

Trichinellosis: A Hidden Threat in Meat Consumption



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ABSTRACT

Trichinella spiralis, the roundworm that causes trichinellosis, is a major global health issue. The main way that the disease is spread is by eating raw or undercooked meat from infected livestock, and wild boars are increasingly playing a role in spreading epidemics. Numerous symptoms, such as myocardial infarction, stomach discomfort, and neurological involvement, are present with the condition. Trichinella species have a complicated life cycle that includes an enteral phase, and a migratory. Trichinellosis is not as common worldwide, it is still a cause for concern, particularly in less developed nations where eating raw or undercooked meat is common. Trichinella species are distributed differently over the world, with T. spiralis being more common in Europe. phase, and a muscle phase. This causes tissue damage and severe inflammation. Due to the small size of larvae and limits in testing methods, difficulties in recognizing contaminated meat continue even in the absence of recorded cases. Trichinellosis vaccines are being developed using a variety of techniques, including DNA, synthetic peptide, live attenuated, and recombinant protein vaccines. The selection of antigens, adjuvants, and variations in immune responses among animal species present challenges in the production of vaccines. Future work should concentrate on developing genetic engineering tools, and comprehending immune evasion mechanisms.

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1. INTRODUCTION

Trichinellosis is a parasitic infection caused by Trichinella spiralis (T. spiralis) (roundworm) (Wu et al. 2022). Through the consumption of contaminated meat, primarily through hunting or scavenging of meat from an infected animal, Trichinella spp. are transmitted to and survive in a variety of hosts (Sgroi et al. 2023). Trichinella can infect more than 150 different species of animals and humans. When consumers eat undercooked or raw meat that has Trichinella infective larvae, they become infected (Hady et al. 2023). Wild animals that are omnivorous and carnivorous serve as the natural reservoirs for Trichinella spp. Trichinella species in domestic and wild animals are not necessarily linked to human diseases (Murakami et al. 2023). The host's dietary habits play an essential role in transmission. The major source of human infection is pigs, particularly those reared in the backyard. However, for the past 30 years, wild boar meat has been a significant factor in epidemics. The worldwide distribution of Trichinella spp. and various cultural food patterns are the major factor that favors human infection in non-industrialized and industrialized countries (Pavel et al. 2023). Higher incidence is frequently observed where eating raw or undercooked meat from domestic animals and wild animals is common.

Compared to other foodborne parasites, Trichinella spp. the infection has a modest global burden. The global disability-adjusted life years (DALY) for trichinellosis were calculated at 76 per billion people per year, yet in recent years, 5751 cases and 5 mortalities have been recorded in 55 countries (Rostami et al. 2017). T. spiralis, a historically acquired species from Eastern Asia, is common in Spain, Poland, Lithuania, and the Balkan different countries of Bulgaria, Romania, Serbia, and Bulgaria (Veronesi et al. 2023). From 1986 to 2009, there were 65,818 incidents reported worldwide, with 56,912 of those incidents occurring in Europe. In the past 20 years, trichinellosis cases have not been documented in nearly half of the EU nations, including Luxembourg, Cyprus, Portugal, and Malta (Pozio 2019). The European Union has identified 5518 cases of trichinellosis over the past 16 years (2002–2017), with a declining trend. In the genus Trichinella, 9 species and 3 genotypes have been identified (Table 1).

Species (genotype)	Larval form	Distribution	Pathogenicity to humans	Major hosts	References
T. spiralis (T1)	Encapsulated	Worldwide	High	Carnivores,	Bruschi and
				wild boar, pigs,	Dupouy-Camet
				rats	2014
T. nativa (T2)	Encapsulated	Europe, and America,	High	Dogs, wild	Bruschi et al. 2002
		areas of Asia, the Arctic	:	carnivores,	
		and Subarctic		Rare in pigs	
T. britovi (T3)	Encapsulated	Asia, middle east	High	Jackal, dog,	Foreyt and Abbott
		countries, Europe,		Wild Boar,	2013
T. pseudospiralis	Nonencapsulated	Australia, Thailand,	High	Birds and	Foreyt and Abbott
(T4)		Nearctic, and palearctic		Mammals	2013
		regions, New Zealand,			
T. murrelli	Encapsulated	Canada and USA	Moderate	Carnivores	Gottstein et al. 2009
T. nelsoni	Encapsulated	Southern Eastern-Africa	Low	Carnivores	Mitreva and Jasmer
					2006
Т. рариае	Nonencapsulated	Thailand, New Guinea,	Moderate	Reptiles and	Pozio 2007
		Рариа		Mammals	

 Table 1: Trichinella species, biological characteristics, and hosts and distribution.



T. zimbabwensis	Nonencapsulated South	Africa,	Ethiopia, Unknown	Reptiles	and Pozio 2001
	Mozambique, Zimbabwe,			Mammals	

2. THE LIFE CYCLE OF TRICHINELLOSIS

The life cycle of Trichinella spp. occurs when muscle tissue carrying first-stage larvae is consumed by the new host (Pozio 2022). The larvae move from the intestines to the lymphatic system, then to the circulation, where they enter skeletal muscle cells and become contagious to the next host. Severe inflammation is the main cause of disease and includes encephalitis, myositis, and myocarditis, the severity of which is determined by the amount of parasites consumed. *T. spiralis* has minimal host specificity in mammals, lives its entire life cycle in one host, lacks a free-living stage, and exists as an intracellular parasite inside a single striated muscle cell (Shinn et al. 2023).

3. ENTERAL PHASE

The infection is passed on by eating meat that is either uncooked or undercooked and carrying the nurse cell-larva combination. The columnar epithelium is at the base of the villus where the young parasites enter (Bonis et al. 2021). They are referred to as intra multi-cellular organisms since they reside there in a row of columnar epithelial cells. The larvae go through four rounds of molting before becoming adults. After mating, the young larvae pass using the bloodstream to the muscles that are regulated voluntarily, where they encyst (Nthiga 2022). Acute immune-mediated inflammation caused by the adult stage, which lives in the epithelial layer of the host's small intestine, results in physiological structural and cellular alterations. These changes are linked with notable changes in epithelial cells, the release of inflammatory mediators, and increasing in inflammatory cells (Muthumalage et al. 2019). A little infection causes minimal harm. Severe infection, on the other hand, can induce serosa petechiae, hyperemia increased, enlarged Peyer's patches, mucous secretion, and intestinal loop dilatation (Bandyopadhyay et al. 2022). In the jejunum, histopathology of the small intestine indicates an intense inflammatory reaction with diverse cellular infiltration primarily of neutrophils, lymphocytes, and eosinophils. T.spiralis can also produce trophic modifications in the longitudinal smooth and circular muscle layers of the ileum and jejunum with crypt hyperplasia and villous atrophy. These are correlated with significant changes in epithelial cells and an increase in mediators and inflammatory cell types (Robinson et al. 2019).

4. MIGRATORY PHASE

The pathology in the migratory phase is produced by larvae discharged into the intestinal mucosa, then migrate to the blood vessels. They transmit through the body until they reached the striated skeletal muscles. Transferring Trichinella larvae and their byproducts generate an instantaneous reaction, resulting in pathological, immunological, and metabolic abnormalities, as well as the different clinical manifestations seen during the acute stage of infection (Saha and Saroj 2022) shown in Fig. 1.

5. MUSCULAR PHASE

Larvae develop a major set of cell physiological changes after invading skeletal muscles. These modifications cause the completely differentiated muscle cell to convert into a nurse cell, helping in the development and growth of the larva (Huang et al. 2015). This stage is connected with allergic and inflammatory reactions generated by the invasion of the muscles by wandering larvae. This process can directly or indirectly harm muscle cells by encouraging the infiltration of inflammatory cells, particularly



eosinophils (Bruschi and Gómez-Morales 2014). In trichinellosis patients, there was a link between eosinophil levels with serum muscle enzymes, implying that these granulocytes may be involved in muscle injury (Bruschi and Dupouy-Camet 2022). Thus, increasing eosinophilia is the most important clinical finding of Trichinellosis muscular phase. The infiltration of the accessory muscles and digraph of respiration by the pathogen results in dyspnea.

6. NEUROLOGICAL INVOLVEMENT

Neurotrichinellosis can affect either white or gray matter of the brain, spinal cord, pons, and cerebellum (Tanabe et al. 2021). Damage to the central nervous system can occur directly or indirectly as a result of the release of tumor necrosis factor (TNF), immune-mediated processes, vascular damage, and toxin reactions which result in eosinophil toxicity. Trichinella larvae can move into the CNS and cause common lesions, blood vessel blockage, and inflammatory infiltrates (Garcia et al. 2019). Different pathologic changes may be mediated by larval and muscle breakdown components. The larvae either cause pathological symptoms in tissues before returning to circulation, or they can be trapped and destroyed, resulting in inflammatory reactions (Hady et al. 2023). Punctuate hemorrhages, hyperemia, and Edema occur commonly in the brain tissues. Vascular changes enclosing the larvae are thought to be the primary processes causing neurological injury.

7. CARDIAC INVOLVEMENT

The liver damage can be caused by larval damage directly or indirectly by immune reactions and eosinophils (Leal-Silva et al. 2021). The liver is usually enlarged in these situations due to dystrophic pathologies such as fatty degeneration. Hypoproteinemia is prevalent and can be attributed to hepatic dysfunction, allergic capillaropathy caused by eosinophils, and protein digestion and absorption deficits caused by changes in the intestinal mucosa (Wen et al. 2022). The decrease in total protein is predicated on a decrease in the albumin fraction.





Fig. 1: Life cycle of Trichinellosis (Retrieved from biorender).

8. SYMPTOMS AND SIGNS

The severity of symptoms is determined by the degree of infection and is proportional to the number of larvae per gram of muscle. Infections are divided into three types based on the number of larvae. In subclinical, up to 10 larvae are involved in light infections. In moderate 50 to 500 larvae are involved and in severe infection (Bruschi and Murrell 2020), more than 1000 larvae are involved which can be lethal. In symptomatic situations, symptoms appear in three stages: enteric (intestinal invasion), invasive (laser migration), and encystation in the muscles (Bogoch et al. 2021). The major clinical symptoms are myocardial infarction, abdominal pain, allergic reactions, encephalitis, fever, myalgia, intestinal diarrhea, and facial swelling (Dupouy-Camet et al. 2021).

9. CHALLENGES IN IDENTIFYING TRICHINELLA-INFECTED MEAT

To recognizing Trichinella-infected meat provides multiple challenges due to the parasite's microscopic size, the spread of larvae in the meat, and the limitations of accessible testing procedures. The complicated nature of these problems can make detecting Trichinella contamination in meat difficult, increasing the danger of consuming contaminated products (Chalmers et al. 2020). Trichinella larvae can be found in muscle tissues in comparatively small numbers, rendering ocular inspection insufficient to detect their existence. The larvae are spread throughout the flesh, therefore a tiny sample may not fully represent their distribution (Gabriël et al. 2022). Trichinella larvae are enclosed in cysts in muscular tissues, adding another layer of defense. Because the cysts shield the larvae from external variables like frying and freezing, reaching the larvae during testing is difficult (Álvarez-Guerrero and Alba-Hurtado 2011).

Due to the irregular distribution of Trichinella larvae in the meat, obtaining representative samples for testing can be difficult. A single tiny sample of meat may not adequately represent the full batch. Traditional diagnostic approaches, such as artificial digestion, necessitate a significant amount of time and specialized devices (Bergwerff and Debast 2021). These procedures may not be practicable for large-scale testing or in resource-constrained places (Yang et al. 2022). Some diagnostic procedures may have sensitivity limitations, particularly when it comes to detecting low amounts of Trichinella larvae in meat samples. This may result in false-negative results, underestimating the level of contamination. Rapid and accurate identification of Trichinella is critical for preventing diseased meat from the food supply chain (Thanh et al. 2014). Some methods of testing may take a long time, causing delays in finding contaminated meat. Some advanced testing procedures are costly and may not be feasible for periodic inspection of all beef products, particularly in resourceconstrained areas. There is a potential for cross-contamination between samples during testing, which could result in false-positive results or incorrect identification of contaminated meat (Haiminen et al. 2019).

10. TRICHINELLOSIS IN A GLOBAL ECONOMY

Global trade increases the risk of trichinellosis outbreaks from ready-to-eat pork foods, demanding urgent attention (Bintsis 2017). As a result, certain nations have unique rules for qualifying pigs or pork foods for importation. The European Union (EU) requires inspection of horse and pork meat before they can be imported into EU member countries. Despite these limitations, the eating of imported



horse meat has resulted in an upsurge of trichinellosis in the EU (Bruschi and Dupouy-Camet 2022). Outbreaks caused by consuming examined meat occur worldwide as well. The reason for the ineffective testing is probably due to inadequate quality control measures. The aggregated digestion assay, as recommended by the EU and others, is thought to be able to detect corpses with the smallest larvae load that would induce clinical sickness in humans (Thrastardottir et al. 2021). Use established protocols for meat inspection to ensure accuracy and reliability. To ensure precise test results, it is important to incorporate other elements of a quality assurance program, such as proficiency exchange, document management, critical point control analyst certification, sampling, and trace-back systems. Controlling Trichinella in cattle and food items globally is critical to reverse the pattern of emerging and re-emerging human trichinellosis. Alternative approaches to ensure Trichinella-free pork are now being examined in regions where pig infection has been almost completely eradicated and human trichinellosis is rare. Create efficient methods for managing farms to protect pigs from trichinellosis (Gamble 202). Rodent control and Bio-security are examples of sound management practices, as is the evasion of feeding garbage to pigs. Although the effectiveness of control in many nations, trichinellosis continues to trigger human disease in some areas, and the parasite's biology and epidemiology still need to be studied further to develop consistent, practical, and standardized control programs for all parts of the world.

11. VACCINES AGAINST TRICHINELLOSIS

The antigens for the *T. spiralis* vaccine are typically obtained from excretory-secretory products and basic extracts of whole worms (Zhang et al. 2018). It is widely accepted that inactivated and live attenuated vaccines are 1st generation vaccinations. Various approaches have been used to identify potential antigens for vaccines against trichinellosis, including immunoproteomics, genome, transcriptome, and proteome screening (Tang et al. 2022). Established on these techniques 2nd and 3rd generation vaccines have been accomplished in swine and rodents to investigate their shielding effects such as recombinant protein vaccine, DNA vaccine, and synthetic peptides (Khalid and Poh 2023).

12. RECOMBINANT PROTEIN VACCINES

Some development in the production of recombinant protein-based vaccines against *T. spiralis* disease has been accomplished with the quick progress of genetic engineering (Xu et al. 2020). Applicant antigens were mostly selected from functional proteins, ES products, and antigens implicated in *T. spiralis* attack pathways (Tang et al. 2022). The constituents that are crucial in ES products for *T. spiralis* infection are protease and protease inhibitors. To suppress *T. spiralis* infection, a great quantity of protein vaccine investigation has been conducted in recent years on deoxyribonuclease, serine proteases, cystatins, and serine protease inhibitors.

13. PROTEASES AND PROTEASES INHIBITOR

The serine proteases found in ES products assist *T. spiralis* in invading host cells and evading attacks from the immune system. The protein superfamily known as serine protease inhibitors is responsible for inhibiting the actions of serine proteases, and plays a part in inflammation, complement activation and blood coagulation (Sofronic-Milosavljevic et al. 2015). Worm serpins shield them from host serine proteolysis, helping parasites evade immune response. Mice immunized with recombinant rTsSP had 62.10 and 71.10% lower worm loads of ML and AD, respectively (Song et al. 2018). Mice vaccinated with rTspSP-1.2 had worm burdens of ML and AD reduced by 52.24 and 34.92 %, respectively. For pigs vaccinated with



rTs-Adsp, the worm load of ML was reduced by 50.9% (Tang et al. 2022). Mice immunized with recombinant rTsSPI had 57.25 and 62.2% decreased ML and AD worm loads (Grzelak et al. 2020). At ten days post-infection, animals inoculated with rTs-Serpin showed a 59.95% decrease in mature worms and a 46.41% decrease in larvae muscle (Xu et al. 2017b). The cystatin protein superfamily has the ability to inhibit the action of cysteine proteases. Cystatins serve a significant part in immune elusion and the regulation of the host immunological reaction during parasitic infection in nematodes (Stachyra, and Wesołowska 2023). At 5 days post-inoculation, mice immunized with a cystatin-like protein 64.28% drop in mature worms and 61.21% decrease in larvae muscle. Cysteine proteases are important enzymes that are existing in most living animals, including parasites and viruses. Parasitic cysteine proteases have a significant impact on the attack of host tissue and the survival of parasites within the host. As a result, they are a key focus for the growth of parasite vaccines (Stachyra et al. 2019).

14. DEOXYRIBONUCLEASE II

DNase II is found mostly in nuclei and lysosomes and plays an essential part in pathogen evasion and invasion of the host's immunological reaction (Kumari et al. 2020). T. spiral DNase II protein group is substantially larger than those of other species. Furthermore, investigations have revealed that *T. spiralis* DNase enzymes may play an important role in host-parasite contacts through infection, implying that they could be exploited as applicant antigens to regulate and avoid trichinellosis (Cui et al. 2019). Subcutaneously vaccinated mice with rTs-DNase II-7 and rTsDNase II-1 demonstrated 34.86 and 40.36 % decreases in mature worms at five dpi, respectively, 42.33 and 50.43 % decreases in larvae muscle (Tang et al. 2022). Pigs immunized with DNase II-7 demonstrated a 45.7% decrease in the larvae muscle (Xu et al. 2021). While recombinant protein vaccines are becoming more common, the degree of immunoprotection is linked to the adjuvants, antigens, and delivery routes.

15. LIVE ATTENUATED VACCINES

Live attenuated vaccines lower the risk of *T. spiralis* infection while still stimulating an immune response. Mice immunized with radiation-attenuated larvae showed a 72.5 percent reduction in the muscles of larvae (Hafez et al. 2020). Live attenuated vaccines have significant defensive effectiveness because these methods closely mimic natural infection and provide a similar environment to that of *T. spiralis* infection. Though, their security is called into question owing to the probability of infection. The use of live attenuated vaccine, which are known for their strong protective immunity, is quickly being phased out (Santi et al. 2018).

16. SYNTHETIC PEPTIDE VACCINE

Various epitope peptides have the advantage of existence easier and faster to make than recombinant protein vaccines, and they may contain numerous protective epitopes. Mice who received an immunization of a synthetic peptide made up of forty amino acids and derived from the glycoprotein of *T. spiralis* saw a decrease of 64.3% in adult worms (Wait 2022). Mice vaccinated with a thirty-mer peptide antigen had 33.3% decrease in parasite female fertility. In recent years, vaccines targeting epitopes for bacterial, viral, and parasite infections have been rapidly produced. Epitope vaccines have several drawbacks, such as low immunogenicity and the requirement to be linked to a large transport protein (Parvizpour et al. 2020). To boost the immunogenicity of epitope vaccines, a new method comprising numerous antigenic peptides was created. Epitope-based vaccinations can be developed as chimeric vaccines, by producing various efficient epitopes. By using a chimera vaccination, it is possible to enhance



the protection provided by epitope vaccines or prevent the immune system from being evaded by parasites (Sanches et al.2021). Furthermore, T. spiral life cycle is complicated, resulting in a variety of antigens at distinct phases. *T. spiralis* infection can be effectively controlled with a multiepitope vaccination.

17. DNA VACCINES

DNA vaccines acquired popularity because of their potential to elicit a wide immune reaction and provide long-time immunity (Soleymani et al. 2022). In addition, DNA vaccines have been found to be more steady, economical, easy to produce, and harmless to distribute when linked to traditional protein vaccines. The fundamental drawback of DNA vaccines over protein vaccines is their low immunogenicity (Qin et al. 2021). The primary disadvantage of DNA vaccines beyond protein vaccines is their lack of immunogenicity. Recently, numerous DNA vaccines effective against *T. spiralis* infection have been identified in mouse models. According to a study conducted on mice, who were given a TsPmy DNA vaccine delivered through Salmonella, there was a reduction of 46.6% and 44.8% in ML and AD worm burdens (Wu et al. 2021). In mice treated with the pcDNA3.1(+)-Ts-NBLsp DNA vaccine, the worm burden of ML was reduced by 77.93% (Xu et al. 2020). Overall, DNA vaccines have lower immunogenicity than protein vaccinations due to low amounts of antigen expression. According to research, DNA positive protein immunization is an excellent technique for increasing protective effect and immune response. Mice vaccinated with Ts87 in a DNA-prime/protein-boost approach had 46.1% reduction in ML worm load. More approaches will be developed to improve the efficacy of DNA vaccinations as technology advances. The principal objective is to create a DNA vaccination that can be used safely in humans.

18. SYNTHETIC PEPTIDE VACCINES

Extensive investigation has been conducted to progress vaccines against T. spiralis infection, including recombinant proteins, DNA vaccines, and crude antigens. Currently, only a limited amount of research has been conducted on the effectiveness of peptide vaccines in suppressing T. spiralis infection. Multiepitope peptides have the advantage of being easier and faster to make than recombinant protein vaccines, and they may contain numerous protective epitopes (Gu et al. 2020). Selected a forty-mer synthesized peptide from T. spiral glycoprotein, mice treated with the peptide vaccine had 64.3% decrease in adult worms. Screened a forty-mer synthesized peptide from T. spiral glycoprotein, and mice treated with the peptide vaccine had 64.3% decrease in adult worms. (Gu et al. 2020). Female parasite fecundity was reduced by 33.3% in mice inoculated with a thirty-mer peptide antigen. Lately, vaccines targeting the infection caused by viruses, bacteria, and parasites have been produced rapidly using epitope technology. However, epitope vaccines have numerous drawbacks, including low immunogenicity and the essential to be coupled to a larger transporter protein. To boost the immune response of epitope vaccines, a new method comprising numerous antigenic peptides was created (Kazi et al. 2018). It is possible to create chimeric vaccines using epitope-based vaccines by combining multiple effective epitopes. As a result, a chimera vaccination could boost the epitope vaccine or avoid and protect parasite immune evasion. Furthermore, T. spiral life cycle is complex, resulting in a variety of antigens at distinct phases. T. spiralis infection can be effectively controlled with a multiepitope vaccination.

19. FACTORS OF VACCINE EFFECTIVENESS

Many reasons influence vaccine effectiveness, including antigen composition, transport routes, adjuvants, animal species, coinfection, inoculation doses, infective doses, and immunization strategy. *T. spiralis* has a multiple phases life cycle, which produces distinct antigens at each stage. Because the combination of antigens impacts vaccine effectiveness, identifying great antigens is critical for creating *T. spiralis* vaccines.



During a trichinellosis infection, hosts release Th2-type cytokines (Gao et al. 2022). These cytokines are responsible for increasing mast-cell proliferation and activation, which is necessary for removing the parasite from the intestine. It's important to note that different antigen candidates can induce different immune responses and provide varying levels of protection. Future research into the production of T. spiralis inoculations should concentrate on antigens that can provoke a Th2-type immune reaction. Choosing an appropriate adjuvant is critical in vaccine development. Adjuvants boost immune reactions elicited by parasite antigens and defend them from being, or removed, degraded and diluted by the host (Serradell et al. 2019). The use of Freund's adjuvant is being phased out due to its toxicity and particular damage to experimental animals (Serradell et al. 2023). Although few adjuvants outperform Freund's adjuvant in terms of antibody generation, numerous adjuvants can induce high antibody responses while causing less inflammation and tissue death. In recent decades, alternative adjuvants like Montanide ISA series adjuvants and Montanide IMS series adjuvants have been tested in mouse models to combat T. spiralis infection (Zhang et al. 2018). Coinfection may alter the host's immune response, reducing the efficacy of T. spiralis vaccinations. It needs to be seen if the immune reaction elicited by T. spiralis vaccinations may be inhibited or defused by infection with other organisms. In order to improve T. spiralis vaccines, it is important to consider the immune response generated by various infections. Animal models are well known for their use in vaccine development. Most investigations on vaccination protection have employed mouse representations rather than pig representations (Cai et al. 2022). Vaccine effectiveness, however, may differ based on animal type. Previous research from our group discovered that the immune reaction elicited by the same antigen differs across mice and pigs. To ensure the effectiveness of potential antigens in inducing immunity, it is necessary to validate the significant levels of immunity in swine models after testing in mouse models. Scientists have been working hard to develop and try out different methods for creating vaccines. Although mature immunization regimens have been developed in mice as models, they may not be accessible to pigs or people (Ali et al. 2022). Currently, there is no universally accepted approach for conducting studies using swine models. In order to develop effective T. spiralis vaccines in the future, it is crucial to carefully study the factors that affect their effectiveness.

20. CHALLENGES AND FUTURE PERSPECTIVE

Although there have been significant efforts and progress in searching for potential antigens and developing vaccines for T. spiralis, there are currently no effective vaccinations to prevent T. spiralis infection. Additional immunogenic antigens have been extracted and discovered due to the advancement of genomics, proteomics, and transcriptomics to generate efficient trichinellosis vaccines (Abbas et al. 2023). More and more ways are being used to improve vaccine effectiveness. DNA vaccines are becoming more popular due to their numerous benefits, including low cost and long-lasting protection. In terms of toxoplasmosis vaccinations, a DNA multicomponent vaccine reduced parasite cyst load by 80.22% (Zhang et al. 2018). The combination DNA vaccine could be a potential technique for increasing the efficacy of T. spiralis vaccinations. Furthermore, in mouse models, combined vaccination has been employed as a favorable method to boost the efficacy of T. spiralis vaccines. By combining the advantages of DNA and protein vaccines, the DNA plus protein vaccination technique can stimulate a strong immunological response and provide effective immune protection. VLP vaccines are a commonly used method as they have a tendency to induce strong immune reactions (Keshavarz et al.2019). This technique has been utilized for developing vaccines for toxoplasmosis and presents a new approach for producing vaccines for T. spiralis. Genetic engineering technologies have been used to create live-attenuated toxoplasmosis vaccines through the process of gene editing. The method has been used in the creation of toxoplasmosis vaccines and offers a fresh strategy for the production of *T. spiralis* vaccines. Since the discovery of genetic engineering technologies, gene editing has been used to produce live-attenuated toxoplasmosis vaccines (Zhang et al. 2022). Even though Toxoplasma gondii and T. spiralis have different physiological



characteristics, the same method can be used to develop a vaccine for trichinellosis. T. spiral life cycle in the host is complicated, involving a variety of host response modulation, antigens, and immune evasion. Because of these qualities, it is challenging to establish the best possible defense with a single T. spiralis antigen. With the advancement of genetics, new study thoughts are being offered for boosting the immunity rate of T. spiralis vaccines (Tang et al. 2022). Comprehensive studies on immune evasion and immunosuppression brought on by T. spiralis infection will aid in the development of more potent T. spiralis vaccines. The primary cause of human T. spiralis infection is the consumption of pork and porkassociated products. To present, T. spiralis vaccine research has been conducted in mouse models, and more study in pigs is needed. T. spiralis vaccines employing pig models provide financial and technical challenges, yet this is an important aspect in many vaccinology studies. Because the danger of T. spiralis infection in livestock is minimal under normal management conditions, little emphasis has been paid to the development of T. spiralis vaccinations. Regardless, T. spiralis vaccinations are a nonviolent technique that could help evade medication struggle. As a result, it is critical to instruct people on the significance and benefits of vaccination. Producing a vaccine for Trichinosis is still a major focus of research. To induce protective immunity against trichinellosis infection, scientists were testing numerous vaccine applicants, including recombinant proteins and DNA vaccines. An effective vaccination could help manage Trichinosis by preventing infection in animals and humans.

21. CONCLUSION

Trichinellosis is a serious parasite infection caused by *T. spiralis* that is predominantly transmitted to humans through the ingestion of contaminated meat. While the worldwide incidence of trichinellosis is low in comparison to other foodborne parasites, it remains a public health problem in some areas and can cause severe clinical symptoms if left untreated. Efforts to produce effective vaccinations against *T. spiralis* have shown promise, with many techniques being investigated, including synthetic peptides, DNA vaccines, and recombinant protein vaccines. However, difficulties in recognizing infected meat, as well as the intricacy of the parasite's life cycle, continue to obstruct efficient control. To effectively combat trichinellosis, researchers, public health administrators, and the food sector must work together. Improving surveillance, establishing stringent meat inspection processes, and boosting public knowledge about safe meat consumption habits are critical measures in avoiding and controlling trichinellosis epidemics. Research into new vaccine candidates and technology, as well as advances in diagnostic tools, will be critical in the continued fight against this viral infection. Finally, lowering the worldwide effect of trichinellosis on animal and human health will require a holistic approach that incorporates preventive measures, effective management techniques, and innovative immunization options.

REFERENCES

- Wu J et al., 2022. Extracellular vesicles derived from Trichinella Spiralis larvae promote the polarization of macrophages to M2b type and inhibit the activation of fibroblasts. Frontiers in Immunology 13: 974332.
- Sgroi G et al., 2023. Trichinella britovi in wild boar meat from Italy, 2015–2021: A citizen science approach to surveillance. One Health 16:100480.
- Hady RSEDA et al., 2023. Brief Overview about Trichinellosis; Epidemiology, Pathogenesis, and Clinical Manifestations 12(1):2705-2712.
- Murakami M et al., 2023. Trichinella T9 in wild bears in Japan: Prevalence, species/genotype identification, and public health implications. International Journal for Parasitology: Parasites and Wildlife 21:264-268.
- Pavel R et al., 2023. Trichinellosis in Hospitalized Children and Adults from Western Romania: A 11-Year Retrospective Study. Life 13(4):969.
- Rostami A et al., 2017. Prevalence of Trichinella spp. infections in hunted wild boars in northern Iran. Iranian Journal of Public Health 46(12):1712.



Veronesi F et al., 2023. Wild mesocarnivores as reservoirs of endoparasites causing important zoonoses and emerging bridging infections across Europe. Pathogens 12(2):178.

Pozio E, 2019. Trichinella and trichinellosis in Europe. Veterinarski glasnik 73(2):65-84.

- Pozio E, 2022. The impact of globalization and climate change on Trichinella spp. epidemiology. Food and Waterborne Parasitology 27:e00154.
- Shinn AP et al., 2023. A global review of problematic and pathogenic parasites of farmed tilapia. Reviews in Aquaculture 15: 92-153.
- Bonis V et al., 2021. The intestinal epithelium–fluid fate and rigid structure from crypt bottom to villus tip. Frontiers in Cell and Developmental Biology 9: 661931.
- Nthiga JN, 2022. An investigation on and isolation of parasites that infect humans associated with Catha edulis leaves sold in selected parts of Nairobi County, Kenya (Doctoral dissertation, JKUAT-COHES) 5847.
- Muthumalage T et al., 2019. E-cigarette flavored pods induce inflammation, epithelial barrier dysfunction, and DNA damage in lung epithelial cells and monocytes. Scientific reports 9(1):19035.
- Bandyopadhyay A et al., 2022. Non-neoplastic Lesions of the Appendix. In Surgical Pathology of the Gastrointestinal System: Gastrointestinal Tract 1:481-519.
- Robinson A, 2019. Stem cell therapies for the treatment of enteric neuropathy associated with inflammatory bowel disease (Doctoral dissertation, Victoria University) 39508.
- Saha UB and Saroj SD, 2022. Lactic acid bacteria: Prominent player in the fight against human pathogens. Expert Review of Anti-infective Therapy 20(11): 1435-1453.
- Huang L et al., 2015. Eosinophils and IL-4 support nematode growth coincident with an innate response to tissue injury. PLoS pathogens 11(12):e1005347.
- Bruschi F and Gómez-Morales MA, 2014. The translational immunology of trichinellosis: from rodents to humans. Immune Response to Parasitic Infections—Immunity to Helminths and Novel Therapeutic Approaches 2:125-161.
- Bruschi F and Dupouy-Camet J, 2022. Trichinellosis. In Helminth infections and their impact on global public health. Cham: Springer International Publishing 351-396.
- Tanabe MB et al., 2021. Neurocysticercosis and other CNS helminthic infections. Neurological Complications of Infectious Diseases 225-254.
- Garcia HH et al., 2019. Parasitic infections of the nervous system. In Seminars in neurology (Vol. 39, No. 03, pp. 358-368). Thieme Medical Publishers.
- Leal-Silva T et al., 2021. Detrimental role of IL-33/ST2 pathway sustaining a chronic eosinophil-dependent Th2 inflammatory response, tissue damage and parasite burden during Toxocara canis infection in mice. PLoS Neglected Tropical Diseases 15(7): e0009639.
- Wen, J et al., 2022. Research progress and treatment status of liver cirrhosis with hypoproteinemia. Evidence-based Complementary and Alternative Medicine 2022.
- Bruschi and Murrell K D, 2020. Trichinellosis. In Hunter's Tropical Medicine and Emerging Infectious Diseases (pp. 882-884). Elsevier.
- Bogoch I I et al., 2021. Systemic Parasitic Infections and the Eye. In Albert and Jakobiec's Principles and Practice of Ophthalmology (pp. 1-40). Cham: Springer International Publishing.
- Dupouy-Camet J et al., 2021. Clinical picture and diagnosis of human trichinellosis. In Trichinella and Trichinellosis (pp. 333-352). Academic Press.
- Chalmers R M et al., 2020. Parasite detection in food: Current status and future needs for validation. Trends in Food Science & Technology 99: 337-350.
- Gabriël S et al., 2022. Foodborne parasites and their complex life cycles challenging food safety in different food chains. Foods 12(1): 142.
- Álvarez-Guerrero C and Alba-Hurtado F, 2011. Effect of some physical factors on the viability of third-stage Gnathostoma binucleatum larvae. Journal of Food Protection 74(5): 844-848.
- Yang S M et al., 2022. Microfluidic point-of-care (POC) devices in early diagnosis: A review of opportunities and challenges. Sensors 22(4): 1620.
- Bergwerff A A and Debast S B, 2021. Modernization of control of pathogenic micro-organisms in the food-chain requires a durable role for immunoaffinity-based detection methodology—a review. Foods 10(4):832.



Thanh N T et al., 2014. An inventory of available laboratory diagnostic tests for selected pathogens along the pig value chain in Vietnam.

Haiminen, N et al., 2019. Food authentication from shotgun sequencing reads with an application on high protein powders. NPJ science of food 3(1):24.

Bintsis T, 2017. Foodborne pathogens. AIMS microbiology 3(3):529.

Thrastardottir R et al., 2021. Yellow mealworm and black soldier fly larvae for feed and food production in europe, with emphasis on iceland. Foods 10(11): 2744.

Gamble H R, 2022. Trichinella spp. control in modern pork production systems. Food and Waterborne Parasitology e00172.

Zhang N Li W and Fu B, 2018. Vaccines against Trichinella spiralis: progress, challenges and future prospects. Transboundary and emerging diseases 65(6): 1447-1458.

Tang B et al., 2022. Vaccines as a strategy to control trichinellosis. Frontiers in microbiology 13: 857786.

- Khalid K and Poh C L, 2023. The development of DNA vaccines against SARS-CoV-2. Advances in Medical Sciences 68(2): 213-226.
- Xu D et al., 2020. The immune protection induced by a serine protease from the Trichinella spiralis adult administered as DNA and protein vaccine. Acta tropica 211: 105622.
- Sofronic-Milosavljevic L et al., 2015. Secretory products of Trichinella spiralis muscle larvae and immunomodulation: implication for autoimmune diseases, allergies, and malignancies. Journal of immunology research.
- Song Y Y et al., 2018. Characterization of a serine protease inhibitor from Trichinella spiralis and its participation in larval invasion of host's intestinal epithelial cells. Parasites & vectors 11(1):1-12.
- Grzelak S et al., 2020. Immunoproteomic analysis of Trichinella spiralis and Trichinella britovi excretory-secretory muscle larvae proteins recognized by sera from humans infected with Trichinella. PLoS One 15(11), e0241918.
- Stachyra A and Wesołowska A, 2023. Immunomodulatory in vitro effects of Trichinella cystatin-like protein on mouse splenocytes. Experimental Parasitology 108585.
- Stachyra A et al., 2019. The immunological properties of recombinant multi-cystatin-like domain protein from Trichinella britovi produced in yeast. Frontiers in Immunology 10: 2420.
- Kumari P et al., 2020. AIM2 in health and disease: Inflammasome and beyond. Immunological reviews 297(1): 83-95.
- Cui J et al., 2019. Vaccination of mice with a recombinant novel cathepsin B inhibits Trichinella spiralis development, reduces the fecundity and worm burden. Parasites & vectors 12: 1-12.
- Xu, D et al., 2021. Vaccination with a DNase II recombinant protein against Trichinella spiralis infection in pigs. Veterinary parasitology 297: 109069.
- Hafez E N et al., 2020. The potential protective role of gamma-irradiated vaccine versus Punica granatum treatment against murine trichinellosis. Journal of Radiation Research and Applied Sciences 13(1): 560-567.

Santi A M et al., 2018. Growth arrested live-attenuated Leishmania infantum KHARON1 null mutants display cytokinesis defect and protective immunity in mice. Scientific reports 8(1):11627.

- Wait L F, 2022. When Parasites Interact: Implications for Immunisation Efficacy in Individuals and Populations (Doctoral dissertation, Princeton University).
- Parvizpour S et al., 2020. Epitope-based vaccine design: a comprehensive overview of bioinformatics approaches. Drug Discovery Today, 25(6): 1034-1042.
- Sanches R C et al., 2021. Immunoinformatics design of multi-epitope peptide-based vaccine against Schistosoma mansoni using transmembrane proteins as a target. Frontiers in immunology 12, 621706.
- Soleymani S et al., 2022. An overview of progress from empirical to rational design in modern vaccine development, with an emphasis on computational tools and immunoinformatics approaches. Computers in biology and medicine 140: 105057.
- Qin, F et al., 2021. A guide to nucleic acid vaccines in the prevention and treatment of infectious diseases and cancers: from basic principles to current applications. Frontiers in cell and developmental biology 9, 633776.

Wu Z et al., 2021. Proteomics of Trichinella. In Trichinella and Trichinellosis 103-183). Academic Press.

Xu D et al., 2020. The immune protection induced by a serine protease from the Trichinella spiralis adult administered as DNA and protein vaccine. Acta tropica 211: 105622.



- Gu Y et al., 2020. A multiple antigen peptide vaccine containing CD4+ T cell epitopes enhances humoral immunity against Trichinella spiralis infection in mice. Journal of immunology research 2020.
- Kazi A et al., 2018. Current progress of immunoinformatics approach harnessed for cellular-and antibody-dependent vaccine design. Pathogens and global health 112(3): 123-131.
- Gao X et al., 2022. Extracellular vesicles from Trichinella spiralis: Proteomic analysis and protective immunity. PLOS Neglected Tropical Diseases 16(6): e0010528.
- Serradell M C et al., 2019. Efficient oral vaccination by bioengineering virus-like particles with protozoan surface proteins. Nature communications 10(1): 361.
- Cai L et al., 2022. Factors limiting the translatability of rodent model–based intranasal vaccine research to humans. AAPS PharmSciTech 23(6): 191.
- Ali A et al., 2022. Recent advancement, immune responses, and mechanism of action of various vaccines against intracellular bacterial infections. Life Sciences 121332.
- Abbas M N et al., 2023. Recent Advances in Tick Antigen Discovery and Anti-Tick Vaccine Development. International Journal of Molecular Sciences 24(5): 4969.
- Keshavarz M et al., 2019. Influenza vaccine: Where are we and where do we go?. Reviews in medical virology 29(1): e2014.
- Zhang Y et al., 2022. Toxoplasmosis vaccines: what we have and where to go? npj Vaccines 7(1): 131.
- Foreyt, W. J., & Abbott, R. C. (2013). Trichinosis (No. 1388). US Geological Survey.
- Gottstein, B., Pozio, E., & Nöckler, K. (2009). Epidemiology, diagnosis, treatment, and control of trichinellosis. *Clinical microbiology reviews*, 22(1), 127-145.

Mitreva, M., & Jasmer, D. P. (2006). Biology and genome of Trichinella spiralis.