Chapter 51

Herbal Remedies to Replace Tilmicosin in Poultry

Shamreza Aziz^{1*}, Farooq Hussain², Muhammad Zishan Ahmad³, Muhammad Mobashar⁴, Qamar un Nisa⁵, Nadia Nazish⁶, Minahal Fatima⁷ and Muhammad Ifham Naeem¹

ABSTRACT

Poultry has been used as a protein source in many countries around the world as an alternative to beef and mutton. The spread of chronic respiratory diseases results in a great loss of poultry production. *Mycoplasma gallisepticum* is one of the major microbial agents causing chronic respiratory infections in poultry. Tilmicosin is widely used to treat chronic respiratory infections in poultry and after five to seven days of regular treatment the bird gets better. The use of antibiotics against the *Mycoplasma gallisepticum* gives very satisfying results but the long term use of these antibiotics produce resistance in the microbial agent against these antibiotics. This situation comes up with a new solution which is the use of herbal medicine to treat poultry flocks. The concept of using herbal products against diseases is not new, in China many herbal products have been used to treat many medical conditions for almost the last 4000 years. Many herbal products including Andrographolide, Chinese herbal medicine formula, *Lonicera japonica* extract, Meniran Extract, Methanol extracts Garlic, Glycyrrhiza and Neem, and Extract of Indonesian wild ginger are being used to treat chronic respiratory diseases. There is no risk of resistance while using herbal medicine also there are no hidden side effects of them.

KEYWORDS

Chronic respiratory disease, poultry, herbal products, *Mycoplasma gallisepticum*, tilmicosin, Andrographolide, Chinese herbal medicine formula, *Lonicera japonica* extract, Meniran Extract, Methanol extracts Garlic, Glycyrrhiza and Neem, and Extract of Indonesian wild ginger.

Received: 19-Jun-2024 Revised: 03-Jul-2024 Accepted: 10-Aug-2024



A Publication of Unique Scientific Publishers

Cite this Article as: Aziz S, Hussain F, Ahmad MZ, Mobashar M, Nisa QU, Nazish N, Fatima M and Naeem MI, 2024. Herbal remedies to replace tilmicosin in poultry. In: Abbas RZ, Khan AMA, Qamar W, Arshad J and Mehnaz S (eds), Complementary and Alternative Medicine: Botanicals/Homeopathy/Herbal Medicine. Unique Scientific Publishers, Faisalabad, Pakistan, pp: 439-445. https://doi.org/10.47278/book.CAM/2024.400

INTRODUCTION

The huge problem in the treatment and the control of *Mycoplasma gallisepticum* is the development of different phenotypic variations. Also the current situation of the antibiotic resistance of different bacteria makes it difficult to control the disease. Tilmicosin is widely used against the *M. gallisepticum* in poultry for a long time. The long term use of antibiotics in poultry results in the development of resistance in the poultry industry. The use of antibiotics as a growth promoter in poultry is another way to resist (Gharaibeh and Al-Rashdan, 2011). Antibiotic residues in milk, meat and eggs is another side effect of long term use of antibiotics and the aftermath of this is the transfer of antibiotic resistant bacteria to the humans. Other side effects include allergic reactions, immunological and pathological reactions. Vaccination and antibiotics are the only way to treat the chronic respiratory diseases but the resistance of the bacteria against the antibiotics impart the need for another way of treatment. This results in the use of herbal medicine to treat different diseases including *M. gallisepticum* infections. The use of herbal products as a treatment regime is incredibly increasing worldwide (Ishfaq et al., 2021).

Tilmicosin against CRD in Poultry

For a long time, macrolides have been used against different bacterial infections. These antibiotics did their actions by inhibiting the synthesis of bacterial protein. The antibiotic binds to the ribosomal RNA of bacteria and stops the protein

¹KBCMA College of Veterinary and Animal Sciences, Narowal, Sub-campus UVAS-Lahore, Pakistan

²Shaheed Benazir Bhutto University of Veterinary and Animal Sciences, Sakrand, Pakistan

³Department of Veterinary Pathology, PMAS-Arid Agriculture University, Rawalpindi-Subcapmus Khushab, Pakistan

⁴Department of Animal Nutrition, Faculty of Animal Husbandry and Veterinary Sciences, University of Agriculture, Peshawar, Pakistan

⁵Department of Pathology, University of Veterinary and Animal Sciences, Lahore, Pakistan.

⁶Department of Zoology University of Sialkot, Sialkot, Pakistan

⁷Department of Zoology, Wildlife and Fishries, University of Agriculture, Faisalabad, Pakistan

^{*}Corresponding author: shamrezaaziz@gmail.com

synthesis (Vester and Douthwaite, 2001). Tilmicosin is a macrolide which is a broad- spectrum antibiotic. Tilmicosin has a bacteriostatic effect which is synthesized from tylosin only for veterinary use. Tilmicosin is very effectively used against *Mycoplasma* spp. and many Gram-positive organisms (Prescott and Baggot, 1993). In poultry, Tilmicosin is used to treat many chronic respiratory infections caused by *M. gallisepticum* (MG), *Mycoplasma synoviae*, *rhinotracheale*, and *Pasteurela multocida* (Figure 1) (Jordan and Horrocks, 1996; Kempf et al., 1997; Varga et al., 2001).

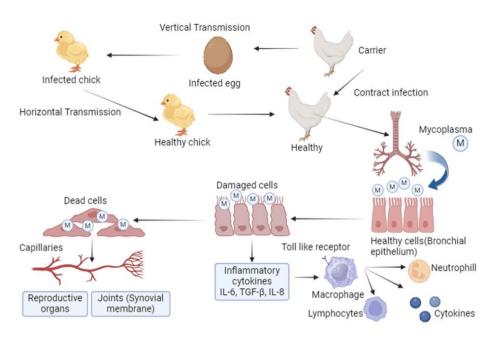


Fig. 1: Transmission of *Mycoplasma gallisepticum* and its pathogenesis (enters in the airways and destroys the cilia of epithelial cells, systemic invasion, causes immunosupression, srops protein and DNA synthesis) in poultry.

Mode of action of Tilmicosin

The classification of antibiotics is based on their mode of action as all types of antibiotics for example penicillin, cephalosporin, tetracycline, macrolides, aminoglycosides, and quinolones have different ways to response against the microbial agents. Tilmicosin belongs to the macrolide, a group of antibiotics which is isolated from a bacteria belonging to genus Streptomyces, present in soil (Riviere and Papich, 2013). Macrolides did their work by inhibiting the biosynthesis of protein in bacteria (Fig. 2). Tilmicosin is basically prepared from desmycosin after chemical alterations (Yazar et al., 2001).

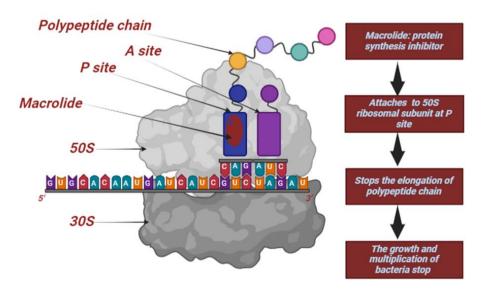


Fig. 2: Mechanism of action of Macrolide (Tilmicosin).

Uses of Tilmicosin in Poultry

The use of tilmicosin in water has very promising effects on chicken against MG including decrease in the formation and degree of lesions on the air sacs (Charleston et al., 1998). The use of tilmicosin at a dose rate of 20mg/kg of body weight for the clinical outbreaks of the MG is thought to be the best way of treatment. This protocol should be followed for five days continuously (Garmyn et al., 2019). The broiler chicken treated with tilmicosin and lincomycin has less or no mean gross air sac and microscopic tracheal lesions compared to those chickens which were untreated. This clears the fact that tilmicosin has very effective results against the mycoplasmosis in chickens (Amer et al., 2009). It is shown by a study that the spiramycin also shows good results against MG in broilers. It reduces the microscopic and gross lesions and also decreases the severity of arthritis. The clinical signs of the disease in treated flock was much lower as compared to those

infected flock which is untreated (Elazab et al., 2021). Many studies reveal that the tilmicosin promisingly reduces microscopic lesions and also decreases the re-isolation rate at 20% and in some other findings the re-isolation rate decreases upto 0% (Abd El-Ghany, 2009; Zakeri and Kashefi, 2011).

Herbal Treatment Options Instead of Tilmicosin in Poultry

The term polypharmacology means the action of an antimicrobial agent at more than one target simultaneously, the latest approach of modern drug discovery (Anighoro et al., 2014; Kuenzi et al., 2017). Through this way natural products are used for treatment (Figure 3) of different diseases which is way better than the same old drugs which are single targeted (Ho et al., 2018). Natural products with the phenomenon of poly-pharmacological profiles defend the host against many diseases with novel therapeutic benefits (Fang et al., 2018; Ishfaq et al., 2021; Wang et al., 2022). The treatment of MG with vaccines is no more effective (Yang et al., 2021). The humoral response of the vaccine against MG is neither as effective nor can clear the pathogen which in turn results in the failure of the vaccine. The genetic variation of MGis also the cause of vaccine failure (Matyushkina et al., 2016). Antibiotics or antimicrobial agents can stop the microbial agent but only for a small period of time. As soon as the drug delivery stops the recurrence of infection is confirmed (Gharaibeh and Al-Rashdan, 2011). These circumstances lead to the need of another effective way of treatment which is not only cheap but also safe to use. The herbal medication is the safest way for treatment of many infections including mycoplasmosis.

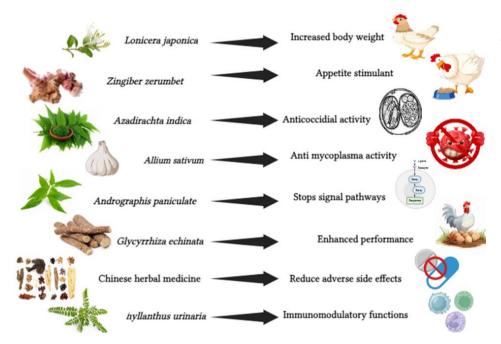


Fig. 3: Herbal treatment options in poultry.

Andrographolide

The main etiological agent for the chronic respiratory disease in chicken is MG The primary symptoms of the chronic respiratory disease is the inflammation of the respiratory tract and apoptosis. Andrographolide (Andro) is a small natural compound. It is famous for its natural properties including Anti-inflammatory action and anti-pathogenic effect (Luo et al., 2022). Andrographis paniculate (A. paniculate) has been used in China, India, Japan, Korea, and some other Asian countries for the last 2000 years as a medicinal food. It is known to treat many diseased conditions in humans for instance myocardial ischemia, pharyngitis, and respiratory diseases (Dai et al., 2019). Andro is a small natural molecule which was extracted from Traditional Chinese Medicine which was made up from A. paniculate (Kim et al., 2019). The natural molecule of Andro works by inhibiting the protein expression of MG which in turn decreases the adhesion of MG to the chicken lungs and AEC II. It has been proved through many studies that MG causes inflammation of the air sac, tracheal tissue and lungs. It also causes damage to the chickens (Wu et al., 2019; Niu et al., 2020). It is confirmed that the administration of Andro is safe in chickens and AEC II and have no adverse side effects. Andro is considered a very effective herbal product against MG as it stops the signal pathways of JAK/PI3K/AKT in the chicken.

Chinese Herbal Medicine Formulae

Recently Chinese herbal medicines have gained scientists' interest with their little toxicity and side effects. Beyond these effects there is no drug resistance and no drug residuals associated with the Chinese herbal medicine (Maryam et al., 2020). In China the use of Chinese herbal medicine to deal with different diseases is almost 4000 years old (Yi and Chang, 2004; Xutian et al., 2009). Old data of use of herbal medicine shows that for the treatment of chronic respiratory disease, a single Chinese herb has very prominent effects (Niu et al., 2020; Zou et al., 2020; Zou et al., 2021). Besides this the Chinese herbal formulae is made by using traditional Chinese medicine formula principles which helps to reduce the adverse side effects and helps in getting better therapeutic benefits (Su et al., 2016). This use of this formula to treat chicken has no

deleterious effects on the health of normal chicken but it can enhance the performance of the chicken which is infected by MG. It also repaired the damage caused by MG to the air sac and tracheal tissue of the chicken. The histopathological data shows that the Chinese herbal formulae also helps in repairing the inflammation of lung and trachea and inhibits the expression of inflammatory cytokines induced by MG (Wang et al., 2022).

Lonicera japonica Extract

Herbs and spices have amazing properties which include antioxidant, antimicrobial, anti-helmintic, immune modulator and growth promoting abilities. These properties belong to the biologically active components of these herbs for instance terpenoids, phenolics, glycosides and alkaloids (Huyghebaert et al., 2011). Lonicera japonica Thunb is a very famous medical plant also known as the Japanese honeysuckle. It has many pharmacological and broad spectrum biological properties which includes anti-oxidative (Kim et al., 1994), anti-tumour (Yip et al., 2006), hepato-protective (Hu et al., 2008), antiviral (Houghton et al., 1993), anti-inflammatory (Lee et al., 1998), and antibacterial activities (Shan et al., 2007; Rahman and Kang, 2009; Shang et al., 2011). The primary natural extracts of L. japonica which act as natural medicine against many diseases are essential oils, organic acids, flavones, saponins, iridoids, and inorganic elements. The most important compound of L. japonica is the chlorogenic acid which is a bioactive organic acid. The chlorogenic acid is found in rich amounts in the flower bud of L. japonica. It has various pharmacological effects against many diseases. Many research studies show that the pharmacological effects of L. japonica extracts have amazing results in the chickens infected with MG. The results of use of L. japonica against MG is as good as achieved by tylosin tartrate. It reduces the chance of body weight loss in chickens infected with MG. Also in the FCR of birds shows that the use of plan based medicinal products have almost the same outcomes as the chemotherapeutic drugs (Giannenas et al., 2003; Saini et al., 2003). To get the best results, chlorogenic acid was given in drinking water to the poultry flock infected with MG at a dose rate of 190ug per day per bird. The liquid chlorogenic acid is prepared at a rate of 1g/ 1000 ml of water which is then used in drinking water for poultry flock. Various studies show that the use of chlorogenic acid results in increased body weight, less individuals with the infection of MG (Müştak et al., 2015).

Meniran (Phyllanthus urinaria) Extract

Currently the use of herbal medicine is widely spreading around the globe specially to treat the disease which affects the production of livestock. Among many of the herbal products used for treatment of animal diseases, Meniran plats are specifically used for the treatment and prevention of chronic respiratory disease. Meniran (Phyllanthus niruri Linn) has been known for the possession of bioactive compounds which have antimicrobial activities. These bioactive compounds are terpenoids, alkaloids, flavonoids, saponins, and tannins (Sreenivas, 1999). The statistical data of various researches shows that the use of Meniran extracts in chicken infected with MG, the body weight increases and the feed conversion ratio (FCR) decreases which shows the excellent performance of the flock (Hidanah et al., 2017). When the chicken is infected with chronic respiratory disease or MG bacteria, it results in a low weight gain and high feed conversion ratio. This happened due to the low appetite which in turn results in less feed consumption by the bird (Bell and Weaver, 2002). Meniran contains chemical compounds which help in enhancing the bird's appetite (Sutton, 2011). Some studies show that Meniran plants also have the benefits of immunomodulatory functions (Gunal et al., 2006). This means that they can help in repairing and restoring the immune system against diseases (Sreenivas, 1999). The natural compounds of Merinan extracts including alkaloids and tannins are well known for their inhibitory response against the microbial agent (Wahju, 2004). The Merinan plant also has the active compounds of phenol class such as flavonoids, alkaloids, saponins, and tannins. Among all the compounds, flavonoids are the one with the ability to denature proteins. With the ability, flavonoids destroy the cell wall proteins of the bacteria which changes its permeability (North and Bell, 1990).

Methanol Extracts Garlic, Glycyrrhiza and Neem

By using the micro dilution method, the minimum inhibitory concentrations (MICs) was determined for the methanol extracts. Herbal plants which include garlic, Glycyrrhiza and Neem were used for the methanol extracts against the three isolates of MG. The procedure was done at the University Diagnostic Lab, University of veterinary and animal sciences, Lahore. The protocols given by (Al-Momani et al., 2007) were used to do this procedure only with minor changes. The results of the study by (Muhammad et al., 2015) reveals that garlic has no adverse side effects on the two isolates of MG. These results were very close to the (Al-Momani et al., 2007) data according to which *Allium sativum* (garlic) shows anti mycoplasma activity against almost six different Mycoplasma species. This shows that garlic can be used in poultry for its amazing therapeutic activities against many gram positive and gram negative bacteria especially against MG (Bakri and Douglas, 2005). Studies show that the aqueous garlic extracts have astonishing therapeutic activities and protect the host from various gram positive and gram negative bacteria (Iwalokun et al., 2004).

Extract of Indonesian wild Ginger

In Indonesia, three species of wild ginger are very popular which are as follows: Zingiber zerumbet (L.) Smith with local name lempuyang gajah, Zingiber amaricans BL. with local name lempuyang pahit and the Zingiber aromaticum (Vahl.) with local name lampuyang wangi. The size of every part of the plant is different from one another in all three species. Zingiber amaricans is the smallest among all of them. The rhizome of Zingiber zerumbet L. Sm. is well known for its high efficiency as

an appetite stimulant, its rhizome is much bigger in size with yellow flesh. The rhizome of *Zingiber amaricans* BL. is also used to enhance appetite but it is smaller in size with bitter taste. In the case of *Zingiber aromaticum*, the rhizome is fragmented and used for its effective results as a slimming agent (Wahyuni et al., 2013). The phytochemical studies about *Zingiber zerumbet* are still ongoing and reveal that the plant is of much Importance. Different compounds were isolated from the plant including sesquiterpenes (Kader et al., 2010; Yob et al., 2011), terpenoid, flavonoids (Nag et al., 2013), tannins (Prakash et al., 2011), and some aromatic compounds. Volatile oil from the rhizome of *Zingiber zerumbet* have different chemical compounds for example zerumbone, humulene, camprene, a-caryophyllene and champhene (Dai et al., 2013).

Conclusion

The resistance of microbial agents against antibiotics is one of the major issues faced by the practitioners and farmers all over the world. This alarming situation draws the attention of pharmacists to find an alternative and better way to fight against the bacteria. This ends with the discovery of herbal products which have been used in many Asian countries for the last 4000 years. Herbal products have many beneficial effects with almost no adverse side effects. The use of herbal products in poultry gives wonderful results and the production increases. Poultry is a source of protein used in many countries but the antimicrobial resistance causes a serious economic loss to poultry industries. The chronic respiratory diseases result in weight loss and mortality of the flock. The herbal products and treatment methods used to fight against the MG and the results are very satisfied.

REFERENCES

- Abd El-Ghany, W. A. (2009). The in vitro and in vivo evaluation of tiamulin and tilmicosin for the treatment of Mycoplasma gallisepticum infected broiler chickens. *International Journal of Poultry Science*, 8(12), 1189-1198. https://doi.org/10.3923/ijps.2009.1189.1198
- Al-Momani, W., Abu-Basha, E., Janakat, S., Nicholas, R., and Ayling, R. (2007). In vitro antimycoplasmal activity of six Jordanian medicinal plants against three Mycoplasma species. *Tropical Animal Health and Production*, 39, 515-519. https://doi.org/10.1007/s11250-007-9033-1
- Amer, M., Hanafei, A., El-Bayomi, K., and Zohair, G. (2009). Comparative study on the efficacy of some antiMycoplasma drugs on the performance of commercial broiler flocks from infected breeders. *Global Veterinaria*, *3*(2), 69-74.
- Anighoro, A., Bajorath, J., and Rastelli, G. (2014). Polypharmacology: challenges and opportunities in drug discovery: miniperspective. *Journal of Medicinal Chemistry*, *57*(19), 7874-7887. https://doi.org/https://doi.org/10.1021/jm5006463
- Bakri, I., and Douglas, C. (2005). Inhibitory effect of garlic extract on oral bacteria. *Archives of Oral Biology*, *50*(7), 645-651. https://doi.org/https://doi.org/10.1016/j.archoralbio.2004.12.002
- Bell, D. D., and Weaver, W. D. J. (2002). Selected References and Suggested Readings. In *Commercial Chicken Meat and Egg Production* (pp. 1241-1265). Springer.
- Charleston, B., Gate, J., Aitken, I., and Reeve-Johnson, L. (1998). Assessment of the efficacy of tilmicosin as a treatment for Mycoplasma gallisepticum infections in chickens. *Avian Pathology*, *27*(2), 190-195. https://doi.org/10.1080/03079459808419322
- Dai, D. N., Thang, T. D., Chau, L. T., and Ogunwande, I. A. (2013). Chemical constituents of the root essential oils of Zingiber rubens Roxb., and Zingiber zerumbet (L.) Smith. *American Journal of Plant Sciences*. https://doi.org/10.4236/ajps.2013.41002
- Dai, Y., Chen, S.-R., Chai, L., Zhao, J., Wang, Y., and Wang, Y. (2019). Overview of pharmacological activities of Andrographis paniculata and its major compound andrographolide. *Critical Reviews in Food Science and Nutrition*, 59(sup1), S17-S29. https://doi.org/10.1080/10408398.2018.1501657
- Elazab, S. T., Elshater, N. S., Hashem, Y. H., Al-Atfeehy, N. M., Lee, E.-B., Park, S.-C., and Hsu, W. H. (2021). Pharmacokinetic/pharmacodynamic modeling of spiramycin against Mycoplasma synoviae in Chickens. *Pathogens*, *10*(10), 1238. https://doi.org/10.3390/pathogens10101238
- Fang, J., Liu, C., Wang, Q., Lin, P., and Cheng, F. (2018). In silico polypharmacology of natural products. *Briefings in Bioinformatics*, 19(6), 1153-1171. https://doi.org/https://doi.org/10.1093/bib/bbx045
- Garmyn, A., Vereecken, M., De Gussem, K., Depondt, W., Haesebrouck, F., and Martel, A. (2019). Efficacy of tylosin and tilmicosin against experimental Mycoplasma gallisepticum infection in chickens. *Avian Diseases*, 63(2), 359-365. https://doi.org/10.1637/11991-110818-Reg.1
- Gharaibeh, S., and Al-Rashdan, M. (2011). Change in antimicrobial susceptibility of Mycoplasma gallisepticum field isolates. *Veterinary Microbiology*, *150*(3-4), 379-383. https://doi.org/https://doi.org/10.1016/j.vetmic.2011.02.005
- Giannenas, I., Florou-Paneri, P., Papazahariadou, M., Christaki, E., Botsoglou, N., and Spais, A. (2003). Effect of dietary supplementation with oregano essential oil on performance of broilers after experimental infection with Eimeria tenella. *Archives of Animal Nutrition*, *57*(2), 99-106. https://doi.org/https://doi.org/10.1080/0003942031000107299
- Gunal, M., Yayli, G., Kaya, O., Karahan, N., and Sulak, O. (2006). The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broilers. *International Journal of Poultry Science*, 5(2), 149-155.
- Hidanah, S., Sabdoningrum, E. K., Wahjuni, R. S., and Arimbi, A. (2017). Implementation of Meniran Extract (Phyllanthus

- Niruri Linn) on the Performance of Broiler Chickens Infected by Mycoplasma gallisepticum Caused Chronic Respiratory Disease. KNE Life Sciences, 296-307. https://doi.org/https://doi.org/10.18502/kls.v3i6.1138
- Ho, T. T., Tran, Q. T., and Chai, C. L. (2018). The polypharmacology of natural products. *Future Medicinal Chemistry*, 10(11), 1361-1368. https://doi.org/https://doi.org/10.4155/fmc-2017-0294
- Houghton, P. J., Boxu, Z., and Xisheng, Z. (1993). A clinical evaluation of the chinese herbal mixture 'Aden-I'for treating respiratory infections. *Phytotherapy Research*, 7(5), 384-386. https://doi.org/10.1002/ptr.2650070513
- Hu, C., Jiang, H., Liu, H., Li, R., and Li, J. (2008). Effects of LJTF on immunological liver injury in mice. *Anhui Medical Pharmaceutical Journal*, *12*, 295-296.
- Huyghebaert, G., Ducatelle, R., and Van Immerseel, F. (2011). An update on alternatives to antimicrobial growth promoters for broilers. *The Veterinary Journal*, 187(2), 182-188. https://doi.org/10.1016/j.tvjl.2010.03.003
- Ishfaq, M., Zhang, W., Liu, Y., Wang, J., Wu, Z., Shah, S. W., and Li, J. (2021). Baicalin attenuated Mycoplasma gallisepticum-induced immune impairment in chicken bursa of fabricius through modulation of autophagy and inhibited inflammation and apoptosis. *Infection*, 101(3), 880-890. https://doi.org/https://doi.org/10.1002/jsfa.10695
- Iwalokun, B., Ogunledun, A., Ogbolu, D., Bamiro, S., and Jimi-Omojola, J. (2004). In vitro antimicrobial properties of aqueous garlic extract against multidrug-resistant bacteria and Candida species from Nigeria. *Journal of Medicinal Food*, 7(3), 327-333. https://doi.org/https://doi.org/10.1089/jmf.2004.7.327
- Jordan, F., and Horrocks, B. K. (1996). The minimum inhibitory concentration of tilmicosin and tylosin for Mycoplasma gallisepticum and Mycoplasma synoviae and a comparison of their efficacy in the control of Mycoplasma gallisepticum infection in broiler chicks. *Avian Diseases*, 326-334. https://doi.org/https://doi.org/10.2307/1592228
- Kader, M. G., Habib, M. R., Nikkon, F., Yeasmin, T., Rashid, M. A., Rahman, M. M., and Gibbons, S. (2010). Zederone from the rhizomes of Zingiber zerumbet and its anti-staphylococcal activity. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*, 9(1), 63-68.
- Kempf, I., Reeve-Johnson, L., Gesbert, F., and Guittet, M. (1997). Efficacy of tilmicosin in the control of experimental Mycoplasma gallisepticum infection in chickens. *Avian Diseases*, 802-807. https://doi.org/10.2307/1592332
- Kim, N., Lertnimitphun, P., Jiang, Y., Tan, H., Zhou, H., Lu, Y., and Xu, H. (2019). Andrographolide inhibits inflammatory responses in LPS-stimulated macrophages and murine acute colitis through activating AMPK. *Biochemical Pharmacology*, 170, 113646. https://doi.org/https://doi.org/https://doi.org/10.1016/j.bcp.2019.113646
- Kim, S., Kim, J., Kim, S., Oh, M., and Jung, M. (1994). Antioxidant activities of selected oriental herb extracts. *Journal of the American Oil Chemists' Society*, *71*, 633-640. https://doi.org/10.1007/BF02540592
- Kuenzi, B. M., Remsing Rix, L. L., Stewart, P. A., Fang, B., Kinose, F., Bryant, A. T., and Rix, U. (2017). Polypharmacology-based ceritinib repurposing using integrated functional proteomics. *Nature Chemical Biology*, 13(12), 1222-1231. https://doi.org/10.1038/nchembio.2489
- Lee, S., Son, K., Chang, H., Kang, S., and Kim, H. (1998). Antiinflammatory activity of Lonicera japonica. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 12(6), 445-447. https://doi.org/https://doi.org/10.1002/(SICI)1099-1573(199809)12:6%3C445::AID-PTR317%3E3.0.CO;2-5
- Luo, R., Wang, Y., Guo, Q., Fan, C., Jiang, G., Wang, L., and Peng, X. J. I. R. (2022). Andrographolide attenuates Mycoplasma gallisepticum-induced inflammation and apoptosis by the JAK/PI3K/AKT signal pathway in the chicken lungs and primary alveolar type II epithelial cells. *International Immunopharmacology and Inflammation Research*, 109, 108819. https://doi.org/10.1016/j.intimp.2022.108819
- Maryam, M., Te, K. K., Wong, F. C., Chai, T. T., Gary, K., and Gan, S. C. (2020). Antiviral activity of traditional Chinese medicinal plants Dryopteris crassirhizoma and Morus alba against dengue virus. *Journal of Integrative Agriculture*, 19(4), 1085-1096. https://doi.org/10.1016/S2095-3119(19)62820-0
- Matyushkina, D., Pobeguts, O., Butenko, I., Vanyushkina, A., Anikanov, N., Bukato, O., and Semashko, T. (2016). Phase transition of the bacterium upon invasion of a host cell as a mechanism of adaptation: a Mycoplasma gallisepticum model. *Scientific Reports*, 6(1), 35959. https://doi.org/10.1038/srep35959
- Muhammad, J., Anjum, A., Rabbani, M., Muhammad, K., Wasim, M., Ahmad, A., and Liaqat, F. (2015). In vitro efficacy of garlic, glycyrrhiza and neem against local isolates of Mycoplasma gallisepticum. *The Journal of Animal and Plant Sciences*.
- Müştak, H., Torun, E., Özen, D., Yücel, G., Akan, M., and Diker, K. (2015). Effect of Lonicera japonica extract on Mycoplasma gallisepticum in naturally infected broiler flocks. *British Poultry Science*, *56*(3), 299-303. https://doi.org/10.1080/00071668.2015.1022711
- Nag, A., Bandyopadhyay, M., and Mukherjee, A. J. J. O. P. (2013). Antioxidant activities and cytotoxicity of Zingiber zerumbet (L.) Smith rhizome. *Journal of Pharmacognosy and Phytochemistry*, 2(3), 102-108.
- Niu, L., Luo, R., Zou, M., Sun, Y., Fu, Y., Wang, Y., and Peng, X. J. I. I. (2020). Puerarin inhibits Mycoplasma gallisepticum (MG-HS)-induced inflammation and apoptosis via suppressing the TLR6/MyD88/NF-κB signal pathway in chicken. *International Immunopharmacology, 88*, 106993. https://doi.org/https://doi.org/10.1016/j.intimp.2020.106993
- North, M., and Bell, D. (1990). *Commercial chicken production manual* https://www.cabidigitallibrary.org/doi/full/10.5555/19920193871

- Prakash, R. O., Kumar, R. K., Rabinarayan, A., and Kumar, M. S. (2011). Pharmacognostical and phytochemical studies of Zingiber zerumbet (I.) Sm. Rhizome. *International Journal of Research in Ayurveda Pharmacy*, *2*(3), 698-703.
- Prescott, J. F., and Baggot, J. D. (1993). *Antimicrobial therapy in veterinary medicine* (Steeve Giguère, John F. Prescott, and P. Dowling, Eds. 5 ed., Vol. 12). Iowa State University Press.
- Rahman, A., and Kang, S. C. (2009). In vitro control of food-borne and food spoilage bacteria by essential oil and ethanol extracts of Lonicera japonica Thunb. *Food Chemistry*, *116*(3), 670-675. https://doi.org/10.1016/j.foodchem.2009.03.014
- Riviere, J. E., and Papich, M. G. (2013). *Veterinary pharmacology and therapeutics* (Jim E. Riviere and M. G. Papich, Eds. 9 ed.). John Wiley and Sons.
- Saini, R., Davis, S., and Dudley-Cash, W. (2003). Oregano essential oil reduces the expression of coccidiosis in broilers. Proceeding of the 52nd Conference on Western Poultry Diseases, Sacramento, CA,
- Shan, B., Cai, Y.-Z., Brooks, J. D., and Corke, H. (2007). The in vitro antibacterial activity of dietary spice and medicinal herb extracts. *International Journal of Food Microbiology*, 117(1), 112-119. https://doi.org/10.1016/j.ijfoodmicro.2007.03.003
- Shang, X., Pan, H., Li, M., Miao, X., and Ding, H. (2011). Lonicera japonica Thunb.: ethnopharmacology, phytochemistry and pharmacology of an important traditional Chinese medicine. *Journal of Ethnopharmacology*, 138(1), 1-21. https://doi.org/10.1016/j.jep.2011.08.016
- Sreenivas, P. (1999). Herbal healing. Far Eastern Agriculture, 31-32.
- Su, X., Yao, Z., Li, S., and Sun, H. (2016). Synergism of Chinese herbal medicine: illustrated by danshen compound. *Evidence-based Complementary Alternative Medicine*, 2016. https://doi.org/https://doi.org/10.1155/2016/7279361
- Sutton, S. (2011). Measurement of microbial cells by optical density. Journal of Validation Technology, 17(1), 46-49.
- Varga, J., Fodor, L., and Makrai, L. (2001). Characterisation of some Ornithobacterium rhinotracheale strains and examination of their transmission via eggs. *Acta Veterinaria Hungarica*, 49(2), 125-130. https://doi.org/10.1556/004.49.2001.2.1
- Vester, B., and Douthwaite, S. (2001). Macrolide resistance conferred by base substitutions in 23S rRNA. *Antimicrobial Agents Chemotherapy*, 45(1), 1-12. https://doi.org/10.1128/aac.45.1.1-12.2001
- Wahju, J. (2004). Poultry Nutrition Science (1 ed., Vol. 1). UGM Press.
- Wahyuni, S., Bermawie, N., and Kristina, N. N. (2013). Karakteristik morfologi, potensi produksi dan komponen utama rimpang sembilan nomor lempuyang wangi. *Industrial Crops Research Journal*, 19(3), 99-107. https://doi.org/10.21082/jlittri.v19n3.2013.99-107
- Wang, Y.-J., Liang, Y.-X., Hu, F.-l., Sun, Y.-F., Zou, M.-Y., Luo, R.-l., and Peng, X.-l. (2022). Chinese herbal formulae defend against Mycoplasma gallisepticum infection. *Journal of Integrative Agriculture*, 21(10), 3026-3036. https://doi.org/10.1016/j.jia.2022.07.038
- Wu, Z., Chen, C., Miao, Y., Liu, Y., Zhang, Q., Li, R., and Li, J. (2019). Baicalin attenuates mycoplasma gallisepticum-induced inflammation via inhibition of the TLR2-NF-κB pathway in chicken and DF-1 Cells. *Infection Drug Resistance*, 3911-3923. https://doi.org/https://doi.org/10.2147/IDR.S231908
- Xutian, S., Zhang, J., and Louise, W. (2009). *New exploration and understanding of traditional Chinese medicine* (Stevenson Xutian, Dongyi Cao, and J. Junion, Eds. Vol. 37). The American Journal of Chinese Medicine.
- Yang, Y., Wang, Y., Zou, M., Deng, G., and Peng, X. (2021). gga-miR-142-3p negatively regulates Mycoplasma gallisepticum (HS strain)-induced inflammatory cytokine production via the NF-kB and MAPK signaling by targeting TAB2. *Journal of Inflammation Research*, 70, 1217-1231. https://doi.org/https://doi.org/10.1007/s00011-021-01499-2
- Yazar, E., Altunok, V., Elmas, M., Traş, B., Baş, A., and Özdemir, V. (2001). Effect of tilmicosin on cardiac muscle and serum creatine kinases activities and serum total protein level in healthy male Balb/C mice. Revue de Médecine Vétérinaire.
- Yi, Y.-D., and Chang, I.-M. (2004). An overview of traditional Chinese herbal formulae and a proposal of a new code system for expressing the formula titles. *Evidence-Based Complementary and Alternative Medicine*, 1, 125-132. https://doi.org/10.1093/ecam/neh019
- Yip, E., Chan, A., Pang, H., Tam, Y., and Wong, Y. H. (2006). Protocatechuic acid induces cell death in HepG2 hepatocellular carcinoma cells through a c-Jun N-terminal kinase-dependent mechanism. *Cell Biology and Toxicology*, *22*, 293-302. https://doi.org/http://dx.doi.org/10.1007/s10565-006-9000-z
- Yob, N., Jofrry, S. M., Affandi, M., Teh, L., Salleh, M., Zakaria, Z. J. E.-B. C., and Medicine, A. (2011). Zingiber zerumbet (L.) Smith: a review of its ethnomedicinal, chemical, and pharmacological uses. *Evidence-Based Complementary and Alternative Medicine*, 2011. https://doi.org/https://doi.org/10.1155/2011/543216
- Zakeri, A., and Kashefi, P. (2011). Comparative therapeutic efficacy of tiamulin and pulmotil in infected broiler and layer flocks with Mycoplasma gallisepticum. *African Journal of Pharmacy and Pharmacology*, 5(15), 1778-1781.
- Zou, M., Yang, L., Niu, L., Zhao, Y., Sun, Y., Fu, Y., and Peng, X. (2021). Baicalin ameliorates Mycoplasma gallisepticum-induced lung inflammation in chicken by inhibiting TLR6-mediated NF-κB signalling. *British Poultry Science*, *62*(2), 199-210. https://doi.org/https://doi.org/10.1080/00071668.2020.1847251
- Zou, M., Yang, W., Niu, L., Sun, Y., Luo, R., Wang, Y., and Peng, X. (2020). Polydatin attenuates Mycoplasma gallisepticum (HS strain)-induced inflammation injury via inhibiting the TLR6/MyD88/NF-κB pathway. *Microbial Pathogenesis*, *149*, 104552. https://doi.org/https://doi.org/10.1016/j.micpath.2020.104552