but also in human beings. It has significant impacts on both the economic and medical processes. It can cause impairment in the trade of animal products such as milk, meat, and hide by decreasing their quality. It has been reported that imidocarb and diminazene aceturate used as a treatment of babesiosis for many years, but nowadays, several compounds are progressed and assessed as a treatment. This can offer a good point for disease control. Controlling tick-borne diseases is important in developing livestock health services products. Control strategies can be different from country to country and place to place and the most important ones are vaccines and drugs.

Recommendations

- Knowledge, as well as awareness, should be given to the owners about the transmission way, prevention, and control of babesia.
 - Governments and organizations should give attention to control and eradicate babesiosis in order to improve the economy.
 - The surveillance system is important in Kurdistan Region to prevent bovine babesiosis.
- New drugs and vaccines should be developed to eradicate the carrier states.

REFERENCES

- Abdel Aziz KB et al., 2014. Molecular characterization of babesiosis infected cattle: Improvement of diagnosis and profiling of the immune response genes expression. *Global Veterinaria* 12(2): 197-206.
- Abdel Hamid OM et al., 2014. Biochemical changes associated with Babesiosis infested cattle. *Journal of Applied Chemistry* 7: 87-92.
- Abdela N and Jilo K, 2016. Bovine Babesiosis and its Current Status in Ethiopia: A Systemic Review. *Advances in Biological Research* 10(3): 138-146.
- AbouLaila M et al., 2010. Development and evaluation of two nested PCR assays for the detection of *Babesia bovis* from cattle blood. *Veterinary Parasitology* 172: 65-70.
- Afridi ZK and Ahmad I, 2005. Incidence of anaplasmosis, babesiosis and theileriosis in dairy cattle in Peshawar [Pakistan]. *Sarhad Journal of Agriculture* 21: 311-316.
- Ahmed J, 2002. The role of cytokines in immunity and immunopathogenesis of piroplasmoses. *Parasitology Research* 88: 48-50.
- Akande F et al., 2010. Haemoparasites of cattle in Abeokuta, south west Nigeria. *Science World Journal* 5: 19-21.
- Allsopp M et al., 1994. Phylogeny and evolution of the piroplasm. *Parasitology* 108: 147-152.
- Alvarez JA et al., 2019. Diagnostic Tools for the Identification of *Babesia* sp. in Persistently Infected Cattle. *Pathogens* 8: 143.
- Anon, 2008. The center for food security and public health of bovine babesiosis.
- APHIS, Veterinary Services 2010. Controlling Cattle Fever Ticks; Factsheet; USDA, APHIS, Veterinary Services; National Center for Import/Export; Animals Program
- Aranda FD et al., 2017. A discrete epidemic model for bovine Babesiosis disease and tick populations. *Open Physics* 15: 360-369.
- Bal MS et al., 2016. Diagnosis and management of bovine babesiosis outbreaks in cattle in Punjab state. *Veterinary World* 9(12): 1370-1374.
- Benavides MV and Sacco AM, 2007. Differential *Bostaurus* cattle response to *Babesia bovis* infection. *Veterinary Parasitology* 150: 54-64.
- Beugnet F and Moreau Y, 2015. Babesiosis. *Revue scientifique et technique (International Office of Epizootics)* 34: 627-639.
- Bhat SA et al., 2015. Molecular detection of *Babesia bigemina* infection in apparently healthy cattle of central plain zone of Punjab. *Journal of Parasitology and Diseases* 39: 649-653.
- Bock R et al., 2004. Babesiosis of cattle. *Parasitology* 129: 247-269.
- Brown WC et al., 2006. Immune control of *Babesia bovis* infection. *Veterinary Parasitology* 138: 75-87.
- Carlton WW and McGavin MD, 1995. Thomson's Special Veterinary Pathology. 2nd Ed., Mosby 2nd Year Book Incorporated, USA.
- Center for Food Security and Public Health (CFSPH), 2008. Bovine babesiosis, Iowa State University, Ames, Iowa.
- Chaudhry Z et al., 2010. Molecular detection of *Babesia bigemina* and *Babesia bovis* in crossbred carrier cattle through PCR. *Pakistan Journal of Zoology* 42: 201-204.
- Chowdhury S et al., 2006. Occurrence of common blood parasites of cattle in Sirajgonj Sadar area of Bangladesh. *Bangladesh Journal of Veterinary Medicine* 4: 143-145.
- Costamagna A et al., 2016. A model for the operations to render epidemic free a hog farm infected by the Aujeszky disease. *Applied Mathematics and Nonlinear Sciences* 1(1): 207-228.
- Criado-Fornelio A et al., 2003. Molecular studies on *Babesia*, *Theileria* and *Hepatozoon* in southern Europe. Part II. Phylogenetic analysis and evolutionary history. *Veterinary Parasitology* 114: 173-194.
- Criado-Fornelio A et al., 2009. Development of fluorogenic probebased PCR assays for the detection and quantification of bovine piroplasmids. *Veterinary Parasitology* 162: 200-206.
- Dantas-Torres M, 2015. Climate Change, Biodiversity, Ticks, and Tick-Borne Diseases, the Butterfly Effect. *International Journal for Parasitology: Parasites and Wildlife* 4(3): 452-461.
- De Vos AJ and Potgieter FT, 2004. Bovine babesiosis. In: Coetzer JAW, Tustin RC, editors. *Infectious diseases of livestock* (2nd Ed.). Cape Town: Oxford University Press; pp: 406-424.
- De Vos A and Bock R, 2000. Vaccination against bovine babesiosis and anaplasmosis. *Academic science* 916: 540-545.
- Delilah Caldwell, 2006. Tick-borne diseases of Cattle.
- Demeke D et al., 2018. Review on bovine Babesiosis. *Acta Parasitologica* 9(1): 15-26.
- Demessie Y and Derso S, 2015. Tick Borne Hemoparasitic Diseases of Ruminants: A Review. *Advances in Biological Research* 9(4): 210-224.
- El Moghazy HM et al., 2014. Epidemiological studies on bovine babesiosis and Theileriosis in Qalubia Governorate. *Benha Veterinary Medical Journal* 27(1): 36-48.
- El Sawalhy A, 1999. *Veterinary Infectious Diseases*. Ahram Distribution Agency, Egypt.
- El-Bahy NM et al., 2018. Molecular detection of *Babesia bigemina* and *Babesia bovis* in cattle in Behaira Governorate. *European Journal of Pharmaceutical and Medical Research* 5(12): 441-446.
- El-Fayomy AO et al., 2013. Contribution of *Babesia* to the illness of cows in Port Said Governorate, Egypt. *Global Veterinaria* 11(1): 118-122.
- Enbiyale G et al., 2018. Review on Bovine Babesiosis. *Acta Parasitologica Globalis* 9(1): 15-26.
- Esmail N et al., 2015. Determination of prevalence and risk factors of infection with *Babesia ovis* in small ruminants from west Azerbaijan province, Iran by Polymerase chain reaction. *Journal of Arthropod-Borne Disease* 9: 246-252.
- Esmailnejad B et al., 2015. Determination of prevalence and risk factors of infection with *Babesia ovis* in small ruminants from west Azerbaijan province, Iran by Polymerase chain reaction. *Journal of Arthropod-Borne Disease* 9: 246-252
- Fadly RS, 2012. Prevalence of some blood parasites of some farm animals at Behera Province. *Assiut Veterinary Medical Journal* 58(134): 316-322.

- Fakhar M et al., 2012. An epidemiological survey on bovine and ovine Babesiosis in Kurdistan Province, western Iran. *Tropical Animal Health and Production* 44: 319-322.
- Fernandez PJ and White WR, 2010. Atlas of transboundary animal diseases OIE: 2010.
- Florin-Christensen M et al., 2014. Vaccines against bovine babesiosis: where we are now and possible roads ahead. *Parasitology* 141: 1563-1592.
- Gallego-Lopez MG et al., 2019. Review Interplay between Attenuation- and Virulence-Factors of *Babesia bovis* and Their Contribution to the Establishment of Persistent Infections in Cattle. *Pathogens* 8(97): 1-13.
- Gohil S et al., 2013. Bovine babesiosis in the 21st century: advances in biology and functional genomics. *International Journal for Parasitology* 43: 125-132.
- Government and State Agencies Bord, 2013. Bia Department of Agriculture Food and the Marine (DAFM). Teagasc Parasite Control Leaflet Series 8: 1-25.
- Gray J et al., 2010. Zoonotic babesiosis: overview of the disease and novel aspects of pathogen identity. *Ticks and Tick-Borne Diseases* 1: 3-10.
- Gray JS et al., 2019. Vectors of Babesiosis. *The Annual Review of Entomology* 64: 149-165.
- Gubbels J et al., 1999. Simultaneous Detection of Bovine Theileria and *Babesia* Species by Reverse Line Blot Hybridization. *Journal of clinical microbiology* 37: 1782-1789.
- Guswanto A et al., 2017. Molecular and serological detection of bovine babesiosis in Indonesia. *Parasites and Vectors* 10: 550.
- Hamoda AF et al., 2014. Toxic effect of Babesiosis in cattle and chemotherapeutic treatment in Egypt. *American Journal of Infectious Diseases and Microbiology* 2: 91-96.
- Hamsho A et al., 2015. A Cross-Sectional Study of Bovine Babesiosis in Teltele District, Borena Zone, Southern Ethiopia. *Journal of Veterinary Science and technology, Health Production* 29: 11-15.
- Hashem MA et al., 2018. A study on Bovine Babesiosis and Treatment with Reference to Hematobiochemical and Molecular Diagnosis. *Slovenian Veterinary Research* 55 (20): 165-173
- Hunfeld KP et al., 2008. Babesiosis: recent insights into an ancient disease. *International Journal for Parasitology* 38: 1219-1237.
- Ijaz M et al., 2013. Clinico-epidemiology and therapeutical trials on Babesiosis in Sheep and goats in Lahore, Pakistan. *The Journal of Animal and Plant Sciences* 23: 666-669.
- International Laboratory for Research on Animal Diseases (ILRAD), 1991. Recent developments in the control of Anaplasmosis, Babesiosis and Cowdriosis, Proceedings of a workshop held at Nairobi, Kenya, pp: 1-174.
- Jabbar A et al., 2015. Tick- borne diseases of bovines in Pakistan: major scope for future research and improved control. *Parasites and Vectors* 8: 283.
- Jerram L and Willshire J, 2019. Babesiosis in the UK and approach to treatment. *Livestock* 24: 18-24.
- Kaandorp S, 2004. Transmissible diseases handbook. IDWG Secretariat.
- Kahn C, 2005. The Merck Veterinary Manual, 9th Ed., Merck and Company Incorporated, USA.
- Khan M et al., 2004. Prevalence of blood parasites in cattle and buffaloes. *Pakistan Veterinary Journal* 24: 193-195.
- Kuttler KL, 1980. Pharmacotherapeutics of drugs used in treatment of anaplasmosis and babesiosis. *Journal of the American Veterinary Medical Association* 176: 1103-1108.
- Lauren A et al., 2016. Radical cure of experimental babesiosis in immunodeficient mice using a combination of an endochin-like quinolone and atovaquone. *Journal of Experimental Medicine* 213(7): 1307-1318. doi: <https://doi.org/10.1084/jem.20151519>
- Lefevre PC et al., 2010. Infectious and parasitic diseases of livestock. Lavoisier.
- Lightowers M, 2013. Antiparasitic vaccines. In: Dwight D, editor. Bowman Georgis' Parasitology for Veterinarians 10th Ed. by Saunders, an imprint of Elsevier Inc. ISBN-10: 1455740063
- Mahmoud MM et al., 2016. Molecular detection of babesia infection in young calves in Damietta governorate, Egypt. *Global Journal of Animal Scientific Research* 4(2): 185-193.
- Mandal S, 2012. Veterinary Parasitology, India: Panacea Computer.
- Masih A et al., 2021. Molecular Epidemiology of Bovine Babesiosis in Punjab, Pakistan. *Acta Scientiae Veterinariae* 49: 1809.
- McCosker PJ, 1981. The global importance of babesiosis. In: Ristic M, Kreier JP, editors. Babesiosis: Academic Press, New York; pp: 1-24.
- Menshawy SM et al., 2018. Dynamics of Boophilus Ticks and its Role in Transmission of Piroplasms at Behaira District. *Alexandria Journal of Veterinary Sciences* 56(1): 137-144.
- Menshawy SM, 2020. A Review on Bovine Babesiosis in Egypt. *Egyptian Veterinary Medical Society of Parasitology Journal (EVMSPJ)* 16: 8-19.
- Mezouaghi A et al., 2019. A predictive spatio-temporal model for bovine Babesiosis epidemic transmission. *Journal of Theoretical Biology* 11: 1-27.
- Mosqueda J et al., 2012. Current advances in detection and treatment of Babesiosis. *Current Medicinal Chemistry* 19: 1504-1518.
- Mosqueda J et al., 2012. Current advances in detection and treatment of Babesiosis. *Current Medicinal Chemistry* 19: 1504-1518.
- Mylonakis E, 2001. When to suspect and how to monitor babesiosis. *American family physician* 63: 1969.
- Onoja II et al., 2013. Prevalence of Babesiosis in cattle and goats at Zaria Abattoir, Nigeria. *Journal of Veterinary Advances* 3: 211-214.
- Otify YZ, 2011. Veterinary Parasitology (Arabic language), 2nd Ed. Soot Elkalam El-Araby, Egypt.
- Ozubek S et al., 2020. Review: Bovine Babesiosis in Turkey: Impact, Current Gaps, and Opportunities for Intervention. *Pathogens* 9 (1041): 1-23.
- Pohl A, 2013. Epidemiology study of tick-borne diseases in cattle in Minas Gerais. *Journal of Veterinary Advances* 40: 124-150
- Pritchard JR et al., 2013. Defining principles of combination drug mechanisms of action. *Proceedings of the National Academy of Sciences of the United States of America* 110: 170-179.
- Radostits O et al., 2007. Veterinary Medicine: A textbook of the disease of cattle, sheep, goat, pigs and horses. 10th Ed., Saunders Elsevier, London, UK.
- Radostits OM et al., 2006. Veterinary Medicine E-Book: A textbook of the diseases of cattle, horses, sheep, pigs and goats, Elsevier Health Sciences.
- Rashid A et al., 2010. Prevalence and chemotherapy of babesiosis among Lohi sheep in the Livestock Experiment Station, Qadirabad, Pakistan. *The Journal of Venomous Animals and Toxins including Tropical Diseases* 16: 587-591.
- Rozej-Bielicka W et al., 2015. Human babesiosis. *Przegląd epidemiologiczny* 69: 605-608.
- Saad F et al., 2015. Zoonotic significance and prophylactic measure against babesiosis. *International Journal of Current Microbiology and Applied Sciences* 4: 938-953.

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- Sahinduran S, 2012. Protozoan diseases in farm ruminants. In: Perez Marin C, editor. *A Bird's Eye View of Veterinary Medicine: International Journal of Technology*; pp: 473 - 477.
- Saunsoy R, 1995. Livestock-a driving force for food security and sustainable development. *World Animal Review* 84(85): 5-17.
- Sayin F et al., 1997. Tick-borne diseases in Turkey. *Tropical Animal and Health Production* 29: 1.
- Schnittger L et al., 2012. Babesia: A world emerging. *Infection, Genetics and Evolution* 12: 1788-1809.
- Shams S et al., 2013 Sensitivity and specificity of PCR & microscopy in detection of Babesiosis in domesticated cattle of Khyber Pakhtunkhwa, Pakistan. *International Journal of Advanced Research and Technology* 2(5): 37-41.
- Shang B et al., 2016. An evaluation of quantitative PCR assays (TaqMan and SYBR Green) for the detection of Babesia bigemina and Babesia bovis, and a novel fluorescent-ITS1-PCR capillary electrophoresis method for genotyping B. bovis isolates. *Veterinary Science* 3: 23-38.
- Sharma A et al., 2016. Clinicopatho-biochemical alterations associated with subclinical babesiosis in dairy animals. *Journal of Arthropod-Borne Diseases* 10(2): 259-267.
- Silva MG et al., 2010. Detection of Babesia and Theileria species infection in cattle from Portugal using a reverse line blotting method. *Veterinary Parasitology* 174(3-4): 199-205.
- Simunza MC, 2009. Differential Diagnosis of Tick-borne diseases and population genetic analysis of Babesia bovis and Babesia bigemina. PhD Dissertation, University of Glasgow.
- Skotarczak M, 2008. Babesiosis as a disease of people and dogs. Molecular diagnostics. *Veterinarni Medicina* 53: 229-235.
- Sonenshine ED, 2018. Range Expansion of Tick Disease Vectors in North America: Implications for Spread of Tick-Borne Disease. *International Journal of Environmental Research and Public Health* 15(3): 478.
- Soulsby EJ, 1986. Helminths, Arthropods and Protozoa of Domesticated Animals, 7th Ed., Bailliere Tindall, London, UK.
- Spickler A et al., 2010. Emerging and exotic diseases of animals, 4th edition CFSPH Iowa State University, Iowa, USA.
- Spickler and Anna Rovid, 2016. Bovine Babesiosis; fact sheet, The Center for Food Security & Public Health: <http://www.cfsph.iastate.edu/DiseaseInfo/disease.php?name=bovine-babesiosis> accessed 1/01/2016.
- Suarez CE and Noh S, 2011. Emerging perspectives in the research of bovine babesiosis and anaplasmosis. *Veterinary Parasitology* 180: 109-125. doi: 10.1016/j.vetpar.2011.05.032.
- Suarez CE et al., 2018. Unravelling the cellular and molecular pathogenesis of bovine babesiosis: is the sky the limit? *International Journal for Parasitology* 49(2): 183-197.
- Taylor M, 2007. *Veterinary Parasitology*, 3rd Ed., Blackwell Publishing, USA.
- Terkawi MA et al., 2011. Molecular and serological prevalence of Babesia bovis and Babesia bigemina in water buffaloes in the northeast region of Thailand. *Veterinary Parasitology* 178: 201-207.
- Uilenberg G, 2006. Babesia-A historical overview. *Veterinary Parasitology* 138: 3-10.
- Urquhart G et al., 1996. *Veterinary Parasitology*, Blackwell Science Ltd., UK.
- Vannier E and Krause PJ, 2009. Update on babesiosis. *Interdisciplinary perspectives on infectious diseases*.
- Vial H and Gorenflot A, 2006. Chemotherapy against babesiosis. *Veterinary parasitology* 138: 147-160.
- Wagner GG et al., 2002. Babesiosis and heat water: threats without boundaries. *The Veterinary Clinics of North America Food Animal Practice* 18(3): 417-430.
- Wodaje A et al., 2019. A Review on Bovine Babesiosis. *International Journal of Advanced Research in Biological Sciences* 6(1): 63-70.
- World Organization for Animal Health: OIE Terrestrial Manual. Manual of Diagnostic tests and vaccines for terrestrial animals 2015. Accessed on line. <http://www.oie.int/international-standardsetting/terrestrialhttp://www.oie.int/international-standard-setting/terrestrial-manual/accessonline/manual/access-online/>.
- World Organization for Animal Health: OIE, 2010. Bovine babesiosis, chapter 2 at <http://web.oie.int/eng/norms/mmanual/2008/pdf//2.04>.
- Yadav HS et al., 2004. A study on babesiosis in HF cattle. *Journal of Veterinary Parasitology* 18: 151-153.
- Yadhav C et al., 2015. An overview of Babesiosis. *International Journal of Pharmaceutical Sciences and Research* 3: 287-295
- Yusuf JJ and Jimma P, 2017. Review on bovine babesiosis and its economic importance. *Journal of Veterinary Medicine and Research* 4: 1090.
- Zintl A et al., 2003. Babesia divergens, a bovine blood parasite of veterinary and zoonotic importance. *Clinical Microbiology Reviews* 16: 622-36.
- Zintl A et al., 2013. Babesia divergens, a bovine blood parasite of veterinary and zoonotic importance. *Clinical microbiology reviews* 16: 622-636