

Chapter 16

Plant as Anthelmintic Allies in the Fight against Fasciolosis: A Review

Muhammad Zulqarnain¹, Sidra Abbas¹, Muhammad Arfan Zaman², Erick Maosa Bosire⁵, Ghazanfer Ali³, Naila Noreen Kanwal⁴, Muhammad Zahid Sarfaraz¹, Iqra Khalil¹ and Sajida Bibi¹

¹Department of Zoology, University of Jhang, Jhang, Pakistan

²Department of Pathobiology, College of Veterinary and Animal Sciences, Jhang, sub-campus UVAS Lahore, Pakistan

³Institute of Biological Sciences, Faculty of Science, Universiti Malaya, Malaysia

⁴Department of Zoology, Institute of Molecular Biology and Biotechnology, The University of Lahore, Lahore, Pakistan

⁵Department of Population Medicine and Diagnostic Sciences, College of Veterinary Medicine, Cornell University, Ithaca, New York, USA

*Corresponding author: arfan.zaman@uvas.edu.pk

ABSTRACT

Plants are the primary food source for ruminating mammals; therefore, using different medicinal plants (extracts, oils, and bioactive compounds) could be effective fasciolicides to prevent economic losses due to fasciolosis. The problem of anthelmintic resistance in flukes due to the excessive and inappropriate use of fasciolicides poses an alarming threat and urges the development of new and alternative fasciolicides. Now, the focus is diverging towards plant-based fasciolicides because of their safe and environment-friendly nature. Different plants' essential oils and active compounds are effective in controlling egg hatching and causing the mortality of adult flukes. The *Artemisia* and *Etingera* extracts exhibited significant efficacy against eggs and adults of *Fasciola spp.* The plants (*Zingibar officinale* and *Potentilla fulgens*) also showed considerable efficacy against miracidia, sporocyst, rediae, and cercarial stages. The use of plant oils in controlling parasitism has been practiced all over the world. Oils from (*Pelargonium graveolens*, *Citrus aurantium*, *Helianthus annuus*, and *Cuminum cyminum*) plants actively inhibit egg development. Moreover, plants' active compounds, diterpenoids, thymoquinone, curcumin, flavonoids, acids, artemisinin, and saponins also show promising effects on egg development and adult fluke productivity.

KEYWORDS

Fasciola, Fasciolosis, Anthelmintics, Phytotherapy, Medicinal Plants

Received: 04-Jun-2024

Revised: 01-Jul-2024

Accepted: 05-Aug-2024



A Publication of
Unique Scientific
Publishers

Cite this Article as: Zulqarnain M, Abbas S, Zaman MA, Bosire EM, Ali G, Kanwal NN, Sarfaraz MZ, Khalil I and Bibi S, 2024. Plant as anthelmintic allies in the fight against fasciolosis: a review. In: Abbas RZ, Khan AMA, Qamar W, Arshad J and Mehnaz S (eds), Complementary and Alternative Medicine: Botanicals/Homeopathy/Herbal Medicine. Unique Scientific Publishers, Faisalabad, Pakistan, pp: 143-149. <https://doi.org/10.47278/book.CAM/2024.148>

INTRODUCTION

Fasciolosis (Fascioliasis) is a zoonotic disease that affects animals and humans worldwide. A trematode parasite, *Fasciola* (*F. hepatica* and *F. gigantica*) also called liver fluke, causes the disease and can be transmitted to animals and humans through its infective stage (metacercariae). The life cycle of this parasite consists of two hosts. The eggs are shed in the faeces of ruminants as sheep, cattle, and goat, and are hatched into the miracidia that infect the first host snail (Lymnid). There, it progresses asexually into its two stages, sporocysts and rediae, after which snail starts shedding cercariae in the water. These cercariae swim randomly, and after a few hours, they lose their tail, get enclosed in a wall and become metacercariae (infective stage) and attached to the watercress or leaves. When the final host mammals (sheep, cattle, and goat), ingest watercress, undercooked vegetables or leaves, the infective metacercariae become converted into immature flukes which excyst in the duodenum, penetrate the intestinal wall and migrate through liver parenchyma to biliary ducts (FAO, 2020; Mia et al., 2021).

Prevalence of Fasciolosis

Fasciola hepatica is prevalent worldwide, while *Fasciola gigantica* is distributed in the tropics (Urquhart et al., 1996). The overall infection rate is increasing due to climate change, drug resistance, and its ability to intrude new areas. It infects more than 2.4 million people worldwide, with 180 million at risk of infection in 66 countries. In livestock, it is hard to quantify, but according to an estimate, it infects over 600 million domestic ruminants annually, which causes heavy economic losses (Collado et al., 2019). The global prevalence of fasciolosis in livestock ranges from 0.72 to 94% (Khan et

al., 2013). Humans are accidental hosts which may be due to the ingestion of raw or undercooked vegetables. The infection rate in humans is low with an estimated 4.5% of people worldwide are thought to have fasciolosis (Infantes et al., 2023).

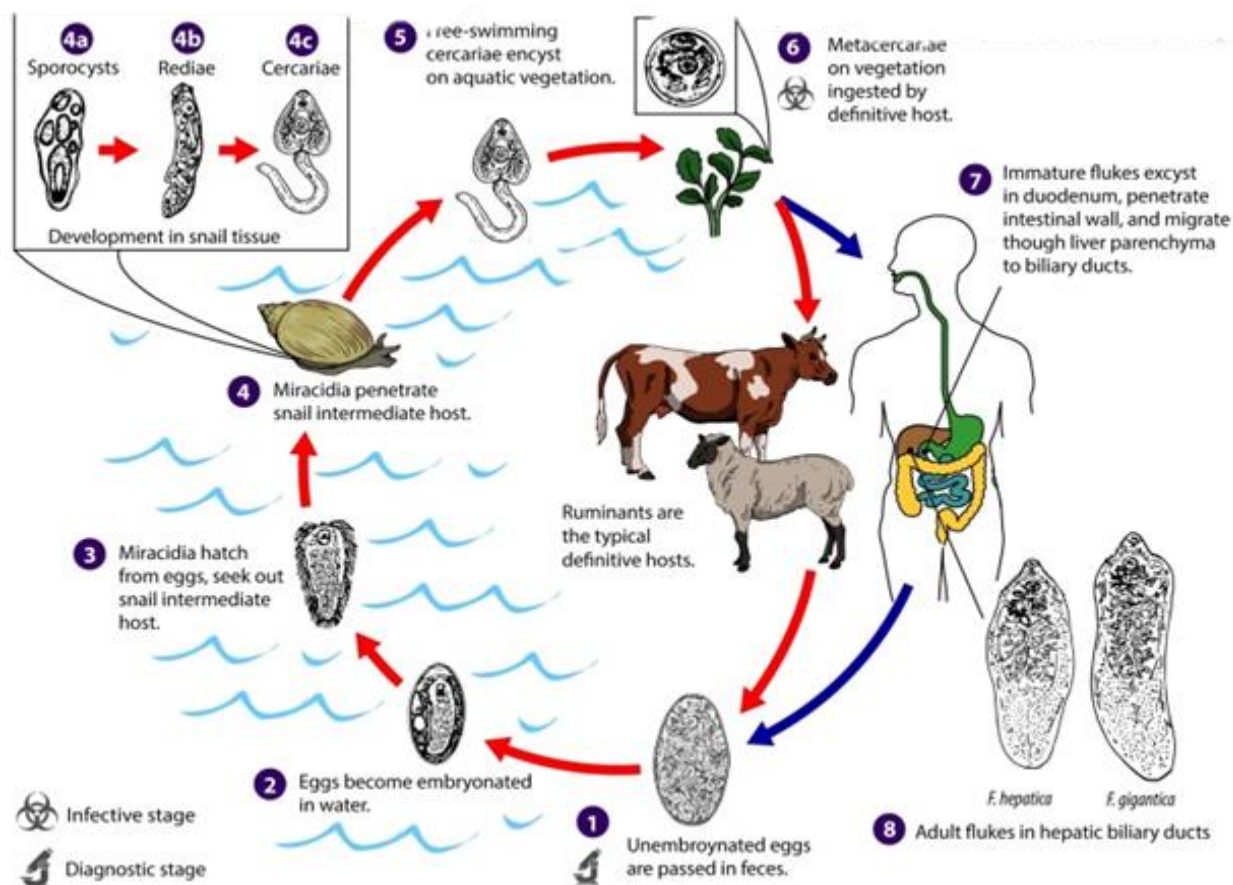


Fig: Life cycle of *Fasciola* spp.

The prevalence of fasciolosis in Pakistan from 2000–2020 was reported in a study. It was 42.70% in Sindh, 25.20% in Baluchistan, 17.70% in Punjab, 10.70% in KPK, and 1.50% in Islamabad. The prevalence was highest among sheep in Punjab at 65.7%, in Baluchistan cows at 28.5%, and in KPK buffalo at 15.9% (Rizwan et al., 2022).

Economic Impact

Helminthes infection costs the agriculture sector an annual loss of US\$20 billion towards animal productivity, out of which, the liver fluke infection was predicted to cause annual losses of about US\$3.20 billion worldwide (Mehmood et al., 2017; FAO, 2020). The following Table 1 summarizes the economic losses throughout the world due to *Fasciola* infection.

Table 1: Economic Losses in Different Regions of the World

Regions	Countries	Economic Loss through	Economic Loss	Reference
Asia	Iraq	Liver condemnation	US\$ 8801.69	Kadir et al. (2012)
	Saudi Arabia		US\$ 0.2 M	Degheidy and Al-Malki (2012)
	Sudan		US\$ 1.94 M	Abebe et al. (2010)
Africa	Uganda	Liver condemnation	US\$ 92 M	Joan et al. (2015)
America	Brazil	Weight loss	35 US\$ PH	Dutra et al. (2010)
	Mexico	Productivity loss through reduced milk and meat yield	US\$ 4.2 M	Rodriguez et al. (2017)
Australia	Australia	Productivity loss through meat and milk, liver condemnation	60–90 M A\$	Toet et al. (2014)
Europe	Switzerland	Reduced milk yield, fertility and meat	52\$ M	Schweizer et al. (2005)

Control of Fasciolosis

Chemotherapy

Chemotherapy has a primary role in controlling parasitic diseases as it is affordable and effective (Khan et al., 2017). The control of fascioliasis is mainly achieved using synthetic anthelmintics such as albendazole, triclabendazole, and

nitroxylnil (Nixon et al., 2020; Kouadio et al., 2021). Among the benzimidazole derivatives, triclabendazole (TCBZ) is the most important and widely used drug of choice that can effectively control fasciolosis (Castro Hermida et al., 2021). It is the only drug that has an efficacy of >98% against adults and especially immature flukes as compared to other flukicides that target only adult flukes (Kahl et al., 2023).

Resistance against Anthelmintics

The liver fluke infection is increasing due to climate change, changes in the land use, increased movement of livestock, and its ability to intrude into new areas, which is compounded by the rising issue of anthelmintic resistance in *Fasciola* spp. (Beesley et al., 2023). The problem with the triclabendazole (TCBZ) resistance may be due to the farmers using less effective substitutes or even administering the dose inadvertently (Fairweather et al., 2012). In the recent years, many studies have shown that TCBZ is ineffective against *F. hepatica* in ruminants all over the world. The threat of drug resistance requires serious attention, as many reports have demonstrated drug resistance worldwide, including Scotland (Sargison and Scott, 2011), Spain (Martinez Valladares et al., 2014), and New Zealand (Hassell and Chapman, 2012), Wales (Gordon et al., 2012), Peru (Ortiz et al., 2013), Australia (Brockwell et al., 2014), Ireland (Hanna et al., 2015), and Argentina (Larroza et al., 2023).

Besides the issue of anthelmintic resistance, chemical residues found in derived products like milk and meat and their environmental effects are additional crucial factors to be aware of (Da Silva et al., 2020). Thus, for sustainable livestock production, the management of fasciolosis towards TCBZ resistance and the development of new alternatives as flukicides are necessary.

Plants as Anthelmintic Allies

Currently, the livestock sector is under threat due to the problem of increasing resistance to anthelmintics, which is because of factors including inappropriate dosage and repeated exposure of anthelmintics for deworming. Hence, search for alternative anthelmintics is necessary. For this, plant-based anthelmintics could be a preference for coping with this problem. Plant-based anthelmintic are becoming a growing trend because they are safer than the synthetic ones. Moreover, plants provide cost-effective substitutes that are more effective than synthetic anthelmintics (Zirintunda et al., 2022).

Around 80% of people use traditional medicine all over the world, and out of them, about 85% depends on medicinal plants (Nascimento et al., 2000; Oyebode et al., 2016; Romero-Benavides et al., 2017; Sanchez et al., 2020). Plants have been used as medicinal agents from millennia and become the integral part of animal's life. Moreover, plants are also immune boosters that protects against many diseases and also provide many vital nutrients to animal's body. Plants have some of the nature's most effective medicinal compounds. Their extracts and bioactive substances have the potential to become new medicinal agents because of their diversity and environment friendly nature (Abbas et al., 2020; Nurlaelasari et al., 2023). Many medicinal plants can be used as antibacterial agents (Chassagne et al., 2021), antiparasitic (Benlarbi et al., 2023), and antifungal agents (Nigussie et al., 2021). Hence identifying and validating different compounds and their extracts as effective fasciolicide is also necessary. For this, during the recent decades, many plants and their extracts have been evaluated for their anthelmintic activity worldwide (Pessoa et al., 2002; Kozan et al., 2006; Eguale et al., 2011; Ahmed et al., 2013; Payne et al., 2013; Acharya et al., 2014; Esteban Ballesteros et al., 2019).

Plant Extracts

Use of plants as a whole requires a lot of plant mass moreover these are not as much effective. Instead, plant extracts and purified fractions must be used which can provide effective alternative to fasciolicides. The efficacy of plant extract depends on the solvent (water, methanol, ethanol, etc.) used for the extraction and also the mechanism of extraction (hot or cold maceration, proclation, soxhlet extraction, etc.). The solvents provide a medium for the extraction of active phytochemicals like phenols, alkalines, tannins, saponins, etc., and these should be extracted in a preferred medium. Plant extracts provide a combination of different phytochemicals that collectively act as fasciolicide. There are many studies that use different plant extracts to validate their efficacy against *Fasciola* spp. Nurlaelasari et al. (2023) conducted an experiment to evaluate the antifasciolic activity of Mugwort (*Artemisia vulgaris*) extract on the both adult and eggs stages of *Fasciola gigantica*. They assessed the ovicidal efficacy by incubating the eggs with the *A. vulgaris* extract at 5%, 2.5%, and 1.25% for 5, 9, 11, 14, and 16 days. The adult mortality assays were performed after incubating with the extract for 5, 10, 20, 40, 80, 160, 320, and 640 min. All the extract concentrations show significant anti-fasciolic activities. Among these, a 5% concentration of extract showed high level of ovicidal activity, whereas a concentration of 20% extract inhibits 66.67% of eggs hatching after 40 min of exposure. The extract also had high flukicidal efficacy, causing disintegration of different physiological structures (vitelline follicles, intestine, acetabulum, tegument, and spine) of adult fluke. This study has indicated that *A. vulgaris* extract exhibits potential antifasciolic properties. In another study conducted by Wulandari et al. (2023), Torch ginger (*Etilingera elatior*) ethanolic extract was evaluated on eggs and adults of *Fasciola gigantica*. The egg-hatching assay was performed at different concentrations. The ova development was reduced by 56.67%, 56.67%, and 36.67%, at 5%, 2.5%, and 1.25% concentrations, respectively, on day 11 post-incubation, and after 14 days, these developed eggs were decreased by 70%, 50%, and 13.33%, respectively. The flukicidal efficacy was recorded at 10% and 20% of the extract concentration, and the death of flukes was noted after 640 min and 80 min, respectively. The skin was damaged, the inner membrane of spina has erosions and syncytium was detached from the tegument. The study showed

that *E. elatior* extract has antifasciolic activity against different stages of flukes. The ginger (*Zingiber officinale*) hydroalcoholic extract was evaluated on the miracidial stage of *Fasciola hepatica* by Ghafari et al. (2021). The hatched miracidia was incubated at 2, 5, and 10 µg/mL concentrations. The extract concentrations of 10, 5, and 2 µg/ml cause the fatality of miracidia within 105, 275, and 520 sec. and reduce the speed of miracidia movement to 0.08, 0.77, and 0.82 mm/s, respectively. The comparative efficacy with triclabendazole shows that the extract has antimiracidial efficacy. Kumar et al. (2020) performed the *in-vitro* larvicidal assay on sporocyst, redia, and cercaria stages of *Fasciola*. The Lined Cinquefoil (*Potentilla fulgens*) organic extracts and column-purified fractions larval motility was time and concentration-dependent. The highest toxicity against sporocyst, redia, and cercaria after 2h (LC50) of exposure was column purified fraction 62.4, 59.5, and 45.1 mg/L, respectively. In contrast, the lowest toxicity at this time was of ethanol extract 66.2, 64.7, and 51.3 mg/L, respectively. The same trend was found after 8h (LC50) of exposure, where column purified fraction has the highest toxicity while ethanol extract has the lowest toxicity.

Plant Oils

A number of plant derived natural products including the natural oils have been shown to possess anti parasitic activity, as they have been used against different parasites *in-vitro* and *in-vivo*. De Mello et al. (2023) evaluated the antifasciolic activity of essential oils (EOs) from two plants, Geranium (*Pelargonium graveolens*) and Sour orange (*Citrus aurantium*), on *Fasciola hepatica*. The essential oils from both plants show 100% efficacy to inactive egg hatching. The Geranium extract showed a mean mortality time of 15 h at both concentrations tested 0.0675 and 0.03375 mg/mL, while Sour orange extract promoted death within 18 h of incubation at concentration of 0.06375 mg/mL. The accumulation of liquid in the tegument was observed. It indicates that essential oils have potential ovicidal and adulticidal activities. Another research conducted by Da Silva et al. (2020) evaluated the Sunflower (*Helianthus annuus*) fixed oil and Cumin (*Cuminum cyminum*) essential oil against *Fasciola hepatica*. The *in-vitro* assay was assessed at different concentrations both fixed and essential oils and also the combination of both oils. The essential oil at concentration of 0.03 mg/mL showed 99% efficacy and the combination of both oils at 0.035+0.03 mg/mL showed 94% effectiveness, while the fixed oil was active insufficiently as an ovicidal. Overall, the results of the experiment showed that the essential oil of Cumin could be used as a new alternative for fascioliasis control.

Plant Active Compounds

Plant compounds and their active ingredients have great potential in the control of various parasites, thus interest is increasing for the search and use of new alternatives. The botanical-derived compounds harboring potential parasitocidal properties are tested as therapeutic agents worldwide. The most common phytochemical constituents of plants are diterpenoids, thymoquinone, curcumin, carbohydrates, terpenoids, fats, enzymes, amino acids, flavonoids, chicoric acid, phenols, polyphenols, alkaloids, artemisinin, saponins, anthocyanins, tannins, isoflavones and carotenoids etc. Interest in the studies with bioactive compounds has been growing as they show direct or indirect negative effects against endoparasites. Several natural diterpenoid molecules have been studied against parasitic trematodes, and their anthelmintic properties have been evaluated. In a study, Chakroborty et al. (2022) evaluated the use of nineteen chemically modified natural active compounds, abietic acid diterpenoid analogues (MC001 to MC088) were first evaluated for their anthelmintic activities against newly excysted juveniles (NEJs) of *Fasciola hepatica*. The six analogues that were proven effective against NEJs (MC008, MC009, MC010, MC052, MC058, and MC061) were secondly evaluated for their anthelmintic activities against adult wild strain flukes. From these analogues MC010 was highly effective against 8-week immature- and 12-week mature Italian strain flukes. The damage to the dorsal side of the fluke was observed. They deduced that the use of abietic acids can be a potential candidate for the development of new anthelmintics. Thymoquinone and curcumin are the active ingredients of *Nigella sativa* and *Curcuma longa*, respectively. Ullah et al. (2017) used these active ingredients as flukicidal agents against *Fasciola gigantica*. The worm motility and egg shedding were both time and concentration-dependent. The adult flukes were exposed to different concentrations (20, 40, 60 µM) of thymoquinone and curcumin. The reduction in motility was observed at 60 µM, but the worms remained alive for 3h post-exposure. The tegumental disruptions and spine erosion were observed in the posterior region and around the acetabulum. Thus, thymoquinone and curcumin have the potential to have a flukicidal effect.

Conclusion

Fasciolosis control using plant-based medicines is proving to be more effective than synthetic anthelmintic, i.e. albendazole. Moreover, plant-based medicines are a growing trend towards a sustainable and nature-friendly environment. In phytotherapy, the time and concentration-dependent factors are the most important; hence, determining toxicity is necessary. Further research using different plant extracts, oils, and their active compounds in this field should be evaluated. However, to confirm which concentrations effectively control eggs, miracidia, larvae, and adult stages is required. Moreover, the mechanism of action should also be understood through different methods like the molecular docking method to eradicate fasciolosis from the livestock industry completely.

REFERENCES

Abbas, R. Z., Zaman, M. A., Sindhu, D., Sharif, M., Rafique, A., Saeed, Z. and Ahmad, M. (2020). Anthelmintic Effects and

- Toxicity Analysis of Herbal Dewormer against the Infection of *Haemonchus Contortus* and *Fasciola Hepatica* in Goat. *Pakistan Veterinary Journal*, 40(4), 455-460.
- Abebe, R., Abunna, F., Berhane, M., Mekuria, S., Megersa, B., and Regassa, A. (2010). Fasciolosis: Prevalence, financial losses due to liver condemnation and evaluation of a simple sedimentation diagnostic technique in cattle slaughtered at Hawassa Municipal abattoir, southern Ethiopia. *Ethiopian Veterinary Journal*, 14(1), 39-52.
- Acharya, J., Hildreth, M. B., and Reese, R. N. (2014). In Vitro Screening Of Forty Medicinal Plant Extracts From The United States Northern Great Plains For Anthelmintic Activity Against *Haemonchus Contortus*. *Veterinary Parasitology*, 201(1-2), 75-81.
- Ahmed, M., Laing, M. D., and Nshahai, I. V. (2013). In Vitro Anthelmintic Activity Of Crude Extracts Of Selected Medicinal Plants Against *Haemonchus Contortus* From Sheep. *Journal of Helminthology*, 87(2), 174-179.
- Beesley, N. J., Cwiklinski, K., Allen, K. A., Hoyle, R. C., Spithill, T. W., La Course, E. J., Williams, D., Paterson, S., and Hodgkinson, J. E. (2023). A major locus confers triclabendazole resistance in *Fasciola hepatica* and shows dominant inheritance. *PLOS Pathogens*, 19(1), e1011081.
- Benlarbi, F., Mimoune, N., Chaachouay, N., Souttou, K., Saidi, R., Mokhtar, M. R., and Benaissa, M. H. (2023). Ethnobotanical survey of the traditional antiparasitic use of medicinal plants in humans and animals in Laghouat (Southern Algeria). *Veterinary World*, 16(2), 357.
- Brockwell, Y., Elliott, T., Anderson, G. R., Stanton, R., Spithill, T. W., and Sangster, N. (2014). Confirmation of *Fasciola hepatica* resistant to triclabendazole in naturally infected Australian beef and dairy cattle. *International Journal for Parasitology, Drugs and Drug Resistance*, 4(1), 48-54.
- Calderon Montano, J. M., Martinez-Sanchez, S. M., Jimenez-Gonzalez, V., Burgos Moron, E., Guillen Mancina, E., Jimenez-Alonso, J. J., and Lopez Lazaro, M. (2021). Screening for selective anticancer activity of 65 extracts of plants collected in western Andalusia, Spain. *Plants*, 10(10), 2193.
- Castro Hermida, J. A., Gonzalez Warleta, M., Martinez Sernandez, V., Ubeira, F. M., and Mezo, M. (2021). Current Challenges for Fasciolicide Treatment in Ruminant Livestock. *Trends in Parasitology*, 37(5), 430-444.
- Chakroborty, A., Pritchard, D., Bouillon, M. E., Cervi, A., Cookson, A., Wild, C., Fenn, C., Payne, J. E., Holdsworth, P., Capner, C., O'Neill, J., Padalino, G., Forde-Thomas, J., Gupta, S., Smith, B. G., Fisher, M., Lahmann, M., Baird, M. S., and Hoffmann, K. F. (2022). Flukicidal effects of abietane diterpenoid derived analogues against the food borne pathogen *Fasciola hepatica*. *Veterinary Parasitology*, 309, 109766. <https://doi.org/10.1016/j.vetpar.2022.109766>
- Chassagne, F., Samarakoon, T., Porras, G., Lyles, J. T., Dettweiler, M., Marquez, L., Salam, A. M., Shabih, S., Farrokhi, D. R., and Quave, C. L. (2021). A Systematic Review of Plants with Antibacterial Activities: A Taxonomic and phylogenetic perspective. *Frontiers in Pharmacology*, 11, 2069. <https://doi.org/10.3389/fphar.2020.586548>
- Chaves, N., Santiago, A., and Alias, J. C. (2020). Quantification of the antioxidant activity of plant extracts: Analysis of sensitivity and hierarchization based on the method used. *Antioxidants*, 9(1), 76.
- Collado, D. P., Valdes, J. B., Molento, M. B., Gil, A. V., Vazquez, R. O., and Napoles, C. F. (2019). Economic Losses and Prevalence of *Fasciola Hepatica* in Cattle Slaughtered In Two Cuban Provinces. *Revista Mvz Cordoba*, 25(1), 1610.
- Da Silva, M. a. M. P., Zehetmeyr, F. K., Pereira, K. M., Pacheco, B. S., Freitag, R. A., Pinto, N. B., Machado, R. H., Villarreal, J. Z., De Oliveira Hübner, S., Berne, M. E. A., and Da Silva Nascente, P. (2020). Ovicidal in vitro activity of the fixed oil of *Helianthus annuus* L. and the essential oil of *Cuminum cyminum* L. against *Fasciola hepatica* L. *Experimental Parasitology*, 218, 107984. <https://doi.org/10.1016/j.exppara.2020.107984>
- De Mello, A. B., Baccega, B., Martins, F. O., Da Rosa Farias, N. A., De Giacometti, M., Da Fonseca, R. N., De Oliveira Hübner, S., Soares, M. P., and Oliveira, C. B. (2023). Microscopic alterations in *Fasciola hepatica* treated with the essential oils of *Pelargonium graveolens* and *Citrus aurantium*. *Veterinary Parasitology*, 314, 109863. <https://doi.org/10.1016/j.vetpar.2022.109863>
- Degheidy, N. S., and Al-Malki, J. S. (2012). Epidemiological studies of fasciolosis in human and animals at Taif, Saudi Arabia. *World Applied Sciences Journal*, 19(8), 1099-1104.
- Dutra, L. H., Molento, M. B., Naumann, C. R. C., Biondo, A. W., Fortes, F. S., Savio, D., and Malone, J. B. (2010). Mapping risk of bovine fasciolosis in the south of Brazil using Geographic Information Systems. *Veterinary Parasitology*, 169(1-2), 76-81.
- Egualde, T., Tadesse, D., and Giday, M. (2011). In Vitro Anthelmintic Activity Of Crude Extracts Of Five Medicinal Plants Against Egg-Hatching And Larval Development Of *Haemonchus Contortus*. *Journal of Ethnopharmacology*, 137(1), 108-113.
- Esteban Ballesteros, M., Sanchis, J., Gutierrez Corbo, C., Balana Fouce, R., Rojo Vazquez, F., Gonzalez-Lanza, C., and Martinez-Valladares, M. (2019). In vitro anthelmintic activity and safety of different plant species against the ovine gastrointestinal nematode *Teladorsagia circumcincta*. *Research in Veterinary Science*, 123, 153-158. <https://doi.org/10.1016/j.rvsc.2019.01.004>
- Fairweather, I., Mcshane, D. D., Shaw, L., Ellison, S. E., Ohagan, N. T., York, E. A., and Brennan, G. P. (2012). Development of an Egg Hatch Assay for the Diagnosis of Triclabendazole Resistance in *Fasciola Hepatica*: Proof of Concept. *Veterinary Parasitology*, 183(3-4), 249-259.
- Ghafari, A., Arbabi, M., Mosayebi, M., Hooshyar, H., and Nickfarjam, A. M. (2021). Evaluation of anti-helmintic activity of Zingiber officinale roscoe extract on *Fasciola hepatica* miracidia In vitro. *International Archives of Health Sciences*, 8(1),

45-50.

- Gordon, D., Zadoks, R. N., Skuce, P., and Sargison, N. (2012). Confirmation of triclabendazole resistance in liver fluke in the UK. *The Veterinary Record*, 171(6), 159–160.
- Hanna, R., McMahon, C., Ellison, S., Edgar, H., Kajugu, P., Gordon, A., Irwin, D., Barley, J., Malone, F., Brennan, G., and Fairweather, I. (2015). *Fasciola hepatica*: A comparative survey of adult fluke resistance to triclabendazole, nitroxylnil and closantel on selected upland and lowland sheep farms in Northern Ireland using faecal egg counting, coproantigen ELISA testing and fluke histology. *Veterinary Parasitology*, 207(1–2), 34–43.
- Hassell, C., and Chapman, V. (2012). Case report: suspect lack of triclabendazole efficacy in liver fluke in sheep in the Taranaki. *Proc. Soc. Sheep Beef Cattle Vet. New Zealand Veterinary Association*, 2012(4.14), 1-144.
- Infantes, L. R. R., Yataco, G. a. P., Ortiz-Martínez, Y., Mayer, T., Terashima, A., Franco-Paredes, C., González-Díaz, E., Rodríguez-Morales, A. J., Bonilla-Aldana, D. K., Barahona, L. V., Grimshaw, A., Chastain, D. B., Sillau, S., Marcos, L. A., and Henao-Martínez, A. F. (2023). The global prevalence of human fascioliasis: a systematic review and meta-analysis. *Therapeutic Advances in Infectious Disease*, 10. <https://doi.org/10.1177/20499361231185413>
- Joan, N., Stephen, M. J., Bashir, M., Kiguli, J., Orikiriza, P., Bazira, J., and Stanley, I. J. (2015). Prevalence and economic impact of bovine fasciolosis at Kampala City Abattoir, Central Uganda - *Open Archive Press*. (n.d.). <http://library.2pressrelease.co.in/id/eprint/1486/>
- Kadir, M. A., Ali, N. H., & Ridha, R. G. M. (2012). Prevalence of helminthes, pneumonia and hepatitis in Kirkuk slaughter house, Kirkuk, Iraq. *Iraqi Journal of Veterinary Sciences*, 26(2012), 83-88.
- Kahl, A., Von Samson Himmelstjerna, G., Helm, C., Hodgkinson, J. E., Williams, D., Weiher, W., Terhalle, W., Steuber, S., Ganter, M., and Krücken, J. (2023). Efficacy of flukicides against *Fasciola hepatica* and first report of triclabendazole resistance on German sheep farms. *International Journal for Parasitology, Drugs and Drug Resistance*, 23, 94–105. <https://doi.org/10.1016/j.ijpddr.2023.11.001>
- Khan, M. K., Sajid, M. S., Riaz, H., Ahmad, N. E., He, L., Shahzad, M., and Zhao, J. (2013). The Global Burden Of Fasciolosis In Domestic Animals With An Outlook On The Contribution Of New Approaches For Diagnosis And Control. *Parasitology Research*, 112, 2421-2430. <https://doi.org/10.1007/s00436-013-3464-6>
- Khan, M. N., Sajid, M. S., Rizwan, H. M., Qudoos, A., Abbas, R. Z., Riaz, M., and Khan, M. K. (2017). Comparative Efficacy of Six Anthelmintic Treatments against Natural Infection of *Fasciola Spp.* in Sheep. *Pakistan Veterinary Journal*, 37(1), 65-68.
- Kouadio, J. N., Evack, J. G., Achi, L. Y., Balmer, O., Utzinger, J., N'goran, E. K., and Zinsstag, J. (2021). Efficacy of Triclabendazole and Albendazole against *Fasciola Spp.* Infection in Cattle in Cote Divoire: A Randomised Blinded Trial. *Acta Tropica*, 222, 106039. <https://doi.org/10.1016/j.actatropica.2021.106039>
- Kozan, E., Kupeli, E., and Yesilada, E. (2006). Evaluation of Some Plants Used In Turkish Folk Medicine against Parasitic Infections for Their In Vivo Anthelmintic Activity. *Journal of Ethnopharmacology*, 108(2), 211-216.
- Kumar, P., Sunita, K., Singh, R. N., and Singh, D. K. (2020). Fasciola larvae: Anthelmintic activity of medicinal plant *Potentilla fulgens* against sporocyst, redia and cercaria. *Asian Journal of Advances in Research*, 3(1), 95-101.
- Larroza, M. P., Aguilar, M., Soler, P., Mora, J. C., Roa, M., Cabrera, R., Stanziola, J. P. M., Ceballos, L., and Álvarez, L. I. O. (2023). Triclabendazole resistance in *Fasciola hepatica*: First report in sheep from the Santa Cruz province, Argentinian Patagonia. *Veterinary Parasitology. Regional Studies and Reports*, 45, 100927. <https://doi.org/10.1016/j.vprsr.2023.100927>
- Martinez Valladares, M., Cordero-Perez, C., and Rojo-Vazquez, F. (2014). Efficacy of an anthelmintic combination in sheep infected with *Fasciola hepatica* resistant to albendazole and clorsulon. *Experimental Parasitology*, 136, 59–62. <https://doi.org/10.1016/j.exppara.2013.10.010>
- Mehmood, K., Zhang, H., Sabir, A. J., Abbas, R. Z., Ijaz, M., Durrani, A. Z., Saleem, M. H., Rehman, M. U., Iqbal, M. K., Wang, Y., Ahmad, H. I., Abbas, T., Hussain, R., Ghori, M. T., Ali, S., Khan, A. U., and Li, J. (2017). A review on epidemiology, global prevalence and economical losses of fasciolosis in ruminants. *Microbial Pathogenesis*, 109, 253–262. <https://doi.org/10.1016/j.micpath.2017.06.006>
- Mia, M. M., Hasan, M., and Chowdhury, M. R. (2021). A systematic review and meta-analysis on prevalence and epidemiological risk factors of zoonotic fascioliasis infection among the ruminants in Bangladesh. *Heliyon*, 7(12), e09732.
- Nascimento, G. G., Locatelli, J., Freitas, P. C., and Silva, G. L. (2000). Antibacterial Activity of Plant Extracts and Phytochemicals on Antibiotic-Resistant Bacteria. *Brazilian Journal of Microbiology*, 31, 247-256.
- Nigussie, D., Davey, G., Tufa, T. B., Brewster, M., Legesse, B. A., Fekadu, A., and Makonnen, E. (2021). Antibacterial and antifungal activities of Ethiopian medicinal plants: A systematic review. *Frontiers in Pharmacology*, 12, 633921. <https://doi.org/10.3389/fphar.2021.633921>
- Nixon, S. A., Welz, C., Woods, D. J., Costa, L. M., Zamanian, M., and Martin, R. J. (2020). Where are all the anthelmintics? Challenges and opportunities on the path to new anthelmintics. *International Journal for Parasitology, Drugs and Drug Resistance*, 14, 8–16. <https://doi.org/10.1016/j.ijpddr.2020.07.001>
- Nurlaelasari, A., Caro, T. M., Nugroho, H. A., Sukaryo, S., Cahyadi, M., Kurniawan, W., and Hamid, P. H. (2023). *Artemisia Vulgaris* Anthelmintic Activities to Ova and Adult Stages of *Fasciola gigantica* in Vitro. *Veterinary World*, 16(5), 1141–1153.

- Ortiz, P., Scarcella, S., Cerna, C., Rosales, C., Cabrera, M., Guzman, M., Lamenza, P. And Solana, H. (2013) Resistance of *Fasciola Hepatica* against Triclabendazole in Cattle in Cajamarca (Peru): A Clinical Trial and an *In Vivo* Efficacy Test In Sheep. *Veterinary Parasitology*, 195(1–2), 118–121.
- Oyebode, O., Kandala, N. B., Chilton, P. J., and Lilford, R. J. (2016). Use of traditional medicine in middle-income countries: a WHO-SAGE study. *Health Policy and Planning*, 31(8), 984-991.
- Payne, S. E., Kotze, A. C., Durmic, Z., and Vercoe, P. E. (2013). Australian Plants Show Anthelmintic Activity toward Equine Cyathostomins in Vitro. *Veterinary Parasitology*, 196(1-2), 153-160.
- Pessoa, L. M., Morais, S. M., Bevilaqua, C. M. L., and Luciano, J. H. S. (2002). Anthelmintic Activity of Essential Oil of *Ocimum Gratissimum* Linn. And *Eugenol* against *Haemonchus Contortus*. *Veterinary Parasitology*, 109(1-2), 59-63.
- Rizwan, M., Khan, M. R., Afzal, M. S., Manahil, H., Yasmeen, S., Jabbar, M., and Cao, J. (2022). Prevalence of Fascioliasis in Livestock and Humans in Pakistan: A Systematic Review and Meta-Analysis. *Tropical Medicine and Infectious Disease*, 7(7), 126.
- Rodriguez Vivas, R. I., Grisi, L., De Leon, A. a. P., Villela, H. S., Torres Acosta, J., Sanchez, H. F., Salas, D. R., Rosario-Cruz, R., Saldierna, F., and Carrasco, D. G. (2017). Potential economic impact assessment for cattle parasites in Mexico. Review. *Revista Mexicana De Ciencias Pecuarias*, 8(1), 61–74.
- Romero-Benavides, J. C., Ruano, A., Silva-Rivas, R., Castillo-Veintimilla, P., Vivanco-Jaramillo, S. L., and Bailón-Moscoso, N. (2017). Medicinal plants used as anthelmintics: Ethnomedical, pharmacological, and phytochemical studies. *European Journal of Medicinal Chemistry*, 129, 209–217. <https://doi.org/10.1016/j.ejmech.2017.02.005>
- Sanchez, M., Gonzalez Burgos, E., Iglesias, I., Lozano, R., and Gomez Serranillos, M. P. (2020). Current uses and knowledge of medicinal plants in the Autonomous Community of Madrid (Spain): A descriptive cross-sectional study. *BMC Complementary Medicine and Therapies*, 20(1), 1-13.
- Sargison, N. D., and Scott, P. R. (2011). Diagnosis and Economic Consequences of Triclabendazole Resistance in *Fasciola Hepatica* in a Sheep Flock in South-East Scotland. *Veterinary Record*, 168(6), 159-159.
- Schweizer, G., Braun, U., Deplazes, P., and Torgerson, P. R. (2005). Estimating the financial losses due to bovine fasciolosis in Switzerland. *Veterinary Record*, 157(7), 188-193.
- Toet, H., Piedrafita, D. M., and Spithill, T. W. (2014). Liver fluke vaccines in ruminants: strategies, progress and future opportunities. *International Journal for Parasitology*, 44(12), 915-927.
- Ullah, R., Rehman, A., Zafeer, M. F., Rehman, L., Khan, Y. A., Khan, M. A., Khan, S. N., Khan, A., and Abidi, S. (2017). Anthelmintic Potential of Thymoquinone and Curcumin on *Fasciola gigantica*. *PLoS One*, 12(2), e0171267. <https://doi.org/10.1371/journal.pone.0171267>
- Urquhart, G. M., Armour, J., Duncan, J. L., Dunn, A. M., and Jennings, F. W. (1996). *Veterinary Parasitology*, 2nd Ed., Blackwell Science, London, pp: 242-251.
- Wulandari, A. R., Nurlaelasari, A., Nugroho, H. A., Cahyadi, M., Kurniawan, W., and Hamid, P. H. (2023). Ethanolic extract of *Etlingera elatior* flower exhibits anthelmintic properties to *Fasciola gigantica* in vitro. *Open Veterinary Journal*, 13(5), 576.
- Zirintunda, G., Biryomumaisho, S., Kasozi, K. I., Batiha, G. E., Kateregga, J., Vudriko, P., Nalule, S., Olila, D., Kajoba, M., Matama, K., Kwizera, M. R., Ghoneim, M. M., Abdelhamid, M., Zaghlool, S. S., Alshehri, S., Abdelgawad, M. A., and Acai-Okwee, J. (2022). Emerging anthelmintic resistance in poultry: Can ethnopharmacological approaches offer a solution? *Frontiers in Pharmacology*, 12. <https://doi.org/10.3389/fphar.2021.774896>